A Method to Estimate Synaptic Conductances From Membrane Potential Fluctuations

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Rudolph, Michael, Zuzanna Piwkowska, Mathilde Badoual, Thierry Bal, and Alain Destexhe. A method to estimate synaptic conductances from membrane potential fluctuations. J Neurophysiol 91: 2884–2896, 2004; 10.1152/jn.01223.2003. In neocortical neurons, network activity can activate a large number of synaptic inputs, resulting in highly irregular subthreshold membrane potential (V_m) fluctuations, commonly called “synaptic noise.” This activity contains information about the underlying network dynamics, but it is not easy to extract network properties from such complex and irregular activity. Here, we propose a method to estimate properties of network activity from intracellular recordings and test this method using theoretical and experimental approaches. The method is based on the analytic expression of the subthreshold V_m distribution at steady state in conductance-based models. Fitting this analytic expression to V_m distributions obtained from intracellular recordings provides estimates of the mean and variance of excitatory and inhibitory conductances. We test the accuracy of these estimates against computational models of increasing complexity. We also test the method using dynamic-clamp recordings of neocortical neurons in vitro. By using an on-line analysis procedure, we show that the measured conductances from spontaneous network activity can be used to re-create artificial states equivalent to real network activity. This approach should be applicable to intracellular recordings during different network states in vivo, providing a characterization of the global properties of synaptic conductances and possible insight into the underlying network mechanisms.

INTRODUCTION

Neocortical neurons in vivo are characterized by intense subthreshold synaptic activity, which is often called “synaptic noise.” This activity is particularly intense in activated states with desynchronized electroencephalogram (EEG), as expected from the high levels of firing (5–40 Hz) of cortical neurons during EEG-activated states (Evarts 1964; Steriade et al. 2001), combined with their remarkably dense level of interconnection (Braitenberg and Schüz 1998; DeFelipe and Fariñas 1992). The properties of synaptic noise during EEG-activated states were characterized by recent studies (Destexhe and Paré 1999; Paré et al. 1998), concluding that it is responsible for setting neocortical neurons into a “high-conductance state” (reviewed in Destexhe et al. 2003). The characteristics of high-conductance states are depolarized membrane potential (V_m) of around −65 mV (≈15 mV depolarized with respect to rest), a 3- to 5-fold diminished input resistance, and high-amplitude V_m fluctuations (SD of the V_m of about σ_v = 4 mV).

Several types of computational models have been proposed to investigate high-conductance states. Biophysically detailed computational models can integrate the dendritic morphology of cortical neurons, and simulate active channels in soma and dendrites, as well as the large number of excitatory and inhibitory synapses underlying background activity (Bernander et al. 1991; Destexhe and Paré 1999; Rudolph and Destexhe 2003a). On the other hand, simplified models consider single compartments (“point-neurons”) with global excitatory and inhibitory conductances (Destexhe et al. 2001). In this case, each global conductance represents the sum of a large number of individual synaptic inputs and is modeled by stochastic processes. The advantage of the latter approach is that the stochastic variations of synaptic conductances can be injected in real neurons to re-create high-conductance states in vitro, as shown in a number of recent studies (Chance et al. 2002; Destexhe et al. 2001; Fellous et al. 2003; Prescott and De Koninck 2003; Shu et al. 2003b) using the dynamic-clamp technique (Robinson and Kawai 1993; Sharp et al. 1993). These computational and dynamic-clamp approaches have shown that high-conductance states have a number of computational consequences on cortical neurons (reviewed in Destexhe et al. 2003). The stochastic and intense synaptic activity enhances their responsiveness (Hök and Destexhe 2000), modulates their gain (Chance et al. 2002; Fellous et al. 2003; Shu et al. 2003), sharpens the temporal processing of inputs (Bernander et al. 1991; Shelley et al. 2002; Shu et al. 2003b), or equalizes synaptic efficacies (Rudolph and Destexhe 2003a).

Another consequence of high-conductance states is that the subthreshold activity of any single neocortical neuron contains a large amount of information about the rest of the network. This is attributed to the particularly high level of firing activity of cortical neurons (see above), together with the dense intracortical connectivity (5,000 to 60,000 excitatory synapses per neuron; see DeFelipe and Fariñas 1992). Thus, neocortical neurons should provide a good “sampling” of the activity of a large number of neurons in the network, as indeed shown by the tight correlation between EEG and intracellular activity in cortex (Contreras and Steriade 1995; Creutzfeldt et al. 1996a,b; Klee et al. 1965). In principle it should be possible to deduce properties of network activity by analyzing the subthreshold dynamics of the V_m but unfortunately no such methods are yet available. The main difficulty is to relate collective properties at the network level into identifiable patterns of synaptic activity. At present, only global characterizations are possible, such as for example characterizing the mean rate of firing of the excitatory and inhibitory cells, which should translate into...
the mean value of global excitatory and inhibitory conductances. Interestingly, the variance of global synaptic conductances is related to the average amount of correlation present among presynaptic neurons (Destexhe et al. 2001), but it is presently very difficult to estimate the variance of conductances.

A possible path toward such a characterization is to obtain a good mathematical description of the dynamics of synaptic noise, and deduce useful relations between the \( V_m \) dynamics and presynaptic activity. However, the mathematical description must not be too complex, to allow inverting the relations and obtain characteristics of network activity as a function of the total synaptic current constant external (stimulating) current. Synaptic noise is described by these results have appeared in 2 conference abstracts (Destexhe well as in real neurons during active states in vitro. Part of this method of analysis we propose is tested against computational models of increasing complexity, as cortical neurons. The method of analysis we propose is tested 2 independent current terms.

**METHODS**

**Models of cortical neurons and synaptic noise**

To reproduce the stochastic membrane potential fluctuations and high-conductance state characterizing the dynamics of neocortical neurons in vivo, several types of neuronal models were used and compared (see Fig. 1).

