Cyclophosphamide-induced cystitis reduces ASIC channel but enhances TRPV1 receptor function in rat bladder sensory neurons

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The prevalence of painful bladder syndrome/interstitial cystitis (PBS/IC) is estimated at ~11% in women and 4.6% in men in managed care settings (Clemens et al. 2005, 2007). In addition to increased urinary frequency, urgency, and incontinence, PBS/IC patients also report significant discomfort and pain to increased urinary frequency, urgency, and incontinence, managed care settings (Clemens et al. 2005, 2007). In addition, bladder and bladder DRG neurons from CYP-treated rats showed signs of inflammation (greater myeloperoxidase activity; lower intramuscular wall pH) and increased size (whole cell capacitance), respectively, compared with controls. Most bladder neurons (>90%) responded to protons and capsaicin. Protons produced multiphasic currents with distinct kinetics, whereas capsaicin always triggered a sustained response. The TRPV1 receptor antagonist A-425619 abolished capsaicin-triggered currents and raised the threshold of heat-activated currents. Prolonged exposure to an acidic environment (pH range: 7.2 to 6.6) inhibited proton-evoked currents, potentiated the capsaicin-evoked current, and reduced the threshold of heat-activated currents in LS and TL bladder neurons. CYP treatment reduced current density but not kinetics of all current components triggered by pH 5. In contrast, CYP-treatment was associated with an increased current density in response to capsaicin in LS and TL bladder neurons. Correspondingly, heat triggered current at a significantly lower temperature in bladder neurons from CYP-treated rats compared with controls. These results reveal that cystitis differentially affects TRPV1- and ASIC-mediated currents in both bladder sensory pathways. Acidification of the bladder wall during inflammation may contribute to changes in nociceptive transmission mediated through the TRPV1 receptor, suggesting a role for TRPV1 in hypersensitivity associated with cystitis.

bladder sensory neurons; ASIC; TRPV1 receptors; visceral hypersensitivity; cystitis; cyclophosphamide; capsaicin; proton; acid; pH; whole cell patch clamp

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Cystitis enhances excitability, increases P2X receptor-mediated currents, and induces expression of P2X3 receptors in bladder sensory neurons (Chen and Gehbart 2010; Dang et al. 2008; Yoshimura and de Groat, 1999). We hypothesized that changes in the sensitivity of bladder sensory neurons to endogenous acid-sensing ion channels (ASIC) and/or TRPV1 receptor agonists contribute to bladder dysfunction, discomfort, and pain. Using a well-established rat model of urinary bladder inflammation (Bon et al. 1997, 2003; Lanteri-Minet et al. 1995), we thus examined the effects of bladder inflammation on ASIC and TRPV1 currents in TL and LS bladder neurons.

MATERIALS AND METHODS

Male Sprague-Dawley rats (200–300 g; Harlan, Indianapolis, IN) were used throughout. Rats were housed under a 12:12-h light-dark cycle with free access to food and water. All experimental protocols were approved by the Animal Care and Use Committee of The University of Iowa.

Bladder Inflammation, Myeloperoxidase Activity, and pH Measurements

Systemic administration of cyclophosphamide (CYP), which is metabolized to the bladder irritant acrolein (Cox 1979), causes hemorrhagic cystitis in humans and produces a cystitis-like condition in rodents (Bon et al. 1997; Dang et al. 2008; Lanteri-Minet et al. 1995). Saline (control) or CYP (100 mg/kg) was administered systemically (intraperitoneally, ip) on days 1, 3, and 5. To assess inflammation, myeloperoxidase (MPO) activity in bladder and distal colon tissue was measured in a subgroup of rats on day 6 (n = 4/group). Rats were euthanized with pentobarbital (200 mg/kg ip), and the bladders and 1 cm of distal colon were removed rapidly, minced, homogenized in ice-cold 50 mM phosphate buffer (pH 6) containing 0.5% hexadecyltrimethylammonium bromide, centrifuged at 1,000 rpm for 5 min, and the supernatant retained. In the presence of hydrogen peroxide (30%) and o-diaminobenzidine dihydrochloride (0.5%), the absorbance of the supernatant was determined with a U2001 photometric reader (Hitachi, Naperville, IL). To examine specificity of CYP-induced cystitis, we also measured MPO activity of descending colon (1 cm) from the same rats.

