Computational diversity in the cochlear nucleus angularis of the barn owl

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Abbreviated Title: Computational diversity in nucleus angularis

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Keywords:
Auditory, Bird, Sound level, Brainstem, Evolution

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ABSTRACT

The cochlear nucleus angularis (NA) is widely assumed to form the starting point of a brainstem pathway for processing sound intensity in birds. Details of its function are unclear, however, and its evolutionary origin and relationship to the mammalian cochlear-nucleus complex are obscure. We have carried out extracellular single-unit recordings in the NA of ketamine-anaesthetized barn owls. The aim was to re-evaluate the extent of heterogeneity in NA physiology, since recent studies of cellular morphology had established several distinct types. Extensive characterization, using tuning curves, phase locking, peri-stimulus time histograms and rate-level functions for pure tones and noise, revealed 5 major response types. The most common one was a primary-like pattern that was distinguished from auditory-nerve fibres by showing lower vector strengths of phase locking and/or lower spontaneous rates. Two types of chopper responses were found (chopper-transient and a rare chopper-sustained), as well as onset units. Finally, we routinely encountered a complex response type with a pronounced inhibitory component, similar to the mammalian type IV. Evidence is presented that this range of response types is representative for birds and that earlier conflicting reports may be due to methodological differences.

All 5 response types defined were similar to well-known types in the mammalian cochlear nucleus. This suggests convergent evolution of neurones specialized for encoding different, behaviourally-relevant features of the auditory stimulus. It remains to be investigated whether the different response types correlate with morphological types and whether they establish different processing streams in the auditory brainstem of birds.
INTRODUCTION

The cochlear nucleus is the first brainstem nucleus of the auditory pathway and, in birds, is subdivided into the Nucleus magnocellularis (NM) and the Nucleus angularis (NA), both of which are contacted by collaterals of the same auditory-nerve fibres (review in Carr and Code 2000). Most studies of the avian cochlear nucleus have concentrated on the NM. It is widely accepted that the bird NM is the equivalent of the spherical bushy cell population in the mammalian anteroventral cochlear nucleus. This is supported by many detailed similarities in both anatomy and physiology and the specialized role of that particular cell type in temporal auditory processing for sound localization (e.g. reviews in Carr et al. 2001; Oertel 1999; Trussell 1999). Yet the auditory system does much more than localize sources via interaural time differences; it enables localization using level differences and spectral cues, and a whole range of sound recognition behaviour (e.g. Dooling et al. 2000). With extreme specialization for temporal coding in NM, we look to the other nucleus of the pair, NA, as a likely candidate to lay the foundations for these other tasks.

Studies on the cellular morphology of NA agree that it is a heterogeneous nucleus with several different neurone types (Boord and Rasmussen 1963; Häusler et al. 1999; Soares and Carr 2001). Soares et al. (2002) have recently used in vitro whole-cell recording to characterize the intrinsic firing properties and single-cell morphology of NA in the chicken. They showed several distinct physiological classes of cells. Their physiological types correlated with cellular morphology and the morphological types were similar to those found in the owl. These results would predict diversity in NA in vivo responses as well, however, previous recordings in the NA provided conflicting reports on the response types present and indicated an unusual species specificity at such an early stage of auditory processing. For example, responses in the barn owl were reported to be very uniform (Sullivan 1985; Sullivan and Konishi 1984), while both more varied and more complex responses had been found in the redwing blackbird (Sachs and Sinnott 1978). The chicken, while also varied, showed yet other response categories (Warchol and Dallos 1990). In the light of our recent findings on the organization and cell types in NA, we have therefore reexamined the in vivo physiology in the barn owl NA.

MATERIAL AND METHODS

Animals, anaesthesia and homeostasis:

Results are reported from experiments on 6 barn owls (Tyto alba pratincola), 5 males aged between 1 and 13 years and one 6 months old individual of unknown sex. Most animals were used in 2 to 3 separate experiments, spaced several days apart. Anaesthesia was induced by intramuscular injections of 10-14 mg/kg ketamine hydrochloride (“Ketavet” Phoenix, St. Joseph, MO) and 2-3 mg/kg xylazine (“xyla-ject”, Phoenix); supplementary doses of ketamine and xylazine were administered according to individual needs (on average approx. 7 mg/kg/h ketamine and 1.5 mg/kg/h xylazine). Body temperature was continually measured by a probe inserted into the owl’s cloaca and kept constant at 39ºC by a feedback-controlled heating blanket wrapped around the owl’s body (Harvard Instruments, Braintree MA). An EKG was recorded by
fine needle electrodes inserted into a leg muscle and a muscle of the contralateral wing; this was periodically displayed on an oscilloscope and/or broadcast on an audiomonitor, to check for muscle potentials (associated with breathing or waning anaesthesia) and frequency and regularity of the heart beat. At the end of experiments after which the animal was allowed to recover, about 0.06 mg/kg buprenorphine hydrochloride (“Buprenex” by Reckitt & Colman Products Ltd., Hill, England) was administered.

