Acute Ethanol Exposure Elevates Muscarinic Tone in the Septohippocampal System

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Ericson M, Sama MA, Yeh HH. Acute ethanol exposure elevates muscarinic tone in the septohippocampal system. J Neurophysiol 103: 290–296, 2010. First published November 11, 2009; doi:10.1152/jn.91072.2008. The septohippocampal system has been implicated in the cognitive deficits associated with ethanol consumption, but the cellular basis of ethanol action awaits full elucidation. In the medial septum/diagonal band of Broca (MS/DB), a muscarinic tone, reflective of firing activity of resident cholinergic neurons, regulates that of their noncholinergic, putatively GABAergic, counterparts. Here we tested the hypothesis that ethanol alters this muscarinic tone. The spontaneous firing activity of cholinergic and noncholinergic MS/DB neurons were monitored in acute MS/DB slices from C57Bl/6 mice. Exposing the entire slice to ethanol increased firing in both cholinergic and noncholinergic neurons. However, applying ethanol focally to individual MS/DB neurons increased firing only in cholinergic neurons. The differential outcome suggested different mechanisms of ethanol action on cholinergic and noncholinergic neurons. Indeed, with bath-perfused ethanol, the muscarinic antagonist methyl scopolamine prevented the increase in firing in noncholinergic, but not cholinergic, MS/DB neurons. Thus, the effect on noncholinergic neuronal firing was secondary to ethanol’s direct action of acutely increasing muscarinic tone. We propose that the acute ethanol-induced elevation of muscarinic tone in the MS/DB contributes to the altered net flow of neuronal activity in the septohippocampal system that underlies compromised cognitive function.

INTRODUCTION

Ethanol (alcohol) is one of the most widely used and abused of psychoactive substances, exerting wide-ranging yet remarkably specific effects on myriad targets in the CNS (Crews et al. 1996; Valenzuela 1997). The septohippocampal system, connecting the medial septum/diagonal band of Broca (MS/DB) with the hippocampus via the fimbria-fornix, is one such prominent target. Hippocampus-dependent cognitive functions are particularly sensitive and vulnerable to the effects of ethanol consumption (Matthews and Silvers 2004; White et al. 2000). Thus, in humans and in animal models, both acute ethanol intoxication and chronic ethanol consumption impair hippocampus-dependent spatial learning and memory (Melia et al. 1996; Stokes et al. 1991; White et al. 1998). At the cellular level, the literature is replete with reports of ethanol exerting an impressive host of effects on neuronal excitability. In the hippocampus, ethanol alters not only the functional properties of resident neurons but also the activity of afferent inputs, notably the septohippocampal system, that contribute to orchestrating synchronized synaptic operations within its circuitry (Givens and McMahon 1995; Hendricson et al. 2002; Lima-Landman and Albuquerque 1989; Simson et al. 1993; Weiner et al. 1997).

The MS/DB has long been implicated in mnemonic functions (Dutar et al. 1995; Givens and Olton 1994; Olton et al. 1978; Pouzet and Herrmann 1990). Memory impairments have been reported following experimental manipulations of the MS/DB and fimbria-fornix (Druchman and Sahakian 1979; Hepler et al. 1985; Kesner et al. 1986; Meck et al. 1987; Warburton 1972). Within the MS/DB, coordinated firing activity of cholinergic and noncholinergic, presumably GABAergic, neurons projecting to the hippocampus (Köhler et al. 1984; Lewis et al. 1967; Rye et al. 1984) underlies the generation of hippocampal theta rhythms and contribute to sustaining hippocampus-dependent forms of learning and memory (Colom 2006; Hasselmo 2005; O’Keefe 1993; Vertes and Kocsis 1997). There are in addition smaller contingents of peptidergic and glutamatergic projection neurons the functional roles of which remain to be elucidated (Colom et al. 2005; Gritti et al. 2006; Peterson and Shurlow 1992; Senut et al. 1989; Sotty et al. 2003).