**Effective point-conductance model**

The first model was a point-conductance model (Destexhe et al. 2001), which consisted in a single-compartment neuron described by the passive stochastic membrane equation

\[
C_m \frac{dV(t)}{dt} = g_L[E_i - V(t)] - \frac{1}{a} I_{nm}(t) + \frac{1}{a} I_m
\]

where \( V(t) \) is the membrane potential, \( a \) is the membrane area, \( C_m \) is the specific membrane capacitance, and \( g_L \) and \( E_i \) are the leak conductance density and reversal potential, respectively. \( I_m \) denotes a constant external (stimulating) current. Synaptic noise is described by the total synaptic current \( I_{syn}(t) \), which was decomposed into a sum of 2 independent current terms

\[
I_{syn}(t) = \sum_{i=1}^{N} g_{A} a E_{e,i} n(t) + \sum_{i=1}^{M} g_{A} a E_{i} m(t)
\]

where \( g_{A} \) and \( g_{i} \) are time-dependent global excitatory and inhibitory conductances, respectively, and \( E_{e} \) and \( E_{i} \) are their respective reversal potentials. \( g_{A} \) and \( g_{i} \) were described by one-variable stochastic processes similar to the Ornstein–Uhlenbeck process (Uhlenbeck and Ornstein 1930)

\[
\frac{d g_{A}(t)}{dt} = -\frac{1}{\tau_{m}} g_{A}(t) + \frac{2g_{i} \sigma_{i}}{\tau_{m}} \chi_{i}(t)
\]

where \( g_{A} \) and \( g_{i} \) are average conductances, \( \tau_{m} \) and \( \tau_{i} \) are time constants, \( \sigma_{i} \) and \( \sigma_{i} \) are noise SD values, and \( \chi_{i}(t) \) and \( \chi_{i}(t) \) denote independent Gaussian white noise processes of unit SD and zero mean.

The model was accessed both analytically and numerically. In the latter case, the membrane area of the compartment was \( a = 34,636 \) \( \mu m^2 \) (corresponding to the layer VI neocortical pyramidal cells from cat parietal cortex used in Destexhe et al. 2001), and passive parameters were \( g_L = 0.0452 mS/cm^2 \), \( E_i = -80 mV \), \( C_m = 1 \mu F/cm^2 \) (Destexhe and Paré 1999; Paré et al. 1998), \( E_e = 0 mV \) and \( E_i = -75 mV \). Other synaptic noise parameter values were chosen to obtain an average membrane potential of about \(-65 mV \) with SD around 4 mV characteristic for in vivo states of cortical neurons (Destexhe and Paré 1999; Paré et al. 1998), and were \( g_{A} = 12.1 nS \), \( g_{i} = 57.3 nS \), \( \sigma_{i} = 12 nS \), \( \sigma_{i} = 26.4 \), \( \tau_{m} = 2.73 ms \), and \( \tau_{i} = 10.49 ms \).

Simulations of the point-conductance model and its comparison with mean-value complex models are illustrated in Fig. 1. The point-conductance model (Fig. 1A) generates irregular subthreshold activity consistent with in vivo measurements (Destexhe et al. 2001). It is characterized by a Lorentzian power spectrum (Fig. 1C, dashed lines), as well as by a symmetric (Gaussian) distribution of excitatory and inhibitory conductances (Fig. 1D, dashed lines), resulting in a nearly symmetric amplitude distribution of the membrane potential \( V_m \) (Fig. 1E, dashed lines).

**Single-compartment model with individual noise sources**

The second model consisted in a single-compartment membrane with a more realistic representation of synaptic inputs, which were modeled by a large number of individual synaptic conductances. In this case, the synaptic current \( I_{syn}(t) \) in Eq. 1 was described by

\[
I_{syn}(t) = \sum_{i=1}^{N} m_{A} a E_{e,i} n(t) + \sum_{i=1}^{M} m_{A} a E_{i} m(t)
\]

where \( N \) and \( M \) denote the total number of excitatory and inhibitory synapses, modeled by \( \alpha \)-amino-3-hydroxy-5-methyl-4-isoxazolepro- pionic (AMPA) and \( \gamma \)-aminobutyric acid (GABA) postsynaptic receptors (Destexhe et al. 1998) with quantal conductances \( g_{A} \) and \( g_{i} \), respectively. \( m_{A} \) and \( m_{i} \) represent the fractions of postsynaptic receptors in the open state at each independent synaptic interface, and were described by the following kinetic equations

\[
\frac{dm_{A}(t)}{dt} = \alpha_{i}(t)[1 - m_{A}(t)] - \beta_{i}(t) m_{A}(t)
\]

where \( \alpha_{i} \) and \( \beta_{i} \) are respectively forward and backward binding rate constants for excitation (index e) and inhibition (index i). When a spike occurred in the presynaptic compartment, a pulse of transmitter was triggered such that \( T[i] = T_{max} \) for a short time period \( t_{dur} \) and \( T[i] = 0 \) until the next release occurs. These kinetic models of synaptic currents were as described previously (Destexhe et al. 1998), with kinetic parameters that were obtained by fitting the model to postsynaptic currents recorded experimentally. To simulate synaptic background activity, all synapses were activated randomly according to independent Poisson processes with mean rates of \( \nu_{A} \) and \( \nu_{i} \) for AMPA and GABA \( \alpha \) receptors, respectively. The activation of \( \alpha \)-methyl-d-aspartate (NMDA) receptors is minimal at the subthreshold levels investigated here, and were not included for simplicity.

The model was simulated numerically with passive properties as in the point-conductance model. Synaptic parameter were \( N = 4,972 \), \( g_{A} = 1,200 pS \), \( g_{i} = 600 pS \), \( \alpha_{i} = 1.1 \times 10^6 \), \( \beta_{i} = 670 s^{-1} \) for AMPA receptors, \( \alpha_{i} = 5 \times 10^7 s^{-1} \), \( \beta_{i} = 1.8 s^{-1} \) for GABA \( \alpha \) receptors, \( T_{max} = 1 \mu m \), \( t_{dur} = 1 \mu s \), \( \nu_{A} = 2.16 Hz \), and \( \nu_{i} = 2.4 Hz \).

Simulations of this model are illustrated in Fig. 1B. The power spectra of the total excitatory and inhibitory conductances are approximately Lorentzian (Fig. 1C, gray), whereas their amplitude distributions take a nearly symmetric (Gaussian) shape (Fig. 1D, gray). Also the \( V_m \) amplitude distribution follows an approximately symmetric behavior (Fig. 1E, gray). It is to be noted that the point-conductance model captures these properties remarkably well (compare gray areas with dashed lines in Fig. 1, C–E), thus suggesting that the Ornstein–
Uhlenbeck stochastic process yields a valid description of synaptic noise.

