Dopamine D2 Receptor–Mediated Presynaptic Inhibition of Olfactory Nerve Terminals

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INTRODUCTION

Odor signals are transduced by olfactory receptor neurons in the nasal epithelium and relayed to the glomeruli of the main olfactory bulb (MOB) via the olfactory nerve (ON). In the glomeruli, ON terminals form glutamatergic, axodendritic synapses with mitral and tufted (M/T) cells (Aroniadou-Anderjaska et al. 1997; Berkowicz et al. 1994; Ennis et al. 1996), the output cells of the MOB, and with juxtaglomerular (JG) interneurons. The first stage of synaptic processing in the olfactory system, little is known about intraglomerular synaptic mechanisms that process sensory input.

Previous studies demonstrate that most JG cells contain GABA (Ribak et al. 1977) and/or dopamine (DA) (Davis and Macrides 1983; Halasz et al. 1981; McLean and Shipley 1988). The DA JG neurons are abundant; the rat MOB contains more DA neurons (100,000–150,000) (McLean and Shipley 1988) than the entire substantia nigra and ventral tegmental area midbrain DA system (~30,000) (Björklund and Lindvall 1984). Many JG cells colocalize GABA and DA (Gall et al. 1987; Kosaka et al. 1985). In the rat, 69% of immunoreactive neurons in the glomerular layer (GL) colocalize DA and GABA, while only 4% contained DA alone (Gall et al. 1987; Kosaka et al. 1985).

The roles of GABAergic and dopaminergic JG neurons in glomerular synaptic processing are not understood, although recent studies suggest that JG interneurons may function, in part, to presynaptically regulate sensory input to the glomeruli. GABA_B receptors are present on ON terminals in the glomeruli (Bonino et al. 1999; Margeta-Mitrovic et al. 1999), where they inhibit glutamate release from the ON (Aroniadou-Anderjaska et al. 2000). Anatomical evidence also indicates that DA D2 receptors are present on ON terminals. Olfactory receptor neurons express DA D2 receptors, and in the MOB, D2 receptors are localized exclusively in the ON and glomerular layers (Coronas et al. 1997; Koster et al. 1999; Nickell et al. 1991). Bullectomy, a manipulation that causes retrograde degeneration of olfactory receptor neurons, eliminates D2 receptor mRNA in the olfactory epithelium (Koster et al. 1999). These findings suggest that D2 receptors are expressed by olfactory receptor neurons and are translocated to ON terminals in MOB.

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Previous studies have shown that D2 receptors in the MOB are functional, since DA reduces Ca^{2+} influx in ON terminals (Wachowiak and Cohen 1999) and reduces ON-evoked synaptic responses of M/T cells in rats (Hsia et al. 1999). However, D2 receptor expression has also been reported in JG cells (Mansour et al. 1990) and D1-like (i.e., D1 and D5 subtypes) ligand binding is present at very low levels in the subglomerular layers of the MOB (Coronas et al. 1997; Nickell et al. 1991). Thus it is still unclear whether DA inhibits transmission from the ON to the MOB via postsynaptic actions, and whether such inhibition is mediated solely by the D2 receptor subtype. Additionally, the influence of D2 receptor activation on spike output from mitral cells, and the role of these receptors in regulating JG neuronal activity are unknown. To address these issues, we investigated the actions of DA on spontaneous and ON-evoked activity in M/T and JG cells in rats, and in wildtype mice and in mice with targeted deletion of the D2 receptor gene.

**METHODS**

**Animals**

The following experimental procedures were conducted so as to minimize animal suffering and the number of animals used, and were approved by the animal welfare committee of the University of Maryland. Juvenile (12–28 day old) male rodents (Sprague-Dawley rats from Zivic Miller, C57BL/6 mice from Jackson Labs, and DA D2 knockout mice from our colony) were used. The generation of DA D2 receptor knockout (D2 knockout) mice has been reported previously (Baik et al. 1995). The D2 knockout line had been backcrossed to C57BL/6J, and all mice were at least 95% C57BL/6J. Mice were genotyped by polymerase chain reaction (PCR) of DNA in tail tip digests. Tail tips were digested at 56°C in 20 μL of proteinase K. The PCR sample (25 μL) contained 1 μL of DNA digest in 1 times PCR buffer, 1.5 mM MgCl₂, 0.2 mM each dTTP, dCTP, dATP, dGTP, and 0.625 U of Taq polymerase. The oligonucleotide primers were 5 pmol/25 μL for neomycin resistance gene (Neo) or 20 pmol/25 μL for the D2 receptor. The thermocycler program was initiated by a 1-min denaturation step at 95°C followed by 40 amplification cycles: 95°C for 1 min, 60°C for 1 min, 72°C for 1 min. 