An early series of studies demonstrated that acute administration of ethanol attenuated hippocampal theta rhythm and long-term potentiation, suppressed spontaneous firing and enhanced GABA-mediated inhibition in MS/DB neurons (Givens 1995; Givens and Breese 1990a,b; Givens and McMahon 1995). However, the acute effects of ethanol on identified neurons and circuit operations within the MS/DB remain outstanding. We and others have shown in rodents that firing rate is a reliable index for discriminating cholinergic versus noncholinergic MS/DB neurons and that the firing activity of the latter neuronal subpopulation is subject to constant regulation by a muscarinic tone that is sustained by the firing of cholinergic neurons (Alreja et al. 2000; Wu and Yeh 2005). Here we asked whether acute exposure to ethanol affected the firing activity of the cholinergic and noncholinergic neuronal subpopulations in the MS/DB and, if so, whether this could be attributed to an altered muscarinic tone.

METHODS

All procedures involving animals were carried out in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals and approved by the Dartmouth Medical School Institutional Animal Care and Use Committee.

This study used brain slices containing the MS/DB derived from postnatal day 10–22 C57Bl/6 mice when slice viability and recording conditions were optimal for stable and prolonged monitoring of spontaneous firing activity in MS/DB neurons. In rat, the septohip-
pocampal system, including the MS/DB, acquires adult features around postnatal day 14 (Bender et al. 1996; Linke and Frotscher 1993, 1995; Milner et al. 1983). In mouse, information on the development of the MS/DB is limited (e.g., Ward and Hagg 1999, 2000). However, because the developmental schedule of the mouse CNS precedes that of the rat by ~2 day (Clancy et al. 2001), the MS/DB in the slices used for our experiments are in a relatively mature state.

**Electrophysiology**

C57Bl/6 mice (postnatal day 10–22) were asphyxiated by carbon dioxide and decapitated. The brain was quickly removed, blocked by removing the cerebellum and hindbrain, and immersed en bloc in ice-cold oxygenated artificial cerebral spinal fluid (ACSF) containing (in mM) 125 NaCl, 2.5 KCl, 2.0 CaCl$_2$, 1.0 MgCl$_2$, 1.25 NaH$_2$PO$_4$, 26 NaHCO$_3$, and 10 glucose. All solutions were bubbled continuously with 5% CO$_2$-95% O$_2$. Coronal brain slices (200–

During electrophysiological recording, the brain slices were continuously perfused at rate of 0.5 ml/min with oxygenated ACSF maintained at 32–33°C by a heated stage fit onto an upright microscope (BXWI50, Olympus, Melville, NY). A CCD camera attached to a video frame grabber board (Integral Technologies, Indianapolis, IN) facilitated real-time visualization of cells under Hoffman Modulation Optics and placement of recording electrodes and drug pipettes (Fig. 1B). Putative cholinergic and noncholinergic MS/DB neurons were identified on-line by firing rate in the whole cell current-clamp mode (Wu and Yeh 2005); cholinergic cells were confirmed off-line by immunohistochemistry (Fig. 1, E–G).

Recording pipettes were filled with internal solution containing (in mM) 140 KCl, 1.8 CaCl$_2$, 1.0 MgCl$_2$, and 5.0 HEPES (pH 7.4), 3.0 Mg$^{2+}$-ATP, and either Lucifer yellow or biocytin (0.1%). Whole cell current and voltage-clamp recordings were conducted using an Axo-Patch 200A amplifier (Axon Instruments, Foster City, CA).

Bioelectric activity was amplified, digitized using Clampfit v8.0 and analyzed with Clampfit v8.0 (Axon Instruments) and Mini Analysis software (Version 6.01, Synaptosoft, Decatur GA). Statistical analysis was performed using SigmaPlot 8.02 (SPSS, Chicago, IL). Data were reported as means ± SE.

Drugs were dissolved in ACSF and either bath-perfused or loaded into a six-barrel multibarrel pipette assembly and were delivered by means of regulated pressure (Picospritzer, General Valve, Fairfield, NJ). The ejection pressure was routinely adjusted to ≤3 psi to avoid mechanical artifacts due to bulk flow. The drug pipette was visually placed 5–10 μm from the cell being recorded. During control and recovery phases of each recording, ACSF solution was continually applied to control for an ethanol-induced effect and to facilitate wash-out of the drug. In experiments involving focal application, the concentration of ethanol (50 mM) and methyl scopolamine (10 μM) indicated throughout this study are those used to fill the drug barrels and represent the maximal limit to which a cell would have been exposed. With constant perfusion of ACSF during electrophysiological recording, the final concentrations are difficult to ascertain but would be expectedly lower than those in the drug barrels due to dilution factors on ejection.