**Detailed biophysical model**

The third model consisted in a compartmental model of a neocortical layer VI pyramidal neuron obtained from morphological reconstructions of cells recorded in cat association cortex (Contreras et al. 1997). The passive properties (see point-conductance model) were adjusted by matching the model to intracellular recordings obtained in the absence of synaptic activity (Destexhe and Paré 1999). In some cases, voltage-dependent conductances were inserted in the soma, dendrites, and axon and were described by Hodgkin and Huxley (1952) type models. The latter model included 2 voltage-dependent currents, a fast Na⁺ current $I_{Na}$ and a delayed-rectifier K⁺ current $I_{Kd}$, for action potential generation (Traub and Miles 1991) with conductance densities of 8.4 and 7 mS/cm² (Huguenard et al. 1988; densities were 10 times higher in the axon), respectively. To account for the spike-frequency adaptation and afterhyperpolarization commonly observed in “regular-spiking” neurons, a slow voltage-dependent K⁺ current $I_{Kd}$ was added. In some simulations, models of cortical neurons with additional fast inactivating A-type K⁺ current $I_{KA}$ (model from Migliore et al. 1999; conductance density from Bekkers 2000), T-type (low threshold) Ca²⁺ current $I_{T}$ (model from Traub et al. 2003; conductance density from Hamill et al. 1991) and hyperpolarization-activated current $I_{h}$ (model and nonuniform conductance density from Stuart and Spruston 1998) were used.

To simulate synaptic inputs, pyramidal cells were divided into different regions (soma, perisomatic dendrites, main dendrites, axon initial segment) and the conductance of AMPA and GABA_A synapses was estimated from morphological studies (see DeFelipe and Farinas 2002; for kinetic parameters see single-compartment model with individual noise sources). The number of synapses per 100 μm² of membrane were: 10–20 (GABA_A, soma and perisomatic dendrites), 40–80 (GABA_A, axon initial segment), 8–12 (GABA_A, dendrites), and 55–65 (AMPA, dendrites), leading to a total of $N = 16,563$ glutamatergic and $M = 3,376$ GABAergic synapses. Synaptic currents were simulated by kinetic models of AMPA and GABA_A receptor types as described above, and synaptic background activity was simulated by random (Poisson-distributed) synaptic events at a mean rate of $v_{syn} = 1$ Hz and $v_{inh} = 5.5$ Hz for AMPA and GABA_A synapses, respectively. This model was described in detail in a previous study (Destexhe and Paré, 1999).

To estimate the conductances underlying synaptic activity, as well as their variances, we followed a procedure identical to that of a previous paper (Destexhe et al. 2001). An “ideal” voltage clamp (without electrode series resistance) was simulated using a somatic electrode. The model was run twice at 2 different clamped voltages (−65 and −55 mV), and using the same random seed (so that the same random numbers were used at each clamp). The leak-subtracted currents obtained were then decomposed into excitatory and inhibitory conductances using the relation...
\[
R(t) = g_{l}(t)(V - E_{l}) + g_{i}(t)(V - E_{i})
\]

where \(V\) is the clamped voltage. This procedure yields “effective” global synaptic conductances \([g_{l}(t), g_{i}(t)]\) as seen from a somatic electrode.

All simulations were performed using the NEURON simulation environment (Hines and Carnevale 1997) and were run on PC-based workstations under the Linux operating system.

**In vitro experiments**

In vitro experiments were performed on 0.4-mm-thick coronal or sagittal slices from the lateral portions of the ferret occipital cortex including primary and secondary (areas 17, 18, and 19) visual cortical areas. Ferrets, 4–12 mo old (Marshall Europe, Lyon), were anesthetized with sodium pentobarbital (30 mg/kg). The slices were maintained in an interface-style recording chamber at 35–36°C. Slices were prepared on a DSK microslicer (Ted Pella, Redding, CA) in a slice solution in which the NaCl was replaced with sucrose while maintaining an osmolarity of 307 mOsm. After transfer to the recording chamber, the slices were incubated in slice solution containing (in mM): NaCl, 124; KCl, 2.5; MgSO\(_4\), 2; NaHPO\(_4\), 1.25; CaCl\(_2\), 2; NaHCO\(_3\), 26; tetrodotoxin, 10, and was bubbled with 95% O\(_2\)–5% CO\(_2\) to a final pH of 7.4. After about 1 h, the slice solution was modified to contain 1 mM MgCl\(_2\), 1 or 1.2 mM CaCl\(_2\), and 3.5 mM KCl (Sanchez-Vives and McCormick 2000). Intracellular recordings after 2 h of recovery were performed in deep layers (layers IV, V, and VI) on electrophysiologically identified regular spiking and intrinsically bursting cells. Electodes for intracellular recordings were made on a Sutter Instruments P-87 micropipette puller from medium-walled glass (WPI, 1BF100) and beveled on a Sutter Instruments beveler (BV-10M). Micropipettes were filled with 1.2 to 2 M potassium acetate and had resistances of 80–100 MΩ after beveling.

Ferret visual cortical slices spontaneously display recurrent periods of activity lasting 0.5–1.5 s, which are separated by periods of quiescence lasting 2–20 s (Sanchez-Vives and McCormick 2000). In intracellular recordings, this active network activity manifests as a depolarized state (“up-state”). During the periods of quiescence (“down-state”), the membrane potential relaxes toward its resting value. To characterize synaptic noise, intracellular recordings in up- and down-states were collected at several different membrane potentials maintained by injection of steady currents through the recording micropipette (current-clamp).

**Dynamic-clamp experiments**

The dynamic-clamp technique (Robinson et al. 1993; Sharp et al. 1993) was used to inject computer-generated conductances in real neurons. Dynamic-clamp experiments were run using the hybrid RT-NEURON environment (developed by G. Le Masson, INSERM U378, Université de Bordeaux), which is a modified version of NEURON (Hines and Carnevale 1997) running under the Windows NT 4.0 operating system (Microsoft) on a PC equipped with a 1.4-GHz Pentium IV processor. NEURON was augmented with the capacity of simulating neuronal models in real time, synchronized with the intracellular recording. To achieve real-time simulations as well as data transfer to the PC for further analysis, we used a PCI DSP board (Innovative Integration, Simi Valley, CA) with 4 analog/digital (inputs) and 4 digital/analog (outputs) 16 bits converters. The DSP board constraints calculations of the models and data transfer process to be made with a high priority level by the PC processor. The DSP board allows input (e.g., the membrane potential of the real cell incorporated in the equations of the models) and output signals (the synaptic current to be injected into the cell) to be processed at regular intervals (time resolution = 0.1 ms). A custom interface was used to connect the digital and analog inputs/outputs signals of the DSP board with the intracellular amplifier (Axoclamp 2B, Axon Instruments) and the data acquisition systems (PC-based acquisition software ELPHY, developed by G. Sadoc, CNRS Gif-sur-Yvette, ANVAR, and Biologic). The dynamic-clamp protocol was used to insert the fluctuating conductances underlying synaptic noise in cortical neurons using the point-conductance model, similar to a previous study (Destexhe et al. 2001). A critical feature for dynamic clamp and for the present method is the accuracy of the absolute membrane voltage measure. A reliable method is to monitor and adjust manually the \(V_{m}\) offset during the course of the recording according to the known statistical value of the onset of action potentials in a given cell type (e.g., –55 mV for ferret pyramidal cells; Shu et al. 2003b). For this purpose, action potentials are triggered by square depolarizing pulses of current injected through the micropipette.

