Dynamic Synchronization of Purkinje Cell Simple Spikes

Soon-Lim Shin and Erik De Schutter
Theoretical Neurobiology, University of Antwerp, Antwerp, Belgium

Submitted 30 May 2006; accepted in final form 13 September 2006

Shin, Soon-Lim and Erik De Schutter. Dynamic synchronization of Purkinje cell simple spikes. J Neurophysiol 96: 3485–3491, 2006. First published September 20, 2006; doi:10.1152/jn.00570.2006. Purkinje cells (PCs) integrate all computations performed in the cerebellar cortex to inhibit neurons in the deep cerebellar nuclei (DCN). Simple spikes recorded in vivo from pairs of PCs separated by <100 μm are known to be synchronized with a sharp peak riding on a broad peak, but the significance of this finding is unclear. We show that the sharp peak consists exclusively of simple spikes associated with pauses in firing. The broader, less precise peak was caused by firing-rate co-modulation of faster firing spikes. About 13% of all pauses were synchronized, and these pauses had a median duration of 20 ms. As in vitro studies have reported that synchronous pauses can reliably trigger spikes in DCN neurons, we suggest that the subgroup of spikes causing the sharp peak is important for precise temporal coding in the cerebellum.

INTRODUCTION

Purkinje cells (PCs) generate two types of spikes, simple spikes (SSs) and complex spikes (CSs). High-frequency SSs are generally assumed to be driven by parallel fibers (PFs) (but see Loewenstein et al. 2005), whereas CSs are generated by climbing fibers (CFs) originating from the inferior olive (Ito 1984). Despite the large overlap of PF inputs impinging on PCs lying along the same PF beam (Solinas et al. 2003), SSs do not show any precise synchronization in PCs separated by >100 μm (Ebner and Bloedel 1981a; Jaeger 2003). Conversely, precise synchronization of SSs has been reported in pairs of adjacent PCs, either on the same electrode or on two electrodes separated by <100 μm (Bell and Grimm 1969; De Zeeuw et al. 1997; Ebner and Bloedel 1981a; Schwarz and Welsh 2001). Many reported synchronous pairs showed a sharp peak riding on a broader peak in the cross-correlogram, but whether these two peaks represent different sources of spikes has not been investigated. In this study, we could separate the mechanisms responsible for each peak.

The inhibitory PCs are not final projecting neurons of the cerebellum but target the real output neurons in the deep cerebellar nuclei (DCN). In vitro dynamic voltage-clamp experiments showed that ~10% synchronization among PCs converging onto a DCN neuron increases the reliability of the neuronal spiking four times more than 1% synchronization (Gauc and Jaeger 2000). Furthermore, DCN spikes can be precisely controlled by synaptic input at the millisecond level, and such spikes mostly followed short decreases in inhibition lasting ≥15 ms (Gauc and Jaeger 2000). However, it is not known whether such short pauses in PC SS trains are synchronized in vivo. Pauses in SS trains can be generated by multiple mechanisms. It has been reported that CSs are always followed by a pause of variable duration: from several tens of milliseconds (Bloedel and Roberts 1971; Granit and Phillips 1956) to several hundreds of milliseconds (Bell and Grimm 1969; Burg and Rubia 1972; Latham and Paul 1971; McDevitt et al. 1982; Murphy and Sabah 1970, 1971; Thach 1967). The CS triggered pauses can be followed by increased, decreased, or unchanged SS firing rates (McDevitt et al. 1982; Sato et al. 1992). On the other hand, pauses can also be generated by intrinsic afterhyperpolarization, decreased PF input, and/or increased on beam or lateral inhibition from interneurons (Cohen and Yarom 2000). Nevertheless, the origin of all these pauses would be indistinguishable in spike trains recorded from PC axons contacting DCN neurons (Monsivais et al. 2005). Hence we have defined pauses in this study as all SS ISIs that are indistinguishable in duration from the pauses after CSs, as the latter are uniquely defined. Applying this criterion, we have investigated the synchronization of SS pauses by analyzing the cross-correlations of nearby synchronous pairs of PCs.