In a separate series of experiments, the effect of inflammation on bladder weight and tissue pH was assessed. On day 6, rats were anesthetized (pentobarbital, 50 mg/kg ip), the bladder was exposed by an abdominal incision, and pH was measured by inserting a needle (1 mg of Evans blue onto the incision after removal of the bladder. The damaged urothelium made it more difficult to reliably determine the extent of penetration of the pH electrode into the bladder wall in CYP-treated rats. To confirm appropriate electrode placement during measurement of bladder wall pH, we withdrew the electrode following determination of bladder wall pH, applied ~1 mg of Evans blue onto the penetration site, and gently compressed the bladder to rule out a small leak. We then removed the bladder and measured its weight after emptying the contents. Euthanasia was accomplished by exsanguina- tion after removal of the bladder.

Bladder Neuron Labeling

Under pentobarbital anesthesia (50 mg/kg ip), the bladder was surgically exposed (lower abdominal incision ~1 cm in length) and 1.1‘-dioctadecyl-3,3,3’,3’-tetramethyl indocarbocyanine methanesulfonate [DiI, 180 mg in 2 ml of DMSO; Molecular Probes, Eugene, OR] was injected into 6–8 sites within the wall of the bladder and around the trigone using a 30-gauge needle (~6 μl per site). Any visible leakage of DiI from the injection site was removed with a cotton swab. The incision was closed with 4.0 silk suture, and rats were allowed to recover. Two-three weeks later, saline (control) or CYP (100 mg/kg) was administered systemically (ip) on days 1, 3, and 5 as described above.

Cell Dissociation and Plating

For patch-clamp experiments, rats were anesthetized on day 6 (pentobarbital, 50 mg/kg ip) and lumbosacral (LS, L6–S2) or thoracolumbar (TL, T13–L2) DRGs were harvested for acute dissociation and whole cell recordings. The general protocols have been previously described (Dang et al. 2005a, 2005b). Briefly, after removal, ganglia were minced and incubated at 37°C, 5% CO2 for 60 min in serum-free, supplemented Neuro-A medium (B27 supplements, 5%; t-gluta-	amine, 0.5 mM; penicillin-streptomycin mixture, 1%; all from Gibco, Invitrogen, Grand Island, NY) containing collagenase (type 4; 2 mg/ml), and trypsin (1 mg/ml; both from Worthington Biochemical, Lakewood, NJ). Tissue fragments were gently triturated to encourage cell dissociation. Cells were collected by 5 min of centrifugation at 150 g, washed 3 times with supplemented Neuro-A medium (without enzymes), and resuspended in supplemented, enzyme free Neuro-A medium. The cells were plated on poly-o-lysine-coated coverslips (Becton Dickinson Labware, Bedford, MA) and incubated at 37°C, 5% CO2 for 2–3 h before electrophysiological studies. Acutely dis- sociated neurons were round and devoid of any processes, thus reducing potential space-clamp errors. Only bladder sensory neurons (i.e., DiI-containing DRG neurons) were studied. All recordings were performed within 10 h after plating.

Solutions and Electrophysiological recordings

Coverslips with cells were transferred to a recording chamber (1 ml) superfused continuously (2 ml/min) with external solution containing (in mM) 140 NaCl, 5 KCl, 2 MgCl2, 2 CaCl2, 10 HEPES, and 10 glucose. The pH was adjusted to 7.4 with NaOH (310 mosM). Under low magnification (×50), bladder neurons were identified by DH content using a rhodamine filter (bright orange/red color under UV light; excitation wavelength, 530–560 nm; and barrier filter, 573–648 nm). Fire-polished micropipettes with tip resistances of 1.5–2.0 MΩ were used for whole cell recordings. The uncompensated series resistance was generally about 7 MΩ or less. The pipette was filled with an internal solution consisting of (in mM) 130 KCl, 1 CaCl2, 1 MgCl2, 10 EGTA, 10 HEPES, 4 NaATP, 0.5 Tris-GTP, and 0.5 GDP. The pH was adjusted to 7.2 using KOH (310 mosM). After the whole cell configuration was established, the voltage was clamped at ~70 mV using an Axopatch 200B amplifier (Axon Instruments, Sunnyvale, CA), digitized at 1 kHz (Digidata 1350; Axon Instruments) and controlled by Clampex software (pClamp 9; Axon Instruments). Disruption of the neuron membrane during DRG dissociation and the use of patch-clamp recordings are unavoidable. We chose whole cell capacitance as a well-established surrogate marker of size to minimize additional confounders due to distortion in neuron shape. Cell capacitance was obtained by reading the value from the Axopatch 200B amplifier. Recordings began 2–3 min after whole cell configura- tion was established, to ensure stable recording conditions.