**Data acquisition and analysis**

Voltage traces from numerical simulations and experimental recordings were analyzed with respect to both their statistical properties and amplitude distribution \(\rho(V)\). In models, simulations were run using either passive models or models with active currents responsible for spike generation and adaptation (see above). In experiments, spontaneous up-states were collected using custom data acquisition software (ELPHY). In all cases, the data acquisition rate was 100 kHz (numerical simulations) or 20 kHz (experiments). To obtain sub-threshold \(V_{m}\) distributions, steady hyperpolarizing current was used such that the average \(V_{m}\) in up-states was between –75 and –65 mV. The remaining action potentials, if present, were cut using a time window of 10 ms centered around the spike (taking advantage of the fact that signals need not to be contiguous to calculate amplitude distributions).

\(V_{m}\) distributions were calculated using bin sizes of 0.2 mV for traces from both simulations and experiments. The distributions obtained were fitted using a Gaussian template function (e.g., Eq. A6 in the appendix), thereby providing directly the values for the average \(V_{m}\), \(\bar{V}\), and its SD, \(\sigma_{V}\). These estimates were also checked with standard statistical analysis tools for discrete data sets (Press et al. 1993).

To calculate these values from the analytic expressions of \(\rho(V)\), which usually do not allow explicit integration, we integrated \(\rho(V)\) numerically, using

\[
\bar{V} = \int_{-\infty}^{\infty} dV \rho(V)
\]

\[
\sigma_{V}^{2} = \int_{-\infty}^{\infty} dV (V - \bar{V})^{2} \rho(V)
\]

for the mean and SD of the voltage distributions, respectively.

Finally, the effective membrane area \((a)\) and the leak conductance density \((g_{L})\) can be estimated from experimental data by using injection of hyperpolarizing current pulses during periods of quiescent activity, yielding estimates of the membrane time constant \((\tau_{m})\) and of the resting input resistance \((R_{m})\).

\[
g_{L} = g_{i}/C_{m}
\]

assuming a fixed value for \(C_{m}\) (1 \(\mu F/cm^2\)). Note that this procedure for estimating \(g_{L}\) may be a potential source of error because even during periods of quiescent network activity, the membrane still receives background synaptic inputs (e.g., residual network activity or minia
tropic synaptic events). Ideally, synaptic currents should be blocked pharmacologically to faithfully estimate \(g_{L}\). However, this procedure is technically difficult, in particular during feedback experiments where the same cell is used for analyzing and re-creating high-conductance states (see results), and was therefore not attempted here.
**RESULTS**

We start by outlining the procedure used for estimating synaptic conductances from membrane potential ($V_m$) amplitude distributions. We next test this procedure against models of increasing complexity. Finally, we demonstrate the application of this approach to in vitro experiments and test the results obtained using dynamic clamp.

The VmD method: estimating synaptic conductances from membrane potential distributions

We consider the $V_m$ probability density function $p(V,t)$ of the point-conductance model defined in Eqs. 1–3. $p(V,t)$ describes the probability density that the membrane potential $V_m$ takes the value $V$ at time $t$. The time evolution of this probability density function is given by the Fokker–Planck equation (Risken 1984; see APPENDIX), which for the point-conductance model yields the following steady-state solution (Rudolph and Destexhe 2003b)

$$p(V) = \exp[A_1(V-E) + B_1(V-E)^2]
  + A_2 \arctan[B_2(V-E) + B_3(V-E)]$$

where $A_1$ and $B_1$ are voltage-independent terms depending on passive membrane and synaptic noise parameters (see Eq. A4 in the APPENDIX for full expression).

This analytic expression for the $V_m$ distribution was derived and analyzed in detail in a previous study (Rudolph and Destexhe 2003b). Here we focus on possible applications of this analytic approach to analyze subthreshold neuronal activity and analyzed in detail in a previous study (Rudolph and Destexhe 2003b). Here we focus on possible applications of this analytic approach to analyze subthreshold neuronal activity and analyzed in detail in a previous study (Rudolph and Destexhe 2003b). Here we focus on possible applications of this analytic approach to analyze subthreshold neuronal activity and analyzed in detail in a previous study (Rudolph and Destexhe 2003b).

Although this estimate could in principle be obtained by fitting Eq. 9 to experimentally measured $V_m$ distributions, in practice, this approach poses a number of problems. The main obstacle is the highly nonlinear dependency of the $V_m$ distribution on its parameters (Eq. 9). In general, fitting a highly nonlinear function of many parameters to experimental data results in local minima that may lead to spurious estimates (Press et al. 1993). To circumvent this difficulty, we need to simplify Eq. 9. Here, we can take advantage of the fact that the $V_m$ distribution is only weakly asymmetric in $V$, especially in the range of $V_m$ values typical of in vivo activity (−70 to −50 mV; see detailed analysis in Rudolph and Destexhe 2003b).

Furthermore, a very convenient symmetric approximation of Eq. 9 can be obtained and takes the form of a Gaussian distribution

$$p(V) = \exp\left[-\frac{(V - \bar{V})^2}{2\sigma_V^2}\right]$$

where $\bar{V}$ is the average $V_m$ and $\sigma_V$ is the standard deviation of the $V_m$. This Gaussian approximation can be obtained formally from Eq. 9 by second-order Taylor expansion around its peak value (see details in the APPENDIX). As we will see below, this expression provides an excellent approximation of the $V_m$ distributions obtained from models and experiments.

The advantage of using a simplified expression such as Eq. 10 is 2-fold. First, one can obtain an expression of the quantities $\bar{V}$ and $\sigma_V$ (very easy to measure in experiments) as a function of the synaptic conductance parameters (see Eqs. A5 and A7 in the APPENDIX), which allows physical interpretation of these quantities. For example, it can be seen that $\bar{V}$ is mostly determined by the static components of the synaptic conductances ($g_{e0}$ and $g_{i0}$), whereas $\sigma_V$ has a more complex dependency on synaptic noise parameters. It depends on both $g_{e0}$ as well as on $\sigma_e$ and $\sigma_i$.