METHODS

Recording

Forty five male Sprague-Dawley rats (300–400 g, Iffa Credo, Brussels) were anesthetized with a mixture of ketamine HCl (75 mg/kg); Ketalar, Parke-Davis, Warner Lambert Manufacturing, Dublin), and xylazine HCl (3.9 mg/kg; Rompun, Bayer, Leverkusen, Germany) in normal saline (0.9% NaCl, Baxter, Lessine, Belgium) by intraperitoneal injection. A craniotomy exposing Crus II of left cerebellar hemisphere was performed (Vos et al. 1999b). Supplemental doses of the anesthetic (one third initial dose) were given intramuscularly to maintain deep anesthesia as evidenced by the lack of a pinch withdrawal reflex and/or lack of whisking.

A new protocol was developed to record from 38 nearby PCs. One electrode was inserted perpendicularly to the cerebellar surface. The second electrode was tilted 58° from a horizontal plane to reduce dimpling and inserted at a distance from the first electrode so that their tips converged at the Purkinje cell layer. As control, we also recorded 12 distant pairs that were recorded from two electrodes separated by ≥400 μm. In all cases, the two electrodes were positioned along the transverse direction relative to each other. Here we describe data from eight selected pairs of nearby PCs (see RESULTS) and control analysis based on 39 single-unit recordings of PCs. After recording spontaneous activity in four synchronous pairs, tactile stimulus-evoked responses were recorded. Perioral receptive fields were explored as reported elsewhere (Vos et al. 1999b). A punctate stimulus was applied at 0.5 Hz. Electric lesions (15 μA, 8 s, cathodal DC current) were made to mark the location of the electrodes at the end of five paired recordings of nearby PCs. The distance between pairs was estimated from the distance between the centers of the lesion sites.

The costs of publication of this article were defrayed in part by the payment of page charges. The article must therefore be hereby marked “advertisement” in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.
Rats received a lethal dose of sodium pentobarbital (120 mg/kg ip; Nembutal, SANTOFI, Libourne, France) at the end of the experiment. The brain was removed and fixed in 4% paraformaldehyde. The cerebellum was embedded in paraffin, and 10-μm coronal sections were cut and stained with cresylviolet to visualize the electrolytic lesions. All experimental methods were approved by the ethical commission of the University of Antwerp and conformed to the guidelines of the European Commission.

Data analysis

After recordings of CSs and SSs, they were separated with off-line spike sorting software (Plexon, Austin, TX). Examples of the waveforms of sorted SSs and CSs from one PC are shown in Fig. 1A, inset. The off-line analysis of PC firing patterns was carried out using Matlab (The Mathworks, Natick, MA) and Microsoft Excel (Microsoft, Redmond, WA). The mean firing rates of SSs and CSs were 41.8 ± 3.6 and 0.73 ± 0.05 (SE) Hz, respectively, and they were not correlated (R² = 0.003). SS trains were constructed by removing all CS spikes from all recordings; any spike occurring within 10 ms of the start of a CS was considered to be part of the CS response as they probably belonged to the CS burst (Monsivais et al. 2005).

The strength of synchrony of SSs between pairs of PCs was measured using a standard score, the Z score of the cross-correlogram (Vos et al. 1999a) (bin = 2 ms). \[ Z = \left( \frac{N_c - N_e}{S_D} \right) \], with \( N_c \) the number of spikes of the central bin, \( N_e \) the mean number of spikes per bin computed in a 2-s window between −1 and 1 s, and \( S_D \) the SD of spikes per bin computed in the same window. Only pairs with \( Z \) scores of ≥3 were considered as synchronized pairs. The broadness of synchronization was measured with the full width at half-height (half-width). To assess the effect of firing rate co-modulation, shuffled spike trains were generated by randomly shuffling the ISIs within blocks of five consecutive ISIs. All P values refer to Student’s paired or unpaired t-test, unless otherwise specified.