**FIG. 1.** Cholinergic and noncholinergic medial septum/diagonal band of Broca (MS/DB) neurons display different rates of spontaneous firing. A: coronal brain slice (200 μm) from a postnatal day 13 C57Bl/6 mouse with MS/DB outlined. B: a magnified view of the recording setup with a patch recording pipette (right) on the neuron being recorded and a multibarrel pipette assembly (left) used to deliver drugs positioned ~10 μm from the cell. C: example of an MS/DB neuron filled with Lucifer yellow during recording (C1) displaying a slow firing rate (C2). D: example of an MS/DB neuron filled with Lucifer yellow during recording (D1) displaying a fast firing rate (D2). Scale bars apply to both C and D: vertical = 20 mV; horizontal = 1 s. E–G: the slice was fixed after electrophysiological recording and processed immunohistochemically with an antibody against choline acetyltransferase (ChAT). The cells illustrated in C and D were recovered and are denoted (→, *) in E and F, respectively. The overlay in F indicates that the slow-firing cell illustrated in C, but not the fast-firing cell illustrated in D, is ChAT immunopositive.
**RESULTS**

Cells in MS/DB slices were selected for recording, and Lucifer yellow or biocytin filled the cells while spontaneous firing rate was monitored. The recorded slices were then fixed and processed for ChAT immunohistochemistry. We found that expression of ChAT immunoreactivity was limited to the fast-firing (≥3 Hz) subpopulation of cells (Fig. 1). This is consistent with our previous study (Wu and Yeh 2005), which also demonstrated that the fast-firing subpopulation of MS/DB neurons typically are GABAergic because they expressed parvalbumin, calbindin, or calretinin. However, other peptidergic and glutamatergic neuronal subpopulations are present (Colom et al. 2005; Gritti et al. 2006; Peterson and Shultrow 1992; Senny et al. 1989; Sotty et al. 2003). Therefore the present study will refer to the slow-firing MS/DB neurons as cholinergic neurons and the fast-firing ones as “noncholinergic neurons” with the recognition that the majority of them are likely to be either GABAergic or glutamatergic neurons because they appear to be the predominant noncholinergic contingent in the MS/DB (Gritti et al. 2006; Sotty et al. 2003).

**Bath perfusion of ethanol acutely increases firing in both cholinergic and noncholinergic MS/DB neurons**

We first assessed the effect of acute ethanol exposure on the firing rates of cholinergic and noncholinergic MS/DB neurons. The spontaneous firing rates of MS/DB neurons were monitored under whole cell current-clamp conditions, and ethanol was included in the ACSF used to perfuse slices. Because the variability in firing rates precluded a systematic determination of ethanol concentration-response relationship (not shown), we chose to use 50 mM ethanol because this concentration produces acute intoxication and has been shown to affect synaptic extrasynaptic neuronal activities (Jia et al. 2008). Figure 2 illustrates that exposing the entire slice to perfusion solution containing ethanol increased firing in both cholinergic and noncholinergic neurons. In the examples of ratemeter records taken from a cholinergic (Fig. 2A) and a noncholinergic MS/DB neuron (Fig. 2B), the firing rates increased gradually in response to a 5-min exposure to ethanol (solid line above the rate meter records). In both cholinergic and noncholinergic neurons (Fig. 2, A and B, respectively), a consistent observation was that recovery from the ethanol-induced effect was partial, as the increase in spontaneous firing rate persisted well beyond the termination of acute ethanol application and remained elevated over the duration of the recording (≥30 min). While incomplete wash-out of ethanol could not be ruled out entirely, two lines of evidence indicate that the slowly reversible ethanol-induced effect was not due to a continual run-up of firing activity typically encountered in injured neurons. First in control experiments in which spontaneous firing activity was monitored without exposure to ethanol in cholinergic and noncholinergic MS/DB neurons, the firing rate remained stable over the same length of time (Supplemental Fig. S1). Second,
the firing activity stabilized, albeit to an elevated level of baseline firing.