Drugs were applied using a fast-step SF-77B superfusion system (Warner Instruments, Hamden, CT) with a three-barrel pipette placed in close proximity (100 μm) to the cell, allowing complete solution exchange within 25 ms (Dang et al. 2005a). Agonists were applied for 4 s, whereas antagonists were superfused for 60 s before the application of agonists. A washout period of 2 min was allowed between agonist applications. Unless otherwise stated, drugs and chemicals were obtained from Sigma-Aldrich (St. Louis, MO) and prepared fresh from stock solutions on the day of the experiment. All experiments were performed at room temperature (21–23°C). We used current increases above 20 pA

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as threshold to identify responses to agonists, because this level exceeded the baseline variability by a factor of 2.

Heat Application

After whole cell configuration was established and voltage was clamped at −70 mV, preheated external solution (∼80°C; same composition as above) was superfused for 30 s. We placed a temperature-sensitive microelectrode (∼100 μm in front of the recording cell (Physitemp, Clifton, NJ). Heat threshold (°C) was defined as the temperature when the line fitted to the heat-induced current intercepts the X-axis (time, s). Only one cell per coverslip was studied with a single application of heat to avoid complications that may result from repeated exposure to high temperatures.

Data Analyses

Data are means ± SE. Analyses were performed using the software Clampfit (Axon Instruments) and GraphPad Prism 4 (GraphPad Software, San Diego, CA). Sigmoidal concentration-response curves were generated using the following equation: \( Y = A/(1 + \exp(-\log EC_{50} - X/B)) \), where \( X \) is the logarithm of concentration and \( Y \) is the response, which starts at 0 and goes to \( A \) with a sigmoidal shape. \( B \) is the Hill slope, \( I_{max} \) is the maximum current, \( EC_{50} \) is the concentration of agonist that evokes half-maximal response, and \( pH_{50} \) is the logarithm of \( EC_{50} \) of protons. To determine the kinetics of response onset, we measured the rise time, which is the time required for a signal to change from 10 to 90% of the peak amplitude. Desensitization kinetics were fitted with a standard exponential function: \( Y = K_0 + K_t \exp(-t/\tau) \), where \( Y \) is the current amplitude at time \( t \), \( K_0 \) is the amplitude of the nondesensitizing component, \( K_0 + K_t \) is the peak current, and \( \tau \) is the time constant. \( K_0 \) and \( K_t \) represent the contribution to current amplitude from the fast and slow components of the current, respectively. For dichotomous variables, a \( \chi^2 \) test was employed. Unless otherwise indicated, results were evaluated using Welch’s \( t \)-test after logarithmic transformation. One-way ANOVA were employed to compare and confirm variables (e.g., pH50 values and desensitization constants of acid-evoked currents) in LS or TL neurons from saline-treated rats. Results were considered to be statistically significant when \( P < 0.05 \).

RESULTS

CYP Produces Bladder Damage and Inflammation (Cystitis)

Consistent with a previous study (Dang et al. 2008), the weight of bladders from CYP-treated rats was significantly greater than that from saline-treated rats (saline: 95 ± 4.8 g, \( n = 12 \); CYP: 273 ± 8.5 g, \( n = 12 \); \( P < 0.001 \)). CYP treatment significantly and selectively increased bladder MPO activity relative to controls (saline: 0.36 ± 0.05 unit/g, \( n = 4 \); CYP: 1.0 ± 0.29 unit/g, \( n = 4 \); \( P < 0.001 \)); colon MPO activity was not affected by systemic administration of CYP (saline: 0.63 ± 0.06 unit/g, \( n = 4 \); CYP: 0.56 ± 0.07 unit/g, \( n = 4 \); \( P > 0.05 \)).

Cystitis Reduces pH of the Bladder Wall and Increases Size of Bladder Sensory Neurons

Although urine pH (36–37°C) from saline- and CYP-treated rats did not differ (saline: 6.3 ± 0.09, \( n = 12 \); CYP: 6.4 ± 0.07, \( n = 12 \); \( P > 0.05 \)), pH of the bladder wall was significantly reduced in CYP-treated rats compared with controls (saline: 7.3 ± 0.03, \( n = 12 \); CYP: 7.0 ± 0.03, \( n = 12 \); \( P < 0.01 \)). Based on whole cell capacitance, bladder inflammation significantly increased the size of bladder sensory neurons (saline LS: 28.6 ± 1.6 pF, \( n = 60 \); saline TL: 49.3 ± 1.2 pF, \( n = 53 \); CYP LS: 36.3 ± 1.2 pF, \( n = 62 \); CYP TL: 57.6 ± 2.3 pF, \( n = 57 \); \( P < 0.05 \)).