Pauses were defined by setting a threshold for ISI duration, any ISI equal or larger than the threshold was considered a pause. A threshold was chosen based on previous reports that CSs are followed by pauses. The distribution of the first SS ISIs preceding CSs was compared with that of pauses after CSs, where the SS ISIs and pauses after CSs were pooled from 39 PC SS trains (Fig. 1A). The median, 10 percentile and 90 percentile of SS ISIs (pauses after CSs) were 10.0 ms (CS: 31.0 ms), 5.9 ms (CS: 17.9 ms), and 29.9 ms (CS: 617.5 ms) respectively. We then selected a threshold that would optimally detect the pauses after CSs if the two distributions were mixed as is the case in the PC axon because CS bursts are not transmitted faithfully (Monsivais et al. 2005). This was achieved by computing the proportion of ISIs categorized as pauses for different thresholds (set at 10, 11, 12, . . . , 20, 30, 50, 100, 200, or 1,000 ms) for all first SS ISIs preceding CSs and for all pauses after a CS (Fig. 1B, inset). The value that maximized the difference between these two proportions was considered as the optimal threshold, 16 ms (Fig. 1B). The smallest threshold which was not significantly different from the optimal, 12 ms, was used as threshold in this study. This threshold included more pauses caused by CSs, resulting in correct detection of 99.3 ± 0.2% of the pauses after CS (comparing with 95.4 ± 1.2% for a 16-ms threshold).

Using this definition of pauses, spikes were classified as either pause times (Pt, spikes beginning or ending a pause) or nonpause times (Nt, the rest of the spikes; Fig. 2A). The sensitivity of Z score and HW to the pause threshold was tested by computing these values for different thresholds. Measurements from each threshold were compared with those measured in overall spikes with the Wilcoxon signed-rank test. To avoid bias caused by small sample sizes, we excluded all recordings with a sample size <200. The sample sizes of Pt and Nt at different thresholds are summarized in supplemental Table 1.1

Responses to punctate tactile stimulation were measured by estimating the firing rates in 200 ms windows before and after stimulation. The significance of responses was determined by comparing numbers of spikes per bin after stimulation with averaged number of spikes per bin over the 100 ms window before stimulation in the peri-stimulus time histogram (bin = 20 ms).

RESULTS

In total, 21 stable recordings of transverse PC pairs were obtained. In 3 of 21 pairs a comparison of their auto- and cross-correlograms indicated that the pairs were probably recorded from the same PC (supplemental Fig. 1). Of the 18...
remaining pairs, 13 pairs showed statistically significant central peaks in their cross-correlograms (Z scores for the amplitude of the central peak of ≥3) (Vos et al. 1999a), which were not obtained in any distant pairs (3 pairs: 2 < Z score < 3; 9 pairs: Z score < 1). The rest of five adjacent pairs showed Z scores of either between 2 and 3 (n = 3) or <1 (n = 2). For the rest of this study, the eight pairs with Z scores >4 were further analyzed. They were separated by 69.8 ± 9.4 μm (range: 50–100 μm, Fig. 3A) and revealed a tight synchronization (Z score 8.2 ± 0.9; half-width 17.5 ± 7.9 ms; Fig. 3, B and C, black), which was not correlated to differences in firing rates between the two PCs. On average 34.6 ± 2.1% (range: 19.5–49.6%) of SSs were precisely synchronized (central window of ±2 ms). In all cases, the peak on the cross-correlograms consisted of a sharp peak (precise synchronization) riding on top of a broader central peak (loose synchronization) as has been reported for nearby PCs in previous studies (Bell and Grimm 1969; Ebner and Bloedel 1981b). Furthermore, spontaneously synchronous pairs responded to sensory tactile stimulation by significantly increasing their firing rate after stimulation from 49.5 ± 7.6 to 71.5 ± 7.1 Hz (n = 8, P < 0.02, Wilcoxon signed-rank test; Fig. 4B). However, their strength of synchronization (Z score = 8.1 ± 1.2) was not significantly different from that for spontaneous activity (Z score = 7.9 ± 1.3, P > 0.99, Wilcoxon signed-rank test).