**Slice preparation**

Rat and mouse MOB slices were prepared as previously described (Aroniadou-Anderjaska et al. 1997, 1999; Ciombor et al. 1999). Horizontal slices (400 μm thick) were transferred to an interface or submersed recording chamber maintained at 25–30°C and were continuously perfused at a rate of 1–2 ml/min with oxygenated artificial cerebrospinal fluid (ACSF) containing (in mM) 125 NaCl, 3.5 KCl, 2.5 CaCl₂, 1.3 MgCl₂, 26 NaHCO₃, and 10 d-glucose (pH 7.4). Experiments were initiated 1–2 h after the slices were placed in the chamber. Focal stimulation pulses were applied to the slice with a bipolar electrode (paired 50-μm-diam stainless steel wires, insulated except for bluntly cut tips). Isolated, constant-current monophasic square-wave stimuli (10–400 μA in amplitude, 0.1 ms in duration) were delivered by a Grass S8800 stimulator.

**Extracellular recordings**

Extracellular recordings from single mitral cells were obtained with glass micropipettes (2–3 μm tip diam, 10–20 MΩ) filled with a 2% solution of pontamine sky blue in 0.5 M sodium acetate. Electrode signals were amplified, discriminated, and displayed using conventional electrophysiological techniques as previously described (Ciombor et al. 1999). Spontaneous and ON-evoked spiking activity of mitral cells was acquired and analyzed using modified CED hardware and software as previously described (Ciombor et al. 1999). Field excitatory postsynaptic potentials (fEPSPs) were recorded in the GL with glass micropipettes (2–4 μm tip diam, 0.5–2 MΩ) filled with 2 M NaCl. fEPSPs were filtered (3 kHz low-pass), and digitized on-line at 10 kHz with Axon Instruments hardware and software (pClamp8). Measurement of fEPSP amplitudes was done with pClamp8 software. A moving average (5 points) of the data was generated using Origin 6 (Microcal Software, Northampton, MA). Group data, expressed as means ± SE, were statistically analyzed with paired t-tests.

**Whole cell recordings**

JG neurons were visualized with an Olympus BX50WI upright microscope equipped with a ×60 water immersion lens and near-infrared-DIC optics. They were identified based on their periglomerular location (Shipley and Ennis 1996), the size of their soma, and their high-input resistance (∼2 GΩ). All recordings were made at 25–28°C. Patch electrodes were prepared from Garner KG-33 glass tubing using a Narishige PP-830 puller. Series resistance (range 8–15 MΩ) was not compensated but was carefully monitored for constancy. Data were discarded when series resistance varied by >20% or was larger than 15 MΩ. The intracellular solution contained (in mM) 135 KCl (or 135 CsCl and 5 mM QX222), 10 HEPES, 2 Mg₂-ATP, 0.2 Na₂-GTP, and 0.5 EGTA; pH and osmolarity were adjusted to 7.3 and 280 mosM, respectively. Electrical signals were recorded using an Axopatch-200B amplifier (Axon Instruments), filtered at 5 kHz, and stored on videocassette. Evoked events were digitized on-line and stored on hard disk. Off-line, signals were filtered at 2 kHz and digitized at 20 kHz for capturing individual spontaneous events, and at 0.5–1 kHz for capturing long stretches of recordings. The liquid junction potential after achieving whole cell was about 4 mV and was subtracted from the membrane potentials presented below. The decay of synaptic currents was fitted to the following function: I(t) = A_f exp(−t/τ_f) + A_s exp(−t/τ_s) + C, in which I(t) was the amplitude of EPSCs at time t, A_f and A_s were the amplitudes of the fast and slow components, respectively, τ_f and τ_s were the decay time constants of the fast and slow components, respectively, and C was the residual current at the end of the fitting interval. As will be presented below, stimulation of the lateral olfactory tract (LOT) or mitral cell layer (MCL) produced prolonged synaptic responses in JG cells. The synaptic charge transfer of these responses was estimated by subtracting the holding current and then numerically integrating the remaining current beginning at the time that the stimulus was applied (Schoppa et al. 1998). All values are means ± SE.