Cystitis Alters Acid- and Capsaicin-Produced Currents in Bladder Neurons

We previously described the effects of acid and capsaicin on rat bladder sensory neurons (Dang et al. 2005a). In bladder neurons from saline-treated rats, we found in the present study that the proportions of responsive TL and LS neurons, peak current density, and kinetics in response to acid and capsaicin were similar to those we previously reported (Dang et al. 2005a), as summarized in Tables 1–4. Briefly, almost all LS and TL bladder neurons responded to acid (pH 5) with mixed current kinetics (a mixture of fast, intermediate, and/or sustained components), and all acid-sensitive bladder neurons responded to capsaicin (1 μM) with a sustained current only. On the basis of complete inhibition of acid-evoked currents by amiloride, an ASIC channel blocker, we concluded that transient currents triggered by acid were mediated by ASICs (Dang et al. 2005a). Capsaizepine and amiloride partially antagonized the sustained component of acid-triggered currents, suggesting that this component was mediated, in part, by TRPV1 receptors (Dang et al. 2005a). A-425619 (100 nM, 60 s), a competitive TRPV1 receptor antagonist (El Kouhen et al. 2005), abolished current triggered by capsaicin (100 nM), confirming involvement of TRPV1 (data not shown).

Biphasic, slow, and triphasic currents. In bladder neurons from CYP-treated rats, acid (pH 5) produced currents in all 62 LS and 52 of 57 TL bladder neurons (not different from saline-treated controls; Table 1). Relative to saline treatment, CYP treatment increased the proportion of TL bladder neurons exhibiting acid-produced triphasic currents (CYP: 28/52, 53.8%; saline: 15/48, 31.3%; \( P < 0.05 \)) and reduced the number of TL bladder neurons exhibiting acid-produced slow currents (CYP: 7/52, 13.5%; saline: 16/48, 33.3%; \( P < 0.05 \)). Proportions of acid-produced biphasic, slow, and triphasic currents in LS bladder neurons were unaffected by CYP treatment (see examples in Fig. 1). See Table 1 for details.

Current density \( I_{max} \) and \( pH_{50} \). Acid (pH 5) produced significantly lower densities of fast, intermediate, slow, and sustained current components in both TL and LS bladder neurons from CYP-treated rats relative to their saline-treated counterparts (Table 2). Because we noted a reduction in current density in response to pH 5, we further examined \( I_{max} \) and \( pH_{50} \) values for fast, intermediate, and sustained currents triggered by acid application, the most prevalent components in bladder neurons. Whereas peak current densities (\( I_{max} \)) were significantly reduced in LS (e.g., Fig. 1, E–G, and Table 2) and TL (e.g., Fig. 1, I–K, and Table 2) bladder neurons from CYP-treated rats, bladder inflammation did not alter \( EC_{50} \) values for proton-triggered current components in these neurons (Table 3).

Current kinetics. Bladder inflammation did not influence activation and inactivation kinetics of acid-evoked currents in bladder sensory neurons (Table 4). Because slowly activating and desensitizing current was identified in only a small number of bladder neurons, we did not systematically examine recovery kinetics. Based on single-exponential fittings, recovery time constants of transient currents in LS and TL bladder neurons from CYP-treated rats did not differ in comparison with neurons from saline-treated rats (e.g., Fig. 2).
Table 1. Number of bladder sensory neurons responding to protons and capsaicin and type of current produced

<table>
<thead>
<tr>
<th></th>
<th>Sal LS</th>
<th>Sal TL</th>
<th>CYP LS</th>
<th>CYP TL</th>
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<tbody>
<tr>
<td>No. of responders</td>
<td>55/60</td>
<td>48/53</td>
<td>62/62</td>
<td>52/57</td>
</tr>
<tr>
<td></td>
<td>(91.7)</td>
<td>(90.6)</td>
<td>(100)</td>
<td>(100)</td>
</tr>
<tr>
<td>Biphasic current with fast and intermediate components</td>
<td>15/55</td>
<td>17/48</td>
<td>16/62</td>
<td>17/52</td>
</tr>
<tr>
<td></td>
<td>(27.3)</td>
<td>(35.4)</td>
<td>(25.8)</td>
<td>(32.7)</td>
</tr>
<tr>
<td>Slow current</td>
<td>14/55</td>
<td>16/48</td>
<td>19/62</td>
<td>7/52*</td>
</tr>
<tr>
<td>Triphasic current with fast, intermediate, and sustained components</td>
<td>26/55</td>
<td>15/48</td>
<td>27/62</td>
<td>28/52*</td>
</tr>
<tr>
<td></td>
<td>(47.2)</td>
<td>(31.3)</td>
<td>(43.5)</td>
<td>(53.6)</td>
</tr>
</tbody>
</table>