To study if there is any difference in the precision of synchronization among SSs depending on whether they are related to pauses or not, we developed a spike separation method based on the ISI duration. This method allowed SS intervals to be categorized into pause (P) and nonpause (N) ISIs by a threshold of 12 ms (see METHODS and Fig. 1). In the eight PC pairs, on average 52.5 ± 4.5% of ISIs were pauses (Fig. 2B, top), which was not different from single-cell recordings of the other 10 PC pairs (P > 0.7). The proportion of pauses in each SS train was quite different from PC to PC, resulting in a wide distribution (Fig. 2B, bottom). The correlation between the proportion of pauses for each PC of a pair was not high (correlation coefficient of 0.54), this wide distribution seems to be due to genuine differences between the recorded PCs as opposed to differences between rats or experiments.

Using the definition of pauses, spikes were classified as either pause times (Pt) or nonpause times (Nt; METHODS and Fig. 2A). On average, 73.2 ± 4.1% of spikes were classified as Pt (Fig. 2C). This classification was used to recompute the cross-correlogram for either all Pt (Fig. 3, B and C, red) or all Nt (green), taken from both cells of the pairs. The sharp peak of the cross-correlogram consisted exclusively of Pt (sample size: 8763 ± 714, Z score 8.1 ± 1.3, range: 3.9–15.9; half-width 4.0 ± 0.7 ms, range: 2–8 ms), whereas the Nt caused the broad peak (sample size: 3,859 ± 840, Z score 5.3 ± 0.6, range: 3.1–7.2; half-width 60.3 ± 10.0 ms, range: 36–110 ms). These observations were robust for the threshold selected (Fig. 3C). For thresholds between 12 and 18 ms, Pt had narrower half-widths and higher Z score than Nt did (P < 0.03, Wilcoxon signed-rank test). Conversely, smaller thresholds gradually destroyed the difference in precision between Pt and Nt (Fig. 3C). The higher synchronization of Pt was not caused by the different sample size as it persisted when the sample sizes for both Pt and Nt were equivalent (threshold of 16 ms, supplemental Table 1).

These results indicate that mainly the spikes around pauses were precisely synchronized. Conversely, two facts suggest that the broad peak consisting of Nt spikes was caused by firing rate co-modulation, a mechanism that causes spurious synchronization between two neurons which have firing rates that co-vary in time (Eggermont and Smith 1995; Maex et al. 2000). First, in properly shuffled spike trains the broad peak was conserved for Nt but not for Pt (Fig. 3B, left inset). Second, precision of synchronization correlated with firing rate as Nt occurring in the central 2-ms window of the broad peak...
the first SS after a CS was removed for all CSs in one of the SS synchronization observed. The correlation of SS trains where investigated whether pauses after CSs were responsible for the having a preferential timing relative to the pause. Next, we test), indicating that the accurately synchronized spikes did not compared with each other (Fig. 6, cyan dotted lines).

Our analysis thus suggests that Pt or Nt of one PC tended to be correlated with corresponding spikes of the other PC. Pt can be correlated with Nt of one PC with the corresponding spikes of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

Our analysis thus suggests that Pt or Nt of one PC tended to be correlated with corresponding spikes of the other PC. Pt can be correlated with Nt of one PC with the corresponding spikes of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

Our analysis thus suggests that Pt or Nt of one PC tended to be correlated with corresponding spikes of the other PC. Pt can be correlated with Nt of one PC with the corresponding spikes of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

Our analysis thus suggests that Pt or Nt of one PC tended to be correlated with corresponding spikes of the other PC. Pt can be correlated with Nt of one PC with the corresponding spikes of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