**Chemicals**

The following agents were diluted in oxygenated ACSF and applied by bath perfusion: quinpirole (40–500 μM), quinpirole (50–100

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Dopamine reduces the ON-evoked fEPSP in the glomerular layer

Previous studies demonstrate that ON shocks elicit a two-component fEPSP in the GL that reflects glutamatergic synaptic currents generated in the apical dendrites of M/T cells (Aroniadou-Anderjaska et al. 1997, 1999). The first experiment investigated the effects of DA, and D2 receptor agonists and antagonists, on the ON-evoked fEPSP in the rat. Bath application of DA (40 μM) reduced the peak amplitude of the ON-evoked fEPSP from 1.2 ± 0.1 (SE) mV to 0.8 ± 0.1 mV (Fig. 1 A; n = 5, P = 0.005), a 30.1% reduction. Subsequent addition of the D2 antagonist sulpiride (100 μM) completely reversed the effect of DA and caused an increase in the fEPSP amplitude to 106% of control (Fig. 1 A, n = 5, * P = 0.0001). The inhibitory action of DA on the fEPSP was mimicked by the selective D2 receptor agonist, quinpirole. Quinpirole (100 μM) reduced the peak amplitude of the fEPSP from 0.9 ± 0.1 to 0.6 ± 0.1 mV (n = 10, P < 0.0001), a decrease of 32.7% (Fig. 1 C). Subsequent addition of the D2 antagonist sulpiride (100 μM) reversed the effects of quinpirole to within 99% of control levels (n = 10; Fig. 1 C). The effects of DA and quinpirole were reversible after wash out.

DA reduces paired-pulse depression of the ON-evoked fEPSP in the glomerular layer

Next, we examined whether D2 receptor activation affects M/T cell synaptic responses to paired-pulse stimulation of the rat ON. Two successive, identical stimuli delivered at various interstimulus intervals (ISIs) may produce depression (paired-pulse depression) or facilitation of the response to the second (test) pulse. Whether paired-pulse depression or facilitation is produced depends largely on the probability of transmitter release in response to the first (conditioning) pulse (Debanne et al. 1996; Manabe et al. 1993; Thompson et al. 1993). If DA acts presynaptically to reduce the probability of glutamate release from ON terminals, then the ratio of conditioning versus test response amplitude should be altered (i.e., the degree of paired-pulse depression or facilitation). By contrast, if DA acts postsynaptically, both conditioning and test responses should be reduced to the same degree, and therefore their amplitude ratio will remain unchanged.

As shown in Fig. 1 B, paired-pulse stimulation of the ON (100-ms ISI) produced a pronounced depression of the second (test) fEPSP; the test fEPSP was 52.3 ± 7.0% of the conditioning shock fEPSP (n = 4). DA (40 μM) disproportionately reduced the conditioning versus test fEPSPs. In the presence of DA, the amplitude of the test fEPSP was 65 ± 10.4% of the conditioning fEPSP, a significant reduction of the degree of paired-pulse depression (Fig. 1 B; n = 4, P = 0.01). In the presence of sulpiride, DA did not affect the degree of paired-pulse depression; the test fEPSP was 52.0 ± 4.6% of the conditioning response (n = 4), similar to paired-pulse depression in control media. These results suggest that DA reduced the probability of glutamate release from ON terminals, but they do not rule out concomitant postsynaptic effects.

DA reduces spontaneous and ON-evoked spiking of mitral cells

To determine whether D2 receptor activation has an impact on the output from M/T cells, we examined the effects of DA...
on ON-evoked mitral cell spiking in the rat. Most mitral cells exhibit a characteristic biphasic excitatory response to single ON shocks consisting of an early, brief excitation, followed by a short period of inactivity, and then a later, prolonged spiking component lasting up to 700 ms (Aroniadou-Anderjaska et al. 1997; Ennis et al. 1996). As shown in Fig. 2, DA application (100–300 μM) reduced the early spiking component in all mitral cells tested by 43.0–100%; mean reduction, 73.1 ± 4.4% (n = 20, P < 0.0001). DA also decreased the late spiking component in all cells by 27.0–100%; mean reduction, 70.3 ± 6.0% (n = 19, P = 0.005). In addition, DA (100–300 μM) decreased the baseline spontaneous discharge rate of 15/18 mitral cells. The mean discharge rate decreased from 2.8 ± 0.6 to 1.7 ± 0.5 spikes/s, a 26.9 ± 12.9% reduction (n = 18, P = 0.02; Fig. 2, A and B). The actions of DA were fully reversible. Similar inhibitory effects were produced by the D2 receptor agonist, quinelorane (100–300 μM), which decreased both the spontaneous and ON-evoked activity of mitral cells (Fig. 2B). Spontaneous discharge rate was reduced by 37.6 ± 8.2% (n = 12, P = 0.005). The early spiking component was reduced by 61.6 ± 12.0% (n = 13, P = 0.0006). The late spiking component was also depressed in six of nine cells, although this trend did not reach statistical significance (mean reduction, 53.5 ± 12.9%, P = 0.06). The four remaining cells did not exhibit a late spiking component.