The rates of spontaneous firing before and 5 min after the onset of ethanol perfusion were quantified, and data obtained from 11 cholinergic neurons and 16 noncholinergic neurons are summarized in Fig. 2, A (inset) and B (inset), respectively. As illustrated in the scatter plots (C and D), ethanol exposure increased the rate of spontaneous firing in 18 of 20 cholinergic neurons (Fig. 2C; 65 ± 15%; means ± SE; \( P < 0.01 \), Student’s t-test) and in 11 of 16 noncholinergic neurons (D; 15 ± 1.7%; \( P < 0.01 \), Student’s t-test). Figure 2 also reveals that ethanol increased firing to a greater extent in cholinergic than noncholinergic neurons and the onset of the increased firing differed temporally in the two populations of MS/DB neurons. As shown in Fig. 2C, the onset of ethanol-induced increase in firing was faster in cholinergic than noncholinergic neurons and the onset of the increased firing differed temporally in the two populations of MS/DB neurons. Although the degree to which ethanol increased the firing rate varied, the effect was consistently seen in all such neurons examined (11 of 11 cases). The ethanol-induced increase in firing was slowly reversible, typically returning to baseline (preethanol exposure) levels longer than 30 min after the termination of focal ethanol application (not shown). By contrast, focal exposure to ethanol did not affect firing rate in any of the noncholinergic neurons tested (13 of 13 cases).

**Ethanol increases muscarinic tone in the MS/DB**

While acute exposure to ethanol could increase firing in both the cholinergic and noncholinergic MS/DB neuronal populations, pending the mode of ethanol exposure, our results prompted us to test the hypothesis that ethanol acts directly on cholinergic neurons but indirectly on the noncholinergic neurons. We postulated that the increase in firing seen in noncholinergic neurons following bath perfusion of ethanol is due to increased muscarinic tone brought about by increased firing activity in cholinergic neurons. If so, then the increase in noncholinergic neuronal firing induced by bath perfusion of ethanol should be mediated by muscarinic receptors and thus blocked in the presence of methyl scopolamine. Indeed this was the experimental outcome. As summarized in Fig. 4, focal application of methyl scopolamine (10 \( \mu M \)) did not block the ethanol-induced increase in firing recorded in cholinergic neurons (A) but blocked that in noncholinergic neurons (B).

It should be noted that methyl scopolamine, by itself, tended to increase firing rate in cholinergic MS/DB neurons (Fig. 4A). Although this did not reach statistical significance (\( P = 0.082 \), Student’s t-test), an issue that arose was whether such an increase might alter ethanol’s effect in increasing the firing rate of cholinergic neurons. To address this, the baseline spontaneous firing rates of cholinergic MS/DB neurons (\( n = 8 \)) were compared with those recorded after exposure to methyl scopolamine alone, methyl scopolamine plus ethanol, and ethanol alone. As summarized in Fig. 4A, muscarinic receptor blockade did not affect the magnitude of the ethanol-induced increase in the firing rate of cholinergic neurons. Importantly, Fig. 4B illustrates that exposure to methyl scopolamine alone suppressed baseline firing in noncholinergic neurons and prevented the increased firing activity normally seen in these cells following exposure to ethanol. Importantly, Fig. 4B illustrates that even a 10-min exposure to methyl scopolamine alone suppressed baseline firing. This is consistent with the notion that the muscarinic receptor antagonist effectively blocked the muscarinic tone that normally maintains firing in noncholinergic MS/DB neurons, thus precluding the effect of bath-perfused ethanol.

**Discussion**

This study examined the acute effect of ethanol on the firing activity of neurons in the MS/DB. Monitoring spontaneous firing from cholinergic and noncholinergic MS/DB neurons allowed us to interpret our data in light of an ethanol-induced effect on muscarinic tone. The main conclusions of our study are that 1) the firing of both cholinergic and noncholinergic MS/DB neurons are increased on acute exposure to ethanol. 2) Ethanol elevates muscarinic tone by acting directly on cholinergic neurons to increase their firing. 3) The firing of noncholinergic neurons, on the other hand, is itself unaffected by ethanol but increases in response to the elevated muscarinic tone. The idea that MS/DB neurons are not uniformly sensitive to the effects of ethanol is reminiscent of early in vivo studies by Givens and Breese reporting site- and cell-type-specific effects of acute ethanol administration in the medial and lateral septum (Givens and Breese 1990a,b).
FIG. 5. Model of acute ethanol action on muscarinic tone in the MS/DB by methyl scopolamine (10 μM) with ethanol (50 mM) bath perfused on cholinergic cells (A) and noncholinergic (B) MS/DB neurons. A: comparison of baseline rate of spontaneous firing in cholinergic neurons with firing rates monitored after exposure to methyl scopolamine only, then to methyl scopolamine plus ethanol, and then to ethanol alone. A statistically insignificant trend for methyl scopolamine to increase cholinergic neuronal firing is evident. However, concomitant exposure to methyl scopolamine did not affect the ethanol-induced increase in firing (P = 0.737). B: prevention of the ethanol-induced effect on increasing spontaneous firing in noncholinergic MS/DB neurons by methyl scopolamine (10 μM). Data are derived from a minimum of 8 neurons for each condition and presented as percentage baseline firing, means ± SE, n = 8–14. Statistics: Student’s t-test, * statistically significant differences.