Our analysis thus suggests that Pt or Nt of one PC tended to be correlated with corresponding spikes of the other PC. Pt can be correlated with Nt of one PC with the corresponding spikes of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

Our analysis thus suggests that Pt or Nt of one PC tended to be correlated with corresponding spikes of the other PC. Pt can be correlated with Nt of one PC with the corresponding spikes of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

Our analysis thus suggests that Pt or Nt of one PC tended to be correlated with corresponding spikes of the other PC. Pt can be correlated with Nt of one PC with the corresponding spikes of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

Our analysis thus suggests that Pt or Nt of one PC tended to be correlated with corresponding spikes of the other PC. Pt can be correlated with Nt of one PC with the corresponding spikes of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

As a control, we correlated the Pt of one PC with the Nt of the other PC. These spikes showed either no synchronization or a small negative correlation and therefore did not make any contribution to the short- or long-duration correlation of the overall cross-correlogram (Fig. 3B, right inset, magenta and cyan dotted lines).

Our analysis thus suggests that Pt or Nt of one PC tended to be correlated with corresponding spikes of the other PC. Pt can be correlated with Nt of one PC with the corresponding spikes of the other PC. These specific correlations did not show significant differences compared with each other (Fig. 6, P > 0.4, Wilcoxon signed-rank test), indicating that the accurately synchronized spikes did not have a preferential timing relative to the pause. Next, we investigated whether pauses after CSs were responsible for the synchronization observed. The correlation of SS trains where the first SS after a CS was removed for all CSs in one of the SS trains (n = 4, mean CS firing rate = 0.9 ± 0.1) did not show any significant changes in the mean firing rates of Pt between 200-ms windows before and after stimulation, 34.3 ± 3.8 to 37.0 ± 2.8 Hz, respectively (P > 0.6, Wilcoxon signed-rank test), suggesting that the precise synchronization of Pt was not caused by CSs.

We analyzed whether tactile stimulation changed the proportion of Pt in the SS trains by constructing peristimulus time histograms using only Pt (Fig. 4C, red and blue). On average, there was no significant difference in the mean firing rates of Pt between 200-ms windows before and after stimulation, 34.3 ± 3.8 to 37.0 ± 2.8 Hz, respectively (P > 0.6, Wilcoxon signed-rank test). However, this average reflects a large variability in responses between different PCs, which could decrease (Fig. 4C, left) or increase (Fig. 4C, right) their proportion of pauses during the response to the stimulus.

The analysis presented in Figs. 3, 4C, and 6 was based on cross-correlation of specific categories of spikes. This does not guarantee that pauses were highly synchronized because a spike at the start of a pause in one PC can be synchronous with a spike at the end of a pause in the other PC. Therefore in a final analysis, we selected all Pt synchronous within ±2 ms and checked whether the corresponding pauses where also synchronized. The pauses starting together (S-sP) represented 4.0% of overall pauses. Similar proportions were found for pauses ending together (S-eP, 3.6%) or followed by another pause (S-iP, 5.4%; Fig. 7A). If this synchrony was caused by chance, these proportions would be similar to the values obtained from randomly shuffled spike trains. However, all cases were significantly more synchronous than chance level (P < 0.002, Wilcoxon signed-rank test). Interestingly, the synchronous pauses (SM-P, the sum of S-sP, S-eP, and S-iP) had a significantly longer duration (median: 19.9 ± 0.6 ms) than the averaged duration of all pauses (Fig. 7B, P < 0.03, Wilcoxon signed-rank test). But, the durations of SM-P were

![FIG. 5. Synchronous nonpause ISIs. The median durations of all nonpause ISIs (allN) compared with that of synchronous nonpause ISIs (SM-N) and that of all other nonpause ISIs (oN). SM-N was significantly shorter (P < 0.001, Wilcoxon signed-rank test), while oN was not different from allN (P > 0.8).](http://jn.physiology.org/figures/96/6/3488/f1_06.jpg)
Synchronization of Pauses

...spikes are precisely synchronized and decreased after stimulation was strongly related to how the stimulation (Fig. 4) were found only during limited time windows, mostly just after 0.04).