**Surgery and stereotaxis:**

The owl’s head was firmly held in a controlled position by a custom-designed setup, using earbars and a beak holder. After removing some feathers, cutting the skin and gently roughing the skull surface, a metal headplate, as well as a short metal pin marking a standardized zero point, were permanently glued to the skull. After this, the ear bars and beak holder were removed and the head held firmly by the headplate alone. Three different approaches were then used to place electrodes into the brainstem: a) stereotactically through the main part of the cerebellum, b) stereotactically through the cerebellar flocculus, and c) using landmarks after aspirating the cerebellum and exposing the brainstem (b and c were used in terminal experiments only). In all cases, after placing the electrodes on the brain surface, they were lowered by a custom-built stepping motor, controlled from outside the experimental chamber.

a) An opening was made in the skull around the desired area relative to the zero point and the dura mater was cut open, taking care to avoid blood vessels. Each electrode was individually centered over the zero point and moved defined amounts in the rostrocaudal and mediolateral axes, before being driven down into the brain. In addition, the electrode was slightly angled laterally in most penetrations. Electrode positions were re-zeroed after any change in angulation, thus keeping the entrance points into the cerebellum reproducible. The NA was found between 1 and 5 mm caudal to the zero point, with mediolateral coordinates dependent upon angulation, and being between 0.5 and 4 mm from zero.

b) A skull opening lateral to the cerebellum was made, providing a view onto the area of the semicircular canals. After removing the bone overlying the surface of the cerebellar flocculus, electrodes were introduced using various combinations of medial and rostrocaudal angles established in past experiments (Köppl et al. 1993).

c) A skull opening was made, exposing the caudal cerebellum on one side, leaving the midsagittal sinus covered, but exposing far laterally. After removing the dura mater, the accessible cerebellum was aspirated, exposing the auditory brainstem. Electrodes were placed under visual control, aiming for the very lateral edge of the brainstem just caudal to the cerebellar peduncle.

**Electrodes and recording setup:**

Owls were placed on a vibration-isolated table within a sound-attenuating chamber (IAC, New York) that was closed during all recordings. Commercial, Epoxylite coated tungsten electrodes (Frederick Haer Corporation, ME) were used, preferably with impedances around 15-20 MΩ. A grounded silver wire, placed under the animal’s skin around the incision, served as the reference. Electrode signals were amplified and filtered by a custom-built headstage and amplifier. The
recording was then passed in parallel to an oscilloscope, a threshold discriminator (Tucker-Davis Technologies (TDT; SD1) and an analog-to-digital converter (TDT DD1) connected to a personal computer via an optical interface (TDT OI). TTL pulses from the threshold discriminator were also registered by the personal computer via an additional timing module (TDT ET1), with a precision of 10µs. The TTL pulses were also fed to the z-axis of the oscilloscope displaying the recording trace, providing a visual aid for adjusting the TTL trigger level. In addition, a continuously refreshed, software-generated display of the waveforms that triggered TTL-pulses aided in trigger judgement. Finally, a continuously refreshed display of the interspike interval distribution helped to judge unit isolation. For well-isolated units, there were no intervals shorter than about 0.6 ms, representing the absolute refractory period. In most cases, neural responses were only saved as TTL timing events. In two experiments, however, examples of analog waveforms were also saved. We did not check for prepotentials in the spike waveforms, assuming that these can only be expected in cells with large, endbulb-like terminals, which are not found in NA (e.g. Carr and Boudreau, 1991).

**Stimulus generation and calibration:**

Acoustic stimuli were digitally generated by custom-written software (“Xdphys” written in Dr. M. Konishi’s lab at Caltech) driving a signal-processing board (TDT DSP2). After passing a digital-to-analog converter (TDT DD1) and an anti-aliasing filter (TDT FT6-2), the signals were variably attenuated (TDT PA4), impedance-matched (TDT HB4) and attenuated by an additional fixed amount before being fed to commercial miniature earphones (SONY MDR-E424). Two separate channels of signals could be generated, passing through separate channels of all associated hardware and driving two separate earphones. The earphones were housed in custom-built, calibrated, closed sound systems, inserted into the owl’s left and right ear canal, respectively. Sound pressure levels were calibrated individually at the start of each experiment, using built-in miniature microphones (Knowles EM3068 or TM3568, in two different sets of sound systems used).

**Stimulus paradigms and unit characterization:**

While lowering the electrode, noise bursts (50 ms duration, 5 bursts/s) were played to the ipsilateral ear as search stimuli. Once auditory responses were discernable, different frequencies and both ipsi- and contralateral stimuli were tested to judge the position of the electrode. After isolating spikes, the characteristic frequency (CF) was estimated audiovisually and the TTL trigger level was adjusted carefully. The following protocol was then tested in full if possible, or until the unit was lost:

*Tuning curve or response map:* Tone bursts (50 ms total duration, 5 ms rise-fall time, 200 ms cycle) of different frequencies and levels were presented in random order. Levels varied in 5dB steps from below the initial threshold estimate to at least 60dB SPL, frequencies varied in 50-250 Hz steps (depending on CF, smaller steps for units of lower CFS). Average discharge rates over 3 repetitions of each stimulus, within the stimulus window (not corrected for neural latency) were calculated. This paradigm was chosen to derive iso-rate tuning curves comparable to those published for the auditory nerve of the barn owl (Köppl 1997a, see there for detailed methods). The rate criteria for excitatory and, if present, inhibitory threshold responses, were