The disposition of cholinergic MS/DB neurons in acute and chronic states of ethanol consumption has commanded intense experimental attention in part because much of the muscarinic receptor-mediated activities implicated in learning and memory have been attributed to the activity of cholinergic neurons in the MS/DB and their efferent projections to the hippocampus. Comparatively little is known about how ethanol affects the noncholinergic, notably the GABAergic, contingent, despite the fact that the latter has also been implicated in playing a significant role in the regulation of the firing activity of noncholinergic neurons.

Acute ethanol exposure (1), in turn, feedback onto the cholinergic neurons to regulate muscarinic tone that regulates the firing activity of noncholinergic neurons (2). The noncholinergic neurons increased firing only when whole slices were perfused and exposed to ethanol, and this effect was blocked by methyl scopolamine. These findings, in toto, pointed to two cell type-specific modes of acute ethanol action: a direct effect on cholinergic neurons and an indirect muscarinic receptor-mediated mechanism in noncholinergic neurons that is manifested through an elevated ambient muscarinic tone.

Acute ethanol exposure (3), in turn, feedback onto the cholinergic neurons to regulate their firing activity (4). Acute ethanol exposure (right) rapidly increases firing in cholinergic neurons (1), which, in turn,
enhances acetylcholine release and elevates muscarinic tone (2). The heightened muscarinic tone increases firing in non-cholinergic neurons (3), notably the GABAAergic and glutamatergic neurons, both of which receive cholinergic innervation (Bialowas and Frotscher 1987; Colom et al. 2005; Van der Zee and Luiten 1994). Within the circuitry of the MS/DB, these findings invoke the following predictions. First the increased firing in noncholinergic excitatory glutamatergic neurons would feed forward to maintain or further elevate firing activity in cholinergic neurons. Second, as depicted in Fig. 5, the increased firing in noncholinergic inhibitory GABAergic neurons would suppress firing activity of cholinergic neurons and muscarinic tone. However, preliminary observations (Yeh, unpublished data) suggest that acute ethanol attenuates GABA_A receptor-mediated responses in the postsynaptic cholinergic neurons (hypothetically depicted in Fig. 5 as decreased number of GABA_A receptors), and this may offset any effect of acute ethanol exposure on facilitating presynaptic GABA release (4). A muted postsynaptic GABA response thus mitigates the normal complement of inhibitory drive onto the cholinergic MS/DB neurons, resulting in a net disinhibition that increases cholinergic neuronal firing.

Indeed a key outstanding issue is how ethanol acutely increases the firing activity in cholinergic MS/DB neurons. Our working model (Fig. 5) favored an involvement of the GABAAergic system because it is particularly sensitive to modulation by physiologically relevant concentrations of ethanol and has been implicated as a key target for many of ethanol’s pre- and postsynaptic actions in the CNS (Grobin et al. 1998; Siggins et al. 2005; Weiner and Valenzuela 2006). In addition, in the MS/DB, cholinergic neurons are innervated by endogenous GABAergic neurons as well as by extra-septal afferents from the hippocampus (Leranth and Frotscher 1989; Töth et al. 1993). Furthermore, ethanol has been reported to modulate GABA-mediated inhibition in the MS/DB (Givens and Breese 1990a,b; Soldo et al. 1994). Clearly, ethanol influences myriad ion channels, neurotransmitter systems and signaling mechanisms in the CNS. Whether and how these and other related factors come into play to confer ethanol’s influence neuronal activity within the operations of the MS/DB circuitry and the septohippocampal system awaits experimental elucidation.

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