As summarized in Fig. 2B, the D2 receptor antagonist eticlopride (10 μM) increased the mean spontaneous discharge rate of mitral cells from 1.5 ± 0.3 to 2.4 ± 0.8 spikes/s (n = 8, P = 0.02), a 60.7 ± 43.4% increase. Eticlopride also increased the magnitude of the early (234.5 ± 122.2%, n = 9,
\[ P = 0.02 \text{ and late (423.8 \pm 320.1\%), } n = 6, P = 0.02 \] spiking components evoked by ON stimulation. These results suggest that D2 receptors may be tonically activated by endogenous DA.

**Characteristics of ON-evoked EPSCs in JG cells**

JG cells receive glutamatergic synaptic input from the ON. Thus if DA presynaptically inhibits ON terminals, then it should also reduce ON-evoked, glutamate-mediated excitatory postsynaptic currents (EPSCs) in JG cells. To test this hypothesis, whole cell patch-clamp recordings were obtained from mouse JG cells. JG cell synaptic responses to ON inputs have not been investigated in mice. Therefore we first characterized the N-methyl-d-aspartate (NMDA) and \( \alpha \)-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptor-dependent excitatory postsynaptic currents (EPSCs) in a JG cell at different holding potentials; the pipette solution contained 135 mM CsCl, and the bath contained 10 \( \mu \text{M} \) bicuculline and 20 \( \mu \text{M} \) 6-cyano-7-nitroquinoxaline-2,3-dione (CNQX). The ON-evoked EPSC reversed around 0 mV. Note that the current-voltage (I-V) curve (B) exhibits a region of negative slope conductivity at holding potentials of \(-40 \text{ to } -80 \text{ mV}\), characteristic of NMDA receptor–mediated currents in the presence of extracellular Mg\(^{2+}\) (1.3 mM). C: kinetics of ON-evoked NMDA receptor–mediated EPSCs at 36 and \(-34 \text{ mV}\). The decay of these currents was slower at positive holding potentials than at negative ones. D and E: linear I-V relation of ON-evoked, AMPA receptor–dependent EPSCs; the pipette solution contained 135 mM CsCl, and the bath contained 10 \( \mu \text{M} \) bicuculline and 50 \( \mu \text{M} \) (–2)-2-amino-5-phosphono-penta-noic acid (APV). The ON-evoked non-NMDA EPSCs exhibited a linear I-V relationship and a reversal potential near 0 mV. Traces are averages of 2–4 sweeps.
over a range of holding potentials (from approximately −100 mV to approximately +50 mV, Fig. 3, A and B). EPSCs were relatively small at −60 mV or more negative holding potentials and were larger at potentials positive to −40 mV. The currents reversed polarity around 0 mV (Fig. 3, A and B). The characteristics of the current–voltage (I-V) curves for the NMDA receptor–mediated currents attest to the excellent voltage control of JG cells. The decay of these currents was faster at negative holding potentials than at positive ones (Fig. 3, C). The I-V relationship was linear after perfusion with a Mg²⁺-free bathing solution (n = 2, data not shown). In the presence of CNQX, application of 50 μM APV blocked ON-evoked EPSCs in all JG cells tested (n = 4, data not shown), confirming that they were mediated by NMDA receptors. These biophysical and pharmacological properties are characteristic for NMDA receptor–mediated currents (Konnerth et al. 1990; Lester et al. 1990; Nowak et al. 1984). Thus JG cells receive typical NMDA receptor–mediated synaptic inputs from ON terminals.

Next we characterized ON-evoked non-NMDA receptor–mediated EPSCs in the presence of 1.3 mM Mg²⁺, 50 μM APV, and 10 μM bicuculline. At a holding potential of −64 mV, ON shocks evoked relatively fast inward synaptic currents in 25 JG cells (Fig. 3, D and E). The peak amplitude was 450 ± 47 pA (n = 25). T½ rise time was 2.3 ± 0.2 ms (n = 25). The decay time course was variable with the 90% decay time ranging from 2 to 40 ms. The I-V relationship for ON-evoked, non-NMDA receptor–mediated EPSCs in JG cells in the presence of 1.3 mM Mg²⁺ was linear with a reversal potential around 0 mV in all JG cells tested (n = 4, Fig. 3, D and E), typical for non-NMDA receptor–mediated currents. ON-evoked EPSCs in the presence of APV were abolished on addition of 20 μM CNQX (data not shown).

**DA suppresses ON-evoked EPSCs in JG cells**

At a holding potential of −66 mV, bath application of 40 μM DA reversibly reduced the amplitude of the ON-evoked EPSCs in all 26 mouse JG cells. Most cells (20 of 26) were tested in the presence of 40–100 μM APV, and the rest were tested without APV. As the results were not different, the two data sets were pooled. EPSC amplitude in the presence of 40 μM DA was 38 ± 1.9% of control level (P < 0.01, n = 26, Fig. 4A). Despite the pronounced effect on ON-evoked EPSCs, we did not observe any DA-induced changes in holding current or input resistance in JG cells. In nine JG cells recorded with a KCl-based intracellular solution, the holding current and input resistance were −5.0 ± 1.5 pA and 2.1 ± 0.3 GΩ in the control condition, and −5.3 ± 1.6 pA and 2.0 ± 0.3 GΩ in the presence of 20–100 μM DA (P > 0.05). These results suggest that DA did not directly (i.e., postsynaptically) act on any of the JG cells sampled in this study.

Paired-pulse ON stimulation at ISIs of 50–400 ms produced pronounced depression (i.e., paired-pulse depression) of the second (test shock) evoked EPSC in JG cells. DA strongly and disproportionately depressed the conditioning versus the test EPSC, thereby decreasing the degree of paired-pulse depression. In control media, paired-pulse ON stimulation (100-ms ISI) reduced the amplitude of the test EPSC to 23 ± 0.3% of the conditioning EPSC. In the presence of 40 μM DA, the test...
EPSC was 82 ± 4% of the conditioning shock EPSC, a significant reduction in the degree of paired-pulse depression (Fig. 4B, P < 0.01, n = 17). The reduction of paired-pulse depression suggests that DA decreases the probability of glutamate release from ON terminals.

JG cells also receive excitatory glutamatergic input from the apical dendrites of M/T cells (Pinching and Powell 1971; Shipley and Ennis 1996). If indeed DA has no direct effects on M/T and JG cells, then JG cell EPSCs evoked by antidromic activation of M/T cells should not be affected by DA. In six mouse JG cells, lateral olfactory tract (1 cell) or mitral cell layer (5 cells) stimulation evoked bursts of EPSCs (Fig. 4C). Due to the bursting nature of the responses, we measured the total charge transfer induced by antidromic stimuli by calculating the integral of the response (see Methods). The charge transfer in the presence of DA (40–100 μM) was only slightly smaller than those under control conditions (90 ± 2% of control). This slight reduction was attributable to gradual rundown because strong stimuli (∼2 times the strength of ON shocks used) evoked antidromic responses that showed a tendency to slowly decline over time in the absence of DA. Under the same conditions, DA reduced ON-evoked EPSCs in JG cells to 30% of control values (see above). These results, taken with the finding that DA did not produce a measurable effect on holding current or input resistance in JG cells, indicate either that D2 receptors are not present in JG cells or that the receptors are too few in number or located at sites too remote (e.g., autoreceptors at DA release sites) to appreciably influence JG cell responses to M/T cell synaptic inputs.

DA depresses spontaneous, but not miniature, EPSCs in JG cells

To further investigate a presynaptic locus of DA action, we compared the effects of DA on the frequency and amplitude of action potential–dependent EPSCs versus action potential–independent miniature EPSCs in mouse JG cells. In four JG cells tested in the presence of 50 μM APV, DA reduced spontaneous EPSC (sEPSC) frequency by 44 ± 7.5% and amplitude by 42 ± 6.5% (P < 0.001; Fig. 5). The reduction in sEPSC frequency is consistent with a presynaptic locus of DA action. The reduction of sEPSC amplitudes could be attributed to a postsynaptic DA effect on JG cells and/or a reduction in the frequency of multiple, overlapping sEPSCs via presynaptic inhibition. To address this issue, we next investigated the effect of DA on the frequency and amplitude of spontaneous miniature EPSCs (mEPSCs). mEPSCs are believed to represent spontaneous, quantal release of transmitter from presynaptic sites in an action potential/Ca2+-independent manner. An effect of DA on the amplitude of mEPSCs would suggest a postsynaptic action of DA. In the presence of 0.5 μM TTX and 50 μM APV, mEPSCs were recorded in seven JG cells at −64 mV (Fig. 6). mEPSCs had a rapid time course with a T10–90 of 0.32 ± 0.01 ms (n = 7). A double exponential function with a fast and slow term was needed to fit the decay of the averaged mEPSC. The fast component had a decay time constant (τf) of 1.2 ± 0.05 ms, and the small slow component had a decay time constant (τs) of 5.0 ± 0.04 ms (n = 7). These kinetics are similar to those of non-NMDA receptor–dependent mEPSCs described elsewhere (Zhou and Hablitz 1997) and, consistent with this, mEPSCs were completely blocked by bath application of 10 μM CNQX (n = 3, data not shown). DA

**FIG. 5.** DA depresses spontaneous EPSCs (sEPSCs) in mouse JG cells. A–C: individual traces from a single cell showing examples of sEPSCs before (A), during (B), and after (C) application of 40 μM DA; the pipette solution contained 135 mM CsCl, and the bath contained 50 μM APV. Note that DA reduces the frequency of sEPSCs. D: histogram showing the cumulative probability of the amplitude of sEPSCs before and during application of 40 μM DA, and after DA wash out. DA significantly reduced the mean amplitude of sEPSCs from 62.5 ± 1.0 pA (control, n = 1,841) to 33.0 ± 0.6 pA (DA, n = 994, P < 0.001). sEPSC amplitudes returned to near control values (52.0 ± 0.9 pA, n = 1,555) after DA wash out. E: histogram showing the cumulative probability of the frequency of sEPSCs before and during application of 40 μM DA, and after DA wash out. DA significantly increased the mean interval between sEPSCs from 0.34 ± 0.1 s (control, n = 1,841) to 0.63 ± 0.3 s (DA, n = 994, P < 0.001). The interval between sEPSCs returned to near control values (0.35 ± 0.54 s, n = 1,555) after DA wash out.

**FIG. 6.** mEPSCs in mouse JG cells. A–C: individual traces from a single cell showing examples of mEPSCs before (A), during (B), and after (C) application of 40 μM DA; the pipette solution contained 135 mM CsCl, and the bath contained 50 μM APV. Note that DA reduces the frequency of mEPSCs. D: histogram showing the cumulative probability of the frequency of mEPSCs before and during application of 40 μM DA, and after DA wash out. DA significantly reduced the frequency of mEPSCs from 0.34 ± 0.1 s (control, n = 1,841) to 0.63 ± 0.3 s (DA, n = 994, P < 0.001). The interval between sEPSCs returned to near control values (0.35 ± 0.54 s, n = 1,555) after DA wash out.
DA has no effect on ON-evoked responses in D2 receptor knockout mice

D2-like actions can be produced by DA receptor subtypes D3 and D4 (Niznik and Van Tol 1992; Sibley and Monsma 1992). Previous studies have shown that ON terminals express D2 receptors, but it is not known whether D3 or D4 receptors are also present and thus contribute to the presynaptic effects shown in the preceding experiments. To address this question and to provide an alternative to pharmacological antagonism of the D2 receptors, we compared the effects of DA and D2 receptor agonists in D2 knockout mice versus wildtype littermates. Quinelorane (100 μM) differentially influenced the ON-evoked fEPSP in the GL recorded simultaneously in MOB slices from wildtype and D2 knockout mice. As shown in Fig. 7A, quinelorane suppressed the ON-evoked fEPSP in slices (n = 4) harvested from wildtype (37.5 ± 4.8% reduction; P < 0.02), but not D2 knockout mice (2.5 ± 2.5% reduction).

Next, we investigated the effect of DA on ON → JG cell mEPSCs in wildtype and D2 knockout mice. DA reduced the frequency, amplitude, or kinetics of mEPSCs (n = 5, P > 0.1; Fig. 6, A–D). Because JG cells receive glutamatergic synaptic inputs from M/T cells as well as ON terminals, the sEPSCs and mEPSCs could arise from either source. However, as the previous experiment showed that M/T cell–evoked EPSCs in JG cells were not affected by DA, these findings taken together support the hypothesis that DA acts to presynaptically inhibit glutamate release from ON terminals.

FIG. 6. DA does not affect miniature EPSCs (mEPSCs) in mouse JG cells. A and B: waveform of averaged mEPSCs before and during application of 40 μM DA; the pipette solution contained 135 mM CsCl, and the bath contained 0.5 μM TTX and 50 μM APV. DA did not alter the kinetics of mEPSCs. C: cumulative probability histogram shows that 40 μM DA did not affect the distribution of mEPSC amplitudes. The mean amplitude of mEPSCs before and during application of DA were 54.0 ± 6.4 pA (n = 217) and 54.0 ± 6.4 pA (n = 183, P = 0.47), respectively. D: cumulative probability histogram shows that 40 μM DA did not affect the distribution of mEPSC intervals. The mean EPSC intervals before and during application of DA were 4.5 ± 0.3 s (n = 217) and 5.2 ± 0.4 s (n = 183, P = 0.27), respectively.

The major finding of this study is that DA, a neurotransmitter contained in JG neurons, substantially decreases sensory input to the MOB by a presynaptic action on ON terminals. DA and D2 agonists markedly attenuated the responses of both M/T and JG cell excitatory responses to monosynaptic ON input. This inhibitory effect of DA is mediated exclusively by D2 receptors as DA was without effect in MOB slices harvested from mice with targeted deletion of the D2 receptor gene.

Dopamine decreases M/T and JG cell responses to ON input

DA significantly reduced the fEPSP recorded in the GL evoked by ON stimulation. This fEPSP reflects, for the most part, glutamatergic synaptic currents generated in the apical dendrites of M/T cells in response to ON input (Aroniadou-Anderjaska et al. 1997, 1999). The reduction of the ON-evoked GL fEPSP observed here with DA and quinpirole is comparable to the effects of quinpirole reported by Hsia et al. (1999). Consistent with the reduction of the ON-evoked fEPSP, DA significantly decreased both spontaneous and ON-evoked action potentials in mitral cells. The inhibitory effects of DA were mimicked by specific D2 receptor agonists, and prevented or reduced by D2 receptor antagonists. A reduction of ON-evoked spiking in mitral cells by DA and D2 receptor antagonists was recently reported in the turtle MOB (Berkowicz and Trombley 2000). Finally, DA also reduced spontaneous and ON-evoked EPSCs in JG cells. Thus DA or D2 agonists reduced spontaneous and ON-evoked synaptic responses in all MOB neurons known to be directly targeted by ON synaptic inputs.
It should be noted that bath-applied DA differs from synaptically released DA with regard to concentration, duration, and/or sites of action. Thus some of the actions of bath-applied DA observed in the present study may differ from the effects of DA released in response to odorant stimuli.

**DA effects are mediated by D2 receptors**

Anatomical studies demonstrate that the D2 receptor is the predominant DA receptor subtype expressed in the MOB (Coronas et al. 1997; Koster et al. 1999; Nickell et al. 1991). However, D1-like (i.e., D1 and D5 subtypes) ligand binding is present at very low levels in the subglomerular layers of the MOB (Coronas et al. 1997; Koster et al. 1999; Nickell et al. 1991). While the DA receptor agonists and antagonists used in the present study support the interpretation that DA’s actions are mediated by D2 receptors, D2 pharmacological reagents are not very precise tools. To circumvent this problem, we took advantage of a mouse line with a targeted deletion of D2 receptors (D2 knockout mice). In these D2 knockout mice, DA was completely ineffective in modulating the ON-evoked fEPSP and ON-evoked EPSCs in JG cells. This strongly supports the idea that D2 receptors, alone, function to presynaptically reduce glutamate release from ON terminals.

**Presynaptic locus of DA action**

Several lines of evidence from the present study indicate that DA presynaptically inhibits glutamate release from ON terminals. First, DA significantly altered the degree of paired-pulse depression of both M/T and JG cell synaptic responses to ON stimulation. In a paired-pulse paradigm, a postsynaptic action of DA should change the amplitudes of the conditioning and test responses proportionally (i.e., with no significant change in the degree of paired-pulse depression). The present experiments showed that DA significantly and disproportionately suppressed the amplitude of the conditioning versus test fEPSP recorded in the GL, thereby attenuating the degree of paired-pulse depression. Further, DA also attenuated the degree of paired-pulse depression of the ON-evoked EPSC in JG cells. Thus DA appears to reduce the probability of glutamate release from ON terminals onto all MOB neurons known to receive ON synaptic inputs.

Second, DA had no discernible influence on the membrane properties of JG cells. Because JG neurons have much smaller cell bodies and dendritic trees than mitral cells (Pinching and Powell 1971), they provide good space-clamp conditions that should optimize the ability to discern postsynaptic actions of DA. DA consistently suppressed both spontaneous and ON-evoked EPSCs in JG cells; this reduction was not accompanied by any detectable change in the holding current or input resistance in JG cells. This indicates that the predominant locus for the reduction of ON → JG cell EPSCs is pre- rather than postsynaptic.

Third, DA did not significantly alter M/T → JG cell synaptic responses. JG cells receive glutamatergic synaptic inputs from the intraglomerular apical dendrites of M/T cells (Pinching and
involved in D2-mediated inhibition of ON terminals in the rodent. Further, DA reduced sEPSCs, but not mEPSCs in JG cells. DA reduced both the frequency and amplitude of AMPA receptor–dependent sEPSCs. The decreased frequency of sEPSCs in the presence of DA is consistent with a presynaptic locus of action; the decreased sEPSC amplitude could be due to a postsynaptic locus of action or, alternatively, to a decreased incidence of temporally overlapping sEPSCs in JG cells. In agreement with this latter possibility, DA had no effect on the amplitude, frequency, and kinetics of mEPSCs recorded in JG cells in the presence of TTX. Since mEPSCs are believed to represent random release of single (i.e., quantal) neurotransmitter packets, the failure of DA to alter their characteristics provides further evidence that DA does not have postsynaptic actions on JG cells. Taken together, the most parsimonious explanation of these findings is that DA reduces glutamate release from ON terminals; a presynaptic mechanism. While the present results cannot rule out some small inhibitory postsynaptic action of DA, the evidence indicates that the majority, if not all, of the reduction of ON-evoked responses by DA is mediated by presynaptic D2 receptors on ON terminals. This is consistent with anatomical data showing that olfactory receptor neurons, whose axons form the ON, contain mRNA for D2 receptors, and that D2 receptor binding sites are restricted to the ON and GL (Coronas et al. 1997; Koster et al. 1999; Nickell et al. 1991).

There are several mechanisms by which DA could presynaptically inhibit glutamate release from ON terminals: D2 receptors are known to reduce transmitter release by suppressing $\text{Ca}^{2+}$ currents (Formenti et al. 1998; Koga and Moriyama 2000; Lledo et al. 1992; Wachowiak and Cohen 1999; Williams et al. 1990). Alternatively, D2 receptors could 1) increase potassium conductances, which would decrease the excitability of ON terminals (Lacey et al. 1988), or 2) directly modulate the release machinery, thereby reducing transmitter release (Man-Son-Hing et al. 1989; Wu and Saggau 1997).

Recent studies in the turtle showed that DA and D2 receptor agonists decrease $\text{Ca}^{2+}$ influx in ON terminals without affecting presynaptic ON action potentials (Wachowiak and Cohen 1999). DA and D2 receptor agonists were also reported to presynaptically inhibit glutamate release in the ventral tegmental area of the rat via a $\text{Ca}^{2+}$-dependent mechanism (Koga and Moriyama 2000). In the present experiments, DA reduced both the frequency and amplitude of sEPSCs but had no effect, however, on mEPSCs recorded in the presence of TTX. On the basis of these findings, we suggest that DA reduces voltage-activated $\text{Ca}^{2+}$ currents in ON terminals. Additional experiments are needed to identify the presynaptic mechanism involved in D2-mediated inhibition of ON terminals in the rodent MOB.

**Presynaptic regulation of ON terminals**

Olfactory receptor neurons expressing the same odorant receptors (an olfactory receptor neuron cohort) converge on the same MOB glomeruli (for review, see Mombaerts 1999; Mori et al. 1999). Odor molecules differentially activate multiple cohorts of olfactory receptor neurons, with the result that different odors evoke specific patterns of glomerular activity (Belluscio and Katz 2001; Friedrich and Korsching 1997; Guthrie et al. 1993; Johnson and Leon 1996; Johnson et al. 1998; Laurent 1996; Shepherd 1994). Less is known about the representation of odor concentration. If increasing concentrations of an odorant molecule cause increased neural activity among olfactory receptor neurons of the same cohort, then the range of concentrations that can be encoded might be limited by concentrations that produce maximal transmitter release by ON terminals. Presynaptic inhibition of ON terminals is a potential mechanism for increasing the range of concentrations that can be processed by MOB neurons: as activity increases in ON terminals, DA JG cells would be more strongly excited and thus exert negative feedback onto ON terminals, effectively increasing the dynamic range of information transfer from olfactory receptor neurons to MOB neurons.

In most mammals, investigative sniffing typically entails repetitive sniffs at 100- to 200-ms intervals (Komisaruk 1970; Welker 1964). Presynaptic inhibition of ON terminals may also play a role in determining spatial and temporal components of glomerular activation during repetitive sniffing by adjusting the level of glomerular excitation as a function of sniff frequency. We recently reported that GABA acting via GABA$_\text{A}$ receptors on ON terminals causes presynaptic inhibition (Aroniadou-Anderjaska et al. 2000) that is similar to that reported here for DA. Why are ON terminals presynaptically regulated by both DA and GABA? One possibility is that the functions conjectured above for DA (e.g., scaling concentration range, modulating ON input across sniff cycles) may be further enhanced by dual transmitter regulation. Another possibility may be related to the differential regulation of DA and GABA in JG neurons by ON activity. Manipulations that reduce ON synaptic activation of JG cells downregulates tyrosine hydroxylase and DA in JG cells (Baker 1990; Baker et al. 1983, 1984; Brunjes et al. 1985) but do not reduce GABA in the same JG neurons (Baker 1990). Thus if certain odor molecules are infrequently encountered, this might lead to downregulation of DA in JG cells in the glomeruli targeted by the olfactory receptor neurons activated by that odor molecule. Exposure to that odor would evoke less presynaptic inhibition by DA. Conversely, if the animal is tonically exposed to a background odor, the glomeruli activated by the odor might release more DA and thus produce greater presynaptic inhibition, which would decrease responses to the maintained background odor. As GABA is not regulated in an activity-dependent manner, this transmitter might function to presynaptically regulate ON terminals independently of the animal’s odor exposure history.

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