The second and main advantage is that these expressions are mathematically simple enough to enable inverting them, which may lead to expressions of the synaptic noise parameters as a function of the $V_m$ measurements, $\bar{V}$ and $\sigma_V$. The stochastic passive membrane equation, Eq. 1, is characterized by 6 parameters describing excitatory and inhibitory conductance noise ($g_{e0}$, $g_{i0}$, $\sigma_e$, $\sigma_i$, $\tau_e$, and $\tau_i$). Two of these parameters, the noise time constants $\tau_e$ and $\tau_i$, are related to the decay time of synaptic currents and thus the kinetics of synaptic transmission. Therefore, these parameters are expected to show little variations from cell to cell, and can be fixed using power spectra of synaptic conductances deduced from voltage-clamp recordings (see Destexhe et al. 2001). In contrast, the remaining 4 parameters, the means ($\bar{V}$) and SDs ($\sigma_e$, $\sigma_i$) of excitatory and inhibitory synaptic conductances, depend on the synaptic inputs converging to the cell as well as the actual network state. Thus these parameters are expected to vary from one situation to the other (e.g., between different network states), as well as from cell to cell (e.g., depending on the connectivity of that particular cell within the network), and should therefore be estimated for each case.

To extract the 4 conductance parameters ($g_{e0}$, $g_{i0}$, $\sigma_e$, $\sigma_i$) from the membrane probability distribution, Eq. 10 is, however, insufficient because it is characterized by only 2 parameters ($\bar{V}$, $\sigma_V$). To solve this problem, one possibility is to consider 2 $V_m$ distributions obtained at 2 different constant levels of injected current $I_{ext1}$ and $I_{ext2}$ (2 current-clamps protocol). In this case, expressing these 2 $V_m$ distributions as $\bar{V}_{1,2}$ and $\sigma_{V1,2}$ values for the $V_m$ SD, we can solve for the conductance parameters ($g_{e0}$, $g_{i0}$, $\sigma_e$, $\sigma_i$). In this case, one obtains

$$g_{e0} = \frac{(I_{ext1}I_{ext2})(\sigma_{V1}^2(E_{i0} - \bar{V})^2 - \sigma_{V2}^2(E_{i0} - \bar{V})^2)}{(\bar{V}_{1} - \bar{V}_{2})(E_{i0} - \bar{V}_{1})(E_{i0} - \bar{V}_{2})}
  - \frac{(I_{ext1}I_{ext2})(E_{i0} - \bar{V}_{1}) + (I_{ext1}I_{ext2})(E_{i0} - \bar{V}_{2}) - (E_{i0} - \bar{V}_{1})(E_{i0} - \bar{V}_{2})}{(E_{i0} - \bar{V}_{1})(E_{i0} - \bar{V}_{2})}$$

$$\sigma_{V1}^2 = \frac{2\sigma_{V1}^2}{\frac{1}{(E_{i0} - \bar{V}_{1})(E_{i0} - \bar{V}_{2})} + \frac{1}{(E_{i0} - \bar{V}_{1})(E_{i0} - \bar{V}_{2})}}$$

These relations enable us to estimate global characteristic of network activity, such as mean excitatory ($g_{e0}$) and inhibitory ($g_{i0}$) synaptic conductances, as well as their respective variances ($\sigma_{e0}^2$, $\sigma_{i0}^2$), from the sole knowledge of the $V_m$ distributions obtained at 2 different levels of injected current. This procedure, which we refer to below as the “VmD method,” is illustrated in Fig. 2 and constitutes the core of the analysis explored in this paper.

It is worth noting that the method can be generalized to various current levels. For one current level (one current-clamp...
FIG. 2. Sketch of the VmD method to estimate synaptic conductances from membrane potential fluctuations. A: membrane potential recordings of network activity at 2 different current levels $I_{\text{ext}1}$ and $I_{\text{ext}2}$ (top traces). Spikes are removed (bottom traces) or the activity is recorded at hyperpolarized levels to yield subthreshold activity. B: computation of membrane potential distributions (gray) at these 2 current levels and fitting with Gaussian function (solid lines), yielding 2 pairs of values for the average $V_m$ ($\bar{V}_1$, $\bar{V}_2$) and $V_m$ SD ($\sigma_{V1}$, $\sigma_{V2}$). These values are used to estimate analytically, by using Eqs. 11 and 12, the mean ($\bar{g}_e$, $\bar{g}_i$) and SD ($\sigma_e$, $\sigma_i$) of the conductances underlying network activity. From these, analytic forms of the membrane potential distributions, Eq. 9, characterizing subthreshold membrane dynamics attributed to synaptic activity can be obtained.

Protocol), 2 synaptic noise parameters can be extracted, such as the ratios between excitatory and inhibitory mean and SD. The $V_m$ distributions stemming from 3 injected current levels (3 current-clamps protocol) allow estimation in addition of the reversal potential of either excitatory or inhibitory conductances. Alternatively, multiple current clamps ($\geq 3$) can be used as consistency conditions for validating the obtained estimates. However, in what follows we restrict discussion to the 2 current-clamps protocol.

Test of the approach using computational models

We now turn to computational models of increasing levels of complexity to test the conductance estimates provided by Eqs. 11 and 12. First, we used the point-conductance model to check for the validity of the expressions obtained. Because of the equivalence of the underlying equations (Eqs. 1, 2, and 3), here the closest correspondence between estimated and actual (i.e., calculated numerically) conductance parameters is expected.

Figure 3 illustrates the procedure applied to the $V_m$ activity of that model (Fig. 3A). Two different values of steady current injection ($I_{\text{ext}1}$ and $I_{\text{ext}2}$) yield 2 $V_m$ distributions (Fig. 3B, gray). These distributions were fitted with a Gaussian function to obtain the means and SDs of the membrane potential at both current levels. Incorporating the values $V_1$, $V_2$, $\sigma_{V1}$, and $\sigma_{V2}$ into Eqs. 11 and 12 yields values for the mean and SD of the synaptic noise, $g_e$ and $g_i$, respectively (Fig. 3C solid line, and Fig. 3D). These estimates were then used to reconstruct the full analytic expression of the $V_m$ distribution using Eq. 9, which was plotted in Fig. 3B (solid lines). There was a very close match between the analytic estimates of $\rho(V)$ and numerical simulations (Fig. 3B, compare gray areas with solid lines). This demonstrates not just that Gaussian distributions are an excellent approximation for the membrane potential distribution in the presence of synaptic noise, but also that the proposed method yields an excellent characterization of the synaptic noise and thus subthreshold neuronal activity. This can also be seen by comparing the reconstructed conductance distributions (Fig. 3C, solid lines) with the actual conductances recorded during the numerical simulation (Fig. 3C, gray). Distributions deduced from the estimated parameters were in excellent agreement with those of the numerical simulations. Thus, this first set of simulations shows that, at least for the point-conductance model, the proposed approach provides a method that allows an accurate estimate of the mean and the variance of synaptic conductances from the sole knowledge of the (subthreshold) membrane potential activity of the cell.

A second test was to apply this method to a more realistic model of synaptic noise, in which synaptic activity was generated by a large number of individual synapses releasing randomly according to Poisson processes (Fig. 1B). An example of the application of the procedure to this type of model is shown in Fig. 4. Starting from the $V_m$ activity (Fig. 4A), membrane potential distributions were constructed and fitted by Gaussians for 2 levels of injected current (Fig. 4B, gray). Estimates of the mean and variance of excitatory and inhibitory conductances were then obtained using Eqs. 11 and 12. The analytic solution reconstructed from this estimate (Fig. 4B, solid lines) is in excellent agreement with the numerical
simulations of this model. Moreover, the reconstructed conductance distributions based on this estimate (Fig. 4C, solid lines) are also in excellent agreement with the total conductance calculated for each type of synapse in the numerical simulations (Fig. 4C, gray; see Fig. 4D for quantitative values and error estimates). Thus, also in case of this more realistic model of synaptic background activity, which markedly differs from the point-conductance model, the estimates of synaptic conductances and their variances from voltage distributions are in excellent agreement with the values obtained numerically. In fact, this agreement can be expected because of the close correspondence between the conductance dynamics in both models (see Fig. 1, C–E).

A third, more severe test was to apply this estimate to a compartmental model in which individual (random) synaptic inputs were spatially distributed in soma and dendrites. In a passive model of a cortical pyramidal neuron from layer VI (Fig. 5A; see METHODS), the $V_m$ distributions obtained at 2 steady current levels were approximately symmetric (Fig. 5B, left panel, gray). Again, applying Eqs. 11 and 12 to estimate synaptic conductances and their variances led to analytic $V_m$ distributions $p(V)$ (Fig. 5B, left panel, solid lines), which captured very well the shape of the $V_m$ distributions obtained numerically, although small deviations are visible at the hyperpolarized and depolarized tail of the distributions. The reconstructed conductance distributions (Fig. 5C, solid lines) were also in excellent agreement with the conductance distributions obtained in this model using an ideal voltage clamp at the soma (Fig. 5C, gray; see method in Destexhe et al. 2001). The quantitative comparison of those values (Fig. 5D, left panel) shows that the estimation from $V_m$ distributions gives comparable estimates as the ideal voltage clamp. This agreement also shows that the dendritic filtering of synaptic inputs, caused by the spatial extension of the dendritic tree, does have only a minor impact on the conductance estimation. Moreover, because of the higher density of GABAergic synapses in the proximal region of cortical neurons, a slight bias in the estimates toward inhibitory conductance is expected. However, our results indicate that this effect is small and has only a minimal impact on the overall conductance estimates.

Finally, by incorporating voltage-dependent currents ($I_{Na}$, $I_{Kd}$ for spike generation, and a slow voltage-dependent $K^+$ current for spike-frequency adaptation, a hyperpolarization-activated current $I_h$, a low-threshold Ca$^{2+}$ current $I_{CaT}$, and an A-type $K^+$ current $I_{KAT}$ with densities typical for cortical neurons; see METHODS) in the detailed biophysical model, we probed the applicability of the proposed method to more realistic situations with active dendrites capable of generating and conducting dendritic spikes. Here, deviations are expected because the method is strictly based on passive neuronal dynamics (see Eq. 1), which might be strongly altered by the presence of active channels at the site of the recording and the presence of regenerative dendritic spikes. Indeed, after removing spikes in a broad (10-ms) time window, the subthreshold activity approximated the passive dynamics, but showed deviations in the membrane potential distribution at its hyperpolarized and depolarized tails (Fig. 5B, gray, compare left and right). However, these deviations had only a minimal impact on the mean and variance of the membrane potential obtained by Gaussian fits, which constitute the input for the VmD method. Applying Eqs. 11 and 12 led to estimates (Fig. 5B, right panel, solid lines) that showed more significant—albeit still small—deviations from the distributions drawn from the corresponding numerical simulations (Fig. 5B, right panel, gray; Fig. 6, gray). In general, the estimated values for synaptic conductances and their variance showed larger errors, especially for $\sigma$ (Fig. 5D, right panel), and yielded $V_m$ distributions that were slightly broader. However, these errors and deviations remained relatively small and the method still provided a good estimate of synaptic conductances, similar to or better than the one provided by ideal voltage clamp (Fig. 6).

**Test of the method using in vitro recordings and dynamic-clamp experiments**

The method was further tested against real network activity. We used the recurrent activity (“up-states”) occurring spontaneously in ferret neocortical slices (see METHODS). Intracelluarly, this activity consists in a depolarized $V_m$ and relatively large-amplitude $V_m$ fluctuations (Fig. 7), as described previously (Sanchez-Vives and McCormick 2000). To test the method, we applied an on-line protocol (Fig. 7A), consisting of estimating synaptic conductances from “natural” up-states (top traces) and compared them to “artificial” up-states obtained by dynamic-clamp injection of the estimated conductances in the same neuron (bottom traces). As above, the estimates (Fig. 7A, solid distributions) were obtained by computing the $V_m$ distributions at 2 different current levels (Fig. 7A, bottom, gray distr...
FIG. 5. Estimation of synaptic conductances from the membrane potential activity of a detailed biophysical model of synaptic background activity. A: example of the membrane potential ($V_m$) activity obtained in a detailed biophysical model of a layer VI cortical pyramidal neuron (scheme on top; same model as in Destexhe and Pare 1999). Synaptic background activity was modeled by the random release of 16,563 AMPA-mediated and 3,376 GABA_A-mediated synapses distributed in dendrites according to experimental measurements. B: $V_m$ distributions obtained in this model at 2 different current levels, $I_{ext1}$ and $I_{ext2}$. Left panel: distributions obtained in a passive model. Right panel: distributions are shown when the model had active dendrites ($Na^+$ and $K^+$ currents responsible for action potentials and spike-frequency adaptation, located in soma, dendrites, axon). In both cases, results from the numerical simulations (gray) and analytic expression (solid lines), obtained by using the conductance estimates, are shown. C: histogram of the total excitatory and inhibitory conductances obtained from the model using an ideal voltage clamp (gray), compared to the distributions reconstructed from the conductance estimates based on $V_m$ distributions. D: bar plot showing the mean and SD of synaptic conductances estimated from $V_m$ distributions. Error bars indicate the statistical significance of the estimates by using different Gaussian approximations of the membrane potential distribution in B. Left panel: passive model; right panel: model with voltage-dependent conductances. Presence of voltage-dependent conductances had minor (<10%) effects on the estimated conductance values.

**DISCUSSION**

In this paper, we have provided a theoretical description of the subthreshold membrane potential activity of neurons under synaptic bombardment, and provided means to estimate the underlying global characteristics of network activity. We discuss here the advantages and problems of this approach, how it relates to previous work, and what perspectives are expected.

Starting from a purely theoretical approach, the mathematical description of the stochastic variations of $V_m$, we derived expressions to relate $V_m$ measurements to global synaptic conductance parameters, such as the mean excitatory ($g_{e0}$) and inhibitory ($g_{i0}$) conductances, as well as their respective variances ($\sigma_e$, $\sigma_i$). The mean synaptic conductances are related to the mean rate of afferent neurons, whereas the variances of these conductances are related to the level of correlation between presynaptic activities (Destexhe et al. 2001). So far, experimental measurements have concentrated on estimating
the mean conductances (Anderson et al. 2000; Borg-Graham 1998; Hirsch et al. 1998; Shu et al. 2003a), which is equivalent to estimating the mean level of afferent activity. Measuring conductance variances can give estimates of the mean level of correlation within afferent activity (Destexhe et al. 2001). However, such estimates were never provided so far, presumably because of the technical difficulty of measuring conductance variances in vivo, which presently requires voltage-clamp methods.

The present $V_m$ distribution method provides such estimates based on current-clamp recordings. It relies on quantities (mean and variance of the $V_m$), which are relatively easy to measure experimentally. Conductances are usually best estimated from voltage-clamp recordings (Borg-Graham et al. 1998), although methods from current-clamp recordings have been used as well (Anderson et al. 2000; Hirsch et al. 1998). The method proposed here relies on standard current-clamp protocols, in which spontaneous activity is recorded with steady current injection, which corresponds to the most current configuration used for performing intracellular recordings in vivo.

We have shown that the method can provide relatively good estimates of global synaptic conductances, even in the case of complex models, including the dendritic morpho-ology and active channels in dendrites (Fig. 5). In the latter case, the method provides measurements of global conductances and their variances with an accuracy comparable to that of an “ideal” voltage clamp. However, it must be noted that the method we used here to estimate conductance variances from voltage clamp required running the models twice with the same seed for random numbers. On the contrary, the proposed VmD method provides such an estimate without requirements of this type.

On the negative side, the method proposed here to estimate conductance variances relies on a series of parameters. First, there must be an estimation of the “effective” leak conductance (i.e., the nonsynaptic conductance) and membrane area $a$. These values can be obtained in vitro by measuring the input resistance and time constant in quiescent periods (see Methods). Second, voltage-dependent currents in soma and dendrites may distort the $V_m$ distribution and cause errors in the estimate, attributed to either the presence of regenerative dendritic spikes or the activation of voltage-dependent membrane conductances. The latter applies to all currents that are active in the subthreshold $V_m$ range, such as the hyperpolarization-activated current $I_h$. Slow $K^+$ currents may also be activated in the subthreshold range, although in our simulations (Fig. 5) there were slow $K^+$ currents, but their presence did not seem to have strong effects on the estimates. Various $K^+$ currents (such as those underlying spike afterhyperpolarization or the A-type $K^+$ current $I_{K_A}$), various $Ca^{2+}$ currents (such as the low-threshold $T$-current $I_{CaT}$), or $I_h$ may also significantly distort...
the \( V_m \) distribution (see Fig. 6), but the distorting effect of these currents can easily be avoided (see METHODS). Moreover, as our numerical simulations showed, despite a significant impact of these active currents, the estimates obtained with the VmD method were closer to the actual synaptic conductances compared to estimates with ideal voltage clamp. However, to minimize voltage-dependent effects, the best is to identify a linear region of the \( I-V \) curve of the neuron, and perform the measurements in that region. The excellent agreement obtained here using experimental recordings (Fig. 8) also suggests that these contaminations are minimal in the voltage range considered. Furthermore, using Gaussian fits of the \( V_m \) distributions effectively suppresses the effect of dendritic spikes arriving at the site of the recording and, thus, improves the applicability of the method. Third, the presented VmD method restricts, so far, only to glutamatergic and GABAA receptors. The impact of other receptor types, such as GABAB and NMDA, remains to be investigated and neglecting them might contribute to errors in the conductance estimates. Moreover, the knowledge of the reversal potential for GABAAergic or glutamatergic synapses is crucial and using wrong values of reversals will result in estimation errors. Ideally, the reversal potentials should be measured for the same preparation in which the analysis is made. Finally, drifts in the membrane potential (e.g., arising from the experimental setup or from unstability of the recording) constitute another source of error. The proposed method requires a stationary recording, which was the case for all cells shown here.

Another potential source of error comes from the spatial aspect of synaptic noise. Here, synaptic inputs received distally will contribute less to the somatic response compared to those close to the soma. This will, in general, result in an underestimation of the total synaptic conductances and their variation. Moreover, a nonuniform distribution of synaptic conductances, such as the higher density of GABAAergic synapses in the proximal region of cortical neurons, or the distance-dependent scaling of quantal conductances or receptor number, will bias the estimates for excitatory and inhibitory conductances. However, our simulations, which took the spatial distribution of (uniform) synaptic channels and the asymmetry in the distribution of glutamatergic and GABAAergic synapses into account, showed only minimal deviations from the corresponding voltage-clamp estimates of the excitatory and inhibitory synaptic conductances. In addition, our method yields estimates for the total synaptic conductances that determine the cellular dynamics at the site of the recording, a quantity that does not depend on a specific assumption of the distribution of synaptic channels within the dendritic tree.

This approach is also applicable to in vivo intracellular experiments. In this case, the mean and variance of synaptic conductances could be estimated across different states of the network. The difficulty, however, would be to estimate the resting parameters \( (g_{r}, a) \), which are not easy to deduce in vivo. However, if quiescent states can be obtained either pharmacologically (Paré et al. 1998) or spontaneously (“down-states”), then estimates of those parameters can be obtained. Alternatively, it is always possible to use the present approach with minor modifications to estimate relative changes of conductances or conductance variances between different states of the network. In particular, measuring changes in conductance variances should allow us to measure changes in correlation in network activity. Such measurements have not been obtained yet, but should be possible in the near future.
In this appendix we briefly summarize the mathematical approach for deducing the steady-state membrane potential distributions of the stochastic passive membrane equation (Eqs. 1–3) describing the subthreshold membrane dynamics in the presence of synaptic noise.

The Fokker–Planck equation

The stochastic passive membrane equation given in Eqs. 1–3 was analytically accessible within the framework of the stochastic differential calculus (Gardiner 2002; Mortensen 1969; van Kampen 1981). Here, using a set of differential rules (Ito rules) for the OU stochastic process allows deduction of the Fokker–Planck equation (Risken 1984), corresponding to the set of stochastic differential equations (Eqs. 1–3)

\[ \dot{\phi}(V, t) = -\frac{F}{C_m} \phi(V, t) + \frac{1}{C_m} \left[ I_{\text{inj}}(t) + I_{\text{ext}}(t) \right] \]

where

\[ f(V) = \frac{1}{aC_m} \left[ I_{\text{inj}}(t) - g_{\text{ext}}(V) - g_{\text{syn}}(V) - g_{\text{clus}}(V) \right] \tag{A2} \]

is a voltage-dependent drift term, \( h_{\text{exc}}(V) \) and \( h_{\text{inh}}(V) \) are voltage-dependent excitatory and inhibitory conductances noise terms, and

\[ 2\sigma_{\text{exc}}(t) = \sigma_{\text{exc}}^2 h_{\text{exc}}(1 - e^{-\tau_{\text{exc}}/\tau_e}) + \frac{1}{2\tau_e} \sigma_{\text{exc}}^2 \dot{h}_{\text{exc}}(t) - \sigma_{\text{exc}}^2 \dot{\phi}(V, t) \tag{A3} \]

where \( \dot{w}_{\text{exc}}(t) = \int_0^t ds_w \sigma_{\text{exc}}(s) \) are the integrated stochastic processes for the stochastic conductances \( \dot{h}_{\text{exc}}(t) \) and \( \dot{h}_{\text{inh}}(t) \) denote effective noise time constants given by \( \tau_{\text{exc}}(t) = 2\tau_{\text{exc}}\tau_e/\tau_e + \tau_e \), with \( \tau_{\text{exc}} = aC_m(a_{\text{exc}} + g_{\text{exc}} + g_{\text{inh}}) \). The Fokker–Planck equation (Eq. A1) describes the time evolution of the probability density function \( \rho(V, t) \) of the membrane potential \( V(t) \) in the presence of excitatory and inhibitory synaptic noise terms characterized by their mean and variances.

The steady-state membrane potential distribution

In the limit \( t \to \infty \), the Fokker–Planck equation (Eq. A1) can be solved analytically. In this case, one obtains the steady-state probability distribution \( \rho(V) \) for the membrane potential \( V \)

\[ \rho(V) = \text{Nexp} \left[ \ln \left( \frac{u(V-E_e)^2}{(aC_m)^2} + \frac{u(V-E_i)^2}{(aC_m)^2} \right) \right] \tag{A4} \]

where the following constants are defined: \( k_e = 2aC_m g_{\text{exc}} \), \( k_i = 2aC_m g_{\text{inh}} \), \( k_{\text{exc}} = 2aC_m g_{\text{exc}} \), \( k_{\text{inh}} = 2aC_m g_{\text{inh}} \), \( a_e = \sigma_{\text{exc}}^2 \tau_e \), and \( a_i = \sigma_{\text{inh}}^2 \tau_i \) as well as the following voltage-independent terms

\[ A_1 = -k_e + k_i + k_{\text{exc}} + k_{\text{inh}} \]

and

\[ A_2 = 2aC_m \frac{(E_i - E_e)(u_{\text{exc}} - u_{\text{inh}})}{(E_e - E_i)^2} \]

FIG. 8. Test of the method using natural and re-created up-states in vitro under dynamic clamp. A: reinjection of conductance estimates. Protocol used was similar as in Fig. 7 and consisted in first extracting conductances from natural up-states (arrow 1 in scheme) and re-creating artificial up-states in the same neuron (arrow 2). The natural up-states (top trace) were used to compute \( \rho(V) \) distribution and estimate conductances (middle panels, gray). These values were then used to generate artificial synaptic noise using stochastically fluctuating conductances \( \sigma_{\text{exc}}(t) \) and \( \sigma_{\text{inh}}(t) \), which were injected in the same neuron using dynamic clamp. The natural up-states \( \rho(V) \) activity shown as \( V(t) \) in bottom traces. \( \rho(V) \) distributions obtained were in excellent agreement (gray: natural up-states, \( V = -66.87 \) mV, \( \sigma_e = 1.53 \) mV; continuous line: re-created up-states, \( V = -66.81 \) mV, \( \sigma_e = 1.36 \) mV; B: analysis of artificial up-states produced by dynamic-clamp injection of known conductances. In this protocol, stochastically varying synaptic conductances were first injected in the neuron (arrow 1 in scheme). Resulting \( \rho(V) \) activity was then used to reestimate the conductances (arrow 2). Middle panel: injected (dark gray) and reestimated (light gray) conductances. Right panel: corresponding \( \rho(V) \) distributions (gray: experimental; solid lines: analytic prediction from the reestimated parameters). There was an excellent agreement between all values (injected conductances: \( g_{\text{exc}} = 2.1 \) nS, \( g_{\text{inh}} = 2.8 \) nS, \( \sigma_i = 1.0 \) nS, \( \sigma_e = 4.5 \) nS; reestimated conductances: \( g_{\text{exc}} = 2.2 \) nS, \( g_{\text{inh}} = 2.5 \) nS, \( \sigma_i = 0.94 \) nS, \( \sigma_e = 4.0 \) nS).
where $N$ denotes a normalization constant such that $\int_{-\infty}^{\infty} dV \rho(V) = 1$. The matching of this analytic solution and the numerical simulations are illustrated in Fig. 1E (dashed lines; see Rudolph and Destexhe 2003b for details).

**Gaussian approximation of the steady-state membrane potential distribution**

Because of the multiplicative coupling of the stochastic conductances to the membrane potential in $I_{syn}$, the membrane potential probability distribution (Eq. A4) takes in general an asymmetric form. However, $\rho(V)$ shows only small deviations from a Gaussian distribution, suggesting an approximation by a symmetric distribution. To this end, the exponent in Eq. A4 was replaced by the 2 first-order terms of its Taylor expansion around the maximum $V$ of the probability distribution $\rho(V)$

$$\rho(V) = \frac{1}{\sqrt{2\pi \sigma^2}} \exp \left( - \frac{(V - \bar{V})^2}{2\sigma^2} \right)$$

with $S_0 = k_L E_L + k_E E_E + u_i u_f + u_i E_E + u_f E_f + 2aC_m\tau_{mem}$ this yields the following Gaussian distribution

$$\rho(V) = \frac{1}{\sqrt{2\pi \sigma^2}} \exp \left( - \frac{(V - \bar{V})^2}{2\sigma^2} \right)$$

with the SD given by

$$\sigma^2 = \frac{S_0^2(u_i u_f + u_i E_E) - 2S_0(u_i E_E + u_i E_f) + S_0(u_i u_f)}{S_0}$$

Using 2 levels of (constant) injected current $I_{syn}$, these relations can be inverted (see Eqs. 11 and 12 and details in text), yielding estimates for the mean and variance of synaptic conductances.

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