...times starting (----), ending (-----), or surrounded by (---) pauses in a representative PC pair. Insert: same but superimposed plots. B: mean ± SE of Z scores in 8 pairs. P > 0.4 (Wilcoxon signed-rank test).

Significant changes of synchronous pauses after stimulation were found only during limited time windows, mostly just after stimulation (Fig. 4C, green). Whether SM-P increased or decreased after stimulation was strongly related to how the proportion of Pt changed in the PCs (supplemental Fig. 2, P < 0.05, Student’s t-test), indicating that the overall level of synchronization of pauses was not affected by tactile stimulation. Indeed, over longer time scales (500 ms before/after stimulus) we did not find any significant change in the proportion of synchronous pauses (before stimulation: 14.0 ± 1.5%, after stimulation: 12.7 ± 1.7%, P > 0.09, Wilcoxon signed-rank test). As the CS response after stimulation showed a different temporal profile (Fig. 4C, cyan and magenta), it is unlikely that SM-Ps during the response were caused by CSs. This is also supported by the flat cross-correlogram between CSs and SM-Ps (supplemental Fig. 3).

Taken together, in this study, we have shown that around half of the pauses are synchronized precisely, either starting or ending precisely together; CSs do not cause the precise synchronization of pauses; and coincident pauses are longer than other pauses, but their durations are not correlated.

Discussion

The cerebellum plays an important role in motor coordination by controlling the relative timing of muscle activation (Berardelli et al. 1996) and also in other tasks requiring the precise representation of temporal information (Ivry and Spencer 2004; Koekkoek et al. 2003). Although PCs can generate accurately timed spikes (Mittmann et al. 2005), this accuracy has to be preserved at the level of targeted DCN neurons. In vitro dynamic current-clamping experiments have demonstrated that synchronized pauses in firing of converging PCs can induce DCN neurons, to spike in a reliable fashion (Gauck and Jaeger 2000). This makes DCN neurons quite different from cerebral cortical neurons, the precise spike timing of which is controlled by increased depolarization (Mainen and Sejnowski 1995). Here, we have shown that ~13% of all pauses are synchronized in nearby PC pairs in vivo. Synchronized pauses had a median ISI of ~20 ms which is longer than the minimum duration of 15 ms reported by Gauck et al. (2000) to reliably evoke a DCN spike, it is also similar to the duration of the IPSC recorded in DCN neurons in vitro (Pedroarena and Schwarz 2003; Telgkamp and Raman 2002).

If we assume that the recorded PC pairs are random samples of the ~1,000 PCs converging onto a DCN neuron in rats (Chan-Palay 1977), we expect that statistically a similar percentage of pauses and spikes will be synchronized among PCs converging on a DCN. The DCN spikes generated during the decrease of inhibition caused by synchronous pauses are most probably postinhibitory rebound spikes (Llinas and Muhlethaler 1988). Other in vitro experiments have shown that DCN neurons generate pronounced postinhibitory rebound spikes with a delay of ~30 ms to the onset of the last inhibitory postsynaptic potential (IPSP) (Aizenman and Linden 1999). Such rebound spikes, which rely on the strength and duration of preceding hyperpolarization (Aizenman and Linden 1999), are a good candidate cerebellar timing signal (Kistler and De Zeeuw 2002; Koekkoek et al. 2005).

What causes the precise synchronization of pauses in nearby pairs of PCs receiving common PF input remains unclear, but...
we could exclude an important contribution of CSs. It has recently been suggested that CFs may evoke transitions between up and down states in PCs in vivo (Loewenstein et al. 2005; but see also Schoneville et al. 2006). It is unlikely that the synchronized pauses we observed correspond to quiescent down states because the latter last on average several seconds (Loewenstein et al. 2005). Moreover we did not find any evidence for a causal relationship between CF occurrence and transitions between high and low firing rate regimes (S. L. Shin, F. E. Hoebeek, M. Schonewille, A. Aertsen, and E. De Schutter, unpublished data). Indirect effects of CFs on molecular layer interneurons or of mossy fiber branching are also unlikely as these are expected to affect PC synchronization over much longer distances (Shinoda et al. 2000; Welsh et al. 1995; Woolston et al. 1981) than observed here.