Values are no. of lumbosacral (LS) and thoracolumbar (TL) bladder sensory neurons from saline (Sal; control)- and cyclophosphamide (CYP)-treated rats that responded to application of protons (pH 5) and capsaicin, with percentage of neurons (in parentheses) expressing biphasic, slow only, and triphasic currents in response to pH 5. *P < 0.05 vs. saline-treated counterparts.

Capsaicin current density is increased. Virtually all bladder neurons responded to capsaicin (1 μM), and the proportion responding to capsaicin did not change after CYP-treatment (Table 1). In contrast to acid-evoked currents, capsaicin triggered significantly greater peak current density in both TL and LS bladder neurons (e.g., Fig. 1, H and L, and Table 2).

Protons inhibit responses to acid but increase responses to capsaicin. Because bladder wall pH is significantly decreased in this model of cystitis, we increased systematically the concentration of protons (pH range 7.4–6.6) by bathing bladder neurons for 60 s before application of acid (pH 5) or capsaicin (100 nM). In both LS and TL bladder neurons, prolonged exposure to pH 7.0 inhibited both rapidly and intermediately desensitizing current components triggered by acid (Fig. 3, A and B). As shown in Fig. 3, C and D, inhibition of acid-sensitive currents by extracellular acidosis was concentration dependent. A decrease from pH 7.4 to pH 7.2 depressed the response triggered by acid by more than one-half in both LS (Fig. 3C) and TL (Fig. 3D) bladder neurons.

In contrast to acid-sensitive currents, lowering bath pH significantly increased the current density evoked by capsaicin in LS bladder neurons (195.0 ± 40, 308.6 ± 88, 333.4 ± 89, and 356.0 ± 101%, n = 6, for pH 7.2, 7.0, 6.8, and 6.6, respectively; Fig. 4, A, B, and E). Similarly, extracellular acidosis significantly increased current triggered by capsaicin in TL neurons (157.2 ± 17, 253.7 ± 35, 286.9 ± 50, and 313.2 ± 75%, n = 5, for pH 7.2, 7.0, 6.8, and 6.6, respectively; Fig. 4, C, D, and E).

Cystitis and Acid Reduce Bladder Neuron Response Thresholds to Heat

Considering the role of TRPV1 in thermal sensation, we examined the effect of increasing the temperature of the bath solution. We defined the heat activation threshold as the temperature at which X- and Y-axes intercept (Fig. 5A). Heat activated inward
currents with a threshold of 41.4 ± 1.2°C (n = 11) and 42.0 ± 0.9°C (n = 11) in LS and TL neurons, respectively, from saline-treated rats. Relative to their saline-treated counterparts, CYP-treated rats had significantly reduced heat activation threshold for both groups of bladder neurons (CYP LS: 37.7 ± 0.9°C, n = 11; TL: 36.9 ± 0.6°C, n = 12; P < 0.01; Fig. 5B).

To assess the role of TRPV1, we superfused A-425619 (300 nM) for 1 min, followed by administration of heated solution with A-425619. In the presence of A-425619, heat activation thresholds were significantly elevated to 44.5 ± 0.4°C (n = 6) and 45.1 ± 1.1°C (n = 6) for LS and TL bladder neurons, respectively (P < 0.05), suggesting that TRPV1 receptors mediate, in part, the current triggered by heat (Fig. 5C).

We further examined modulation of heat-activated threshold by lowering bath solution pH to pH 7.0 for 1 min, followed by application of preheated solutions (pH 7.0, 30 s). Heat triggered currents at significantly lower thresholds in pH 7.0 compared with pH 7.4 solution (LS: 37.1 ± 1.3°C, n = 5; TL: 38.3 ± 0.8°C, n = 6, P < 0.05; Fig. 5D). Effects of CYP treatment and other pharmacological manipulations are presented in Fig. 5, E and F.

**DISCUSSION**

We extended our prior investigations of ASIC and TRPV1 currents in identified bladder sensory neurons and in this study examined the relative contributions and changes of these currents after bladder inflammation. The key finding is a differential effect of cystitis on distinct current components. Whereas bladder inflammation suppressed proton-evoked currents, it increased peak current density in response to TRPV1 activation by capsaicin and lowered the activation threshold of TRPV1 to heat. Despite the attenuation of acid-induced current, acidification of the extracellular milieu, as seen during cystitis, enhanced responses to capsaicin, further supporting a role of TRPV1 rather than other acid-sensitive channels in peripheral sensitization during bladder inflammation.

CYP treatment produced a significant inflammation that was limited to the bladder, consistent with previous reports (Bon et al. 2003; Cox. 1979; Dang et al. 2008). Based on cell capacitance, bladder neurons from CYP-treated rats were increased in size, consistent with previous reports (Dang et al. 2008; Yoshimura and de Groat 1999). Although we did not address the mechanism(s) for the change in cell size, others have shown that neutralization of NGF prevents the increase in size of neurons innervating an inflamed hind paw (Nicholas et al. 1999). NGF increases during cystitis (Liu et al. 2009; Murray et al. 2004; Oddiah et al. 1998), suggesting that NGF may have contributed to the increase in cell size in the present study. The toxic CYP metabolite acrolein activates TRPA1 receptors, which also contribute to nociceptive responses. Both TRPV1 and TRPA1 play important roles in the development of neuropathic inflammation, which may indirectly or directly affect neuron structure (Bautista et al. 2006). As indicated above, we noted a significant decrease in bladder wall pH during inflammation, which may contribute to sensitization of afferent pathways. Similar effects have previously been shown by others (Cohen et al. 2009; Deval et al. 2010; Friesen et al. 2007; Holzer 2011; Nagae et al. 2007), suggesting that in addition to changes in epithelial permeability, inflammation contributes to tissue acidosis. We recognize that other factors may contribute to the increased excitability of primary bladder afferents and bladder over activity (Arms et al. 2013; Dang et al. 2008; Yoshimura and de Groat 1999). The toxic CYP metabolite acrolein directly affects TRPA1 channels through covalent modification of cysteine residues, thus potentially contributing to the development of neuropathic inflammation (Maehelson et al. 2007). Prior studies have demonstrated that TRPV1 and TRPA1 are coexpressed in rat bladder neurons (Streng et al. 2008). These TRP channels are also functionally linked, as has been demonstrated by cross-sensitization and desensitization (Andrade et al. 2006; Du et al. 2007; Ruparel et al. 2008). We did not directly address the role of TRPA1 in the present study, but it is unlikely that it affected acid-evoked currents, the focus of the present investigation. Because experiments were conducted after and not during induction of CYP-produced cystitis, the

**Table 2. Current density in response to protons or capsaicin in bladder sensory neurons**

<table>
<thead>
<tr>
<th>Component</th>
<th>Sal LS</th>
<th>Sal TL</th>
<th>CYP LS</th>
<th>CYP TL</th>
</tr>
</thead>
<tbody>
<tr>
<td>pH50, pA/pF</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fast</td>
<td>55.4 ± 8.3 (41)</td>
<td>58.2 ± 9.0 (32)</td>
<td>33.6 ± 7.5* (43)</td>
<td>32.2 ± 4.3* (45)</td>
</tr>
<tr>
<td>Intermediate</td>
<td>64.4 ± 10.3 (41)</td>
<td>54.8 ± 11.1 (32)</td>
<td>41.9 ± 5.2* (43)</td>
<td>32.9 ± 4.2* (45)</td>
</tr>
<tr>
<td>Slow</td>
<td>55.8 ± 10.3 (44)</td>
<td>64.5 ± 7.3 (16)</td>
<td>29.6 ± 9.0* (19)</td>
<td>36.5 ± 9.2* (7)</td>
</tr>
<tr>
<td>Sustained</td>
<td>91.6 ± 12.9 (26)</td>
<td>32.8 ± 7.1 (15)</td>
<td>49.6 ± 9.4* (27)</td>
<td>14.2 ± 3.9* (28)</td>
</tr>
<tr>
<td>Capsaicin</td>
<td>142.4 ± 19.1 (55)</td>
<td>89.6 ± 11.2 (49)</td>
<td>258.2 ± 21.0* (61)</td>
<td>254.6 ± 15.2* (57)</td>
</tr>
</tbody>
</table>

Values are means ± SE of current density for various components triggered by application of pH 5 or capsaicin in LS and TL bladder sensory neurons taken from saline (control)- and CYP-treated rats (no. of neurons in parentheses). All comparisons were made between saline and CYP treatment. *P < 0.05 vs. saline-treated counterparts.
potential sensitizing effect of acrolein on coexpressed members of the TRP channels is similarly unlikely to skew results.

Bladder neurons exhibit multiple and complex current responses during exposure to protons, indicating that multiple subtypes of ASICs coexist and contribute to heteromultimeric channel complexes (Benson et al. 2002; Hesselager et al. 2004; Holzer 2011; Kobayashi et al. 2009; Mamet et al. 2002; Nagae et al. 2006, 2007; Voilley et al. 2001). Based on EC50 values, the transient proton-evoked currents share similar acid sensitivity. Relative to its potency in heterologous expression systems, acid was significantly less potent at ASICs in the native sensory neurons studied presently. The difference in potency may be due to the presence of accessory and/or regulatory protein(s) in bladder sensory neurons, which has previously been shown for TRPV1 receptors (Kim et al. 2006). Cystitis significantly enhanced during inflammation, suggesting a differential modulation of TRPV1 and ASIC channels. Considering these findings and the importance of TRPV1 in nociception, we examined the modulation of this current during inflammatory conditions. Because capsaicin at nociception, we examined the modulation of this current during inflammatory conditions. Because capsaicin at

Table 4. Current kinetics in response to protons in bladder sensory neurons

<table>
<thead>
<tr>
<th>Time to peak 10–90%, ms</th>
<th>Sal LS</th>
<th>Sal TL</th>
<th>CYP LS</th>
<th>CYP TL</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fast</td>
<td>111.2 ± 10.4 (41)</td>
<td>136.9 ± 23.9 (32)</td>
<td>90.8 ± 11.4 (43)</td>
<td>115.6 ± 10.9 (45)</td>
</tr>
<tr>
<td>Intermediate</td>
<td>639.4 ± 31.0 (41)</td>
<td>630.9 ± 45.3 (32)</td>
<td>586.0 ± 40.3 (43)</td>
<td>715.3 ± 44.8 (45)</td>
</tr>
<tr>
<td>Slow</td>
<td>914.0 ± 79.4 (14)</td>
<td>1,002.0 ± 77.3 (16)</td>
<td>812.5 ± 100.2 (19)</td>
<td>1,049.0 ± 126.3 (7)</td>
</tr>
<tr>
<td>Desensitization constant, ms</td>
<td>Fast 225.1 ± 41.2 (41)</td>
<td>252.9 ± 23.1 (32)</td>
<td>180.5 ± 19.1 (43)</td>
<td>233.5 ± 23.9 (45)</td>
</tr>
<tr>
<td></td>
<td>Intermediate 661.5 ± 96.4 (41)</td>
<td>815.9 ± 168.6 (32)</td>
<td>704.5 ± 78.9 (43)</td>
<td>934.0 ± 191.3 (45)</td>
</tr>
<tr>
<td></td>
<td>Slow 3,433.0 ± 997 (14)</td>
<td>4,022.0 ± 706 (32)</td>
<td>3,033.0 ± 506 (19)</td>
<td>3,349 ± 811 (7)</td>
</tr>
</tbody>
</table>

Values are means ± SE of current kinetics for various components triggered by application of pH 5 in bladder sensory neurons from saline (control)- and CYP-treated rats (no. of neurons in parentheses). *P < 0.05 vs. saline-treated counterparts.
well above the reported EC50 (Correll et al. 2004; Dray et al. 1990; Gunthorpe et al. 2000; McLatchie and Bevan 2001). Based on antagonism of capsaicin by A-425619, cystitis was associated with a significant increase in TRPV1-mediated peak current density, consistent with a previous study reporting similar augmentation of TRPV1-mediated current in a mixture of labeled/unlabeled LS neurons from cats with IC. It is unlikely that allosteric modulation of the channel contributes to the increased current in this study, because allosteric modulation of TRPV1 by ions has been reported to be transient, lasting only minutes after removal from the bath fluid (Caterina et al. 2000; McLatchie and Bevan 2001; Tominaga et al. 1998). In contrast, capsaicin activated TRPV1 receptors with increased current in bladder neurons from CYP-treated rats; these neurons were removed from the rats several hours previously and bathed in control pH (7.4), suggesting long-lasting alteration(s) had occurred. Although we did not directly determine channel expression or other mechanisms contributing to the observed changes, others have shown that inflammation increases the basal activity of PKC and/or the expression or functional properties of TRPV1 receptors in sensory neurons after tissue inflammation/damage (Amaya et al. 2004; Rashid et al. 2003; Zhu et al. 2011). As mentioned above, cystitis significantly increases levels of NGF in urine and receptor tyrosine kinase A (TrkA) in the mouse bladder (Murray et al. 2004; Oddiah et al. 1998). Furthermore, overexpression of NGF in the bladder triggers bladder hyperactivity and hypersensitivity (Boudes et al. 2011; Lamb et al. 2004; Schnegelsberg et al. 2010; Yoshimura et al. 2006). NGF has been shown to upregulate expression of TRPV1, which likely contributed to the effects observed in the present study (Mamet et al. 2002, 2003; Saban et al. 2011; Shimosato et al. 2005; Simonetti et al. 2006; Xue et al. 2007; Zhang et al. 2005).

Inflammation lowers tissue pH, as we confirmed in the present study, prompting us to determine the effect of acidosis on TRPV1-mediated current. Acute exposure to a slightly acidic environment significantly enhanced capsaicin- and heat-evoked currents, thereby potentially further enhancing excitability (see below). We did not directly examine the relative contribution of TRPV1 to the heat response in neurons from CYP-treated rats because we only applied heat once per neuron per coverslip to avoid complications with repeated exposure to heat. Although heat is a nonselective activator of TRPV1 channels, TRPV1 receptors play a role in temperature detection in rat bladder sensory neurons, because heat activation threshold was raised from 41 to 45°C in the presence of A-425619, consistent with previous reports demonstrating a key role of TRPV1 receptors in thermal sensation (Caterina et al. 1997). The present data suggest that enhanced function of TRPV1 may contribute, in part, to the lower heat activation threshold in bladder neurons from CYP-treated rats. Interestingly, the TRPV1 receptor antagonist A-425619 did not completely abolish current triggered by heat, suggesting that other heat modulated channels may be coexpressed in bladder neurons (Caterina et al. 1999; Hiura 2009; Smith et al. 2002; Tamura et al. 2005). Although other TRP channels may be affected by bladder inflammation, it is beyond the scope of this study and remains to be investigated.

**Significance of Plasticity of TRPV1 Receptor Function and Its Modulation by Protons**

Taken together, the present data demonstrate that cystitis differentially regulates the function of ASIC and TRPV1 channels, depressing ASIC-mediated currents while enhancing the TRPV1-mediated current. In addition to changes in channel expression or modulation of expressed channels, the proton...
concentration increases during inflammation, which further modulates channel function. Prolonged exposure to protons during tissue acidosis desensitizes the transient ASIC currents, suggesting that ASIC channels may participate in sensory detection under normal physiological conditions but do not significantly contribute to peripheral sensitization during inflammatory conditions such as cystitis. In contrast, bladder inflammation augmented TRPV1-mediated current and reduced the response threshold to heat; both effects can be enhanced by tissue acidification, suggesting that TRPV1 could be readily activated by normal body temperature and/or endogenous activators such as anandamide and protons. This study supports an important role of TRPV1 in the sensitization of bladder afferent pathways.

**Fig. 4.** Proton enhancement of current triggered by capsaicin in LS (A and B) and TL (C and D) bladder neurons from saline-treated rats. In both LS and TL neurons, capsaicin (100 nM) produced an inward current (A and C, first trace) that was potentiated in the presence of pH 7.0 (A and C, second trace) and remained enhanced after a 2-min washout (WO) following exposure to pH 7.4 (A and C, third trace). Capsaicin-triggered currents recovered 4–6 min after removal of pH 7.0 (B and D). Potentiation of capsaicin-evoked inward currents by protons showed strong pH dependency (E) in both LS and TL bladder neurons.

**Fig. 5.** Activation threshold to heat and its modulation. Bath application of preheated solution triggered an inward current with an activation threshold defined as the temperature at which lines extrapolated from the X (time, s)- and Y (current, pA)-axes intercept. Extrapolation to the temperature ramp (bottom trace) yielded the activation threshold (°C). Examples are given in A–D; data are summarized in E and F for LS and TL neurons, respectively. In bladder neurons from saline-treated rats (A), heat triggered current at a threshold of 42°C. The heat-triggered current threshold was significantly reduced in bladder neurons from CYP-treated rats (B), significantly increased in the presence of A-425619, a selective TRPV1 antagonist (C), and significantly reduced by brief reduction of the external solution to pH 7.0 (D). See text for details. *P < 0.05.
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DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS

K.D., K.B., and G.F.G. conception and design of research; K.D. performed experiments; K.D. analyzed data; K.D., K.B., and G.F.G. interpreted results of experiments; K.D. prepared figures; K.D. drafted manuscript; K.D., K.B., and G.F.G. edited and revised manuscript; K.D., K.B., and G.F.G. approved final version of manuscript.

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