A more likely mechanism for the pauses is synchronized IPSPs caused by feed-forward inhibition, which is an important regulator of the timing of the neuronal responses of cerebellar PCs and molecular layer interneurons (MLIs) (Cohen and Yarom 1998; Eccles et al. 1967; Mittmann et al. 2005; Solinas et al. 2006). The role of common feed-forward inhibition in the precise synchronization is supported by the facts that the sharp synchrony has been observed only in very close pairs (Jaeger 2003) and that interneurons have short axonal collaterals (Paley and Chan-Palay 1974; Solinas et al. 2003; Sultan and Bower 1998). However, feed-forward inhibition is more likely responsible for the synchronization of pauses, not for the synchronous spikes that may have been generated by parallel fiber inputs. As not all excitatory PF inputs trigger feed-forward inhibition (Mittmann et al. 2005), carefully designed follow-up studies will be required to understand their exact contribution to the observed synchrony. Another possible cause of the synchronous pauses is intrinsic afterhyperpolarization, which may be modified by learning processes (V. Steuber, W. Mittmann, M. Häusser, and E. De Schutter, unpublished results). An intrinsic origin of pauses could explain the large variability between PCs (Fig. 2B, bottom) as PCs show a large variability in the expression of voltage gated channels (Achard and De Schutter 2006; Swensen and Bean 2005). Although CSs were not directly responsible for the synchronous pauses, this does not exclude that previous learning processes induced by Cs modified dendritic excitability (Schreurs et al. 1998).

Not only precisely synchronized Pt but also the loosely synchronized Nt may be important for controlling DCN neurons. The latter may regulate DCN baseline firing rates (S. L. Shin, F. E. Hoebeek, M. Schonewille, A. Aertsen, and E. De Schutter, unpublished data), which are linearly correlated to the input frequencies of converging PCs (Gaucck and Jaeger 2000). In this study we have shown that fast firing spikes tend to be only loosely synchronized. This will raise the actual firing rate of the converged high-frequency input, resulting in more efficient suppression of the DCN neurons.

Our estimate of 35 and 13% of synchronization of simple spikes and pauses, respectively, were made during spontaneous activity under anesthesia. Although it was recently reported that there was no difference in the occurrence of pauses of 50 ms or longer between anesthetized and awake mice (Schonewille et al. 2006), we do not know whether this also applies to synchrony of pauses. We did not find a strong effect of tactile stimulation, but this may have been due to the lack of a behavioral context in the anesthetized preparation (Krupa et al. 2004). Indeed, similar spike correlations with different degrees of temporal precision were observed in motor cortex of behaving monkeys (Riehle et al. 1997). In the motor cortex, precise synchronization without rate modulation was observed during internal (expectation) events. To understand the functional significance of dynamic synchronization in cerebellar PCs experiments in awake, behaving animals will be needed, but under these conditions, recording from nearby PCs on different electrodes will be a technical challenge. Nevertheless, at this time we can propose that the dynamic synchronization of PC SSs may be important for fine control of the target neurons in DCN neurons and that, especially, the precisely synchronized pauses may serve to control the timely generation of rebound spikes in DCN neurons.

Acknowledgments

We thank E. De Leenheer and I. Bats for the histology and the photography. We gratefully acknowledge Drs. Reinoud Maex and Pablo Achard for helpful comments on an earlier version of the manuscript.

Grants

This work was supported by grants from University of Antwerp, Inter University Attraction Pole, Queen Elisabeth Medical Foundation and Fund for Scientific Research (Flanders).

References


J Neurophysiol • Vol. 96 • December 2006 • www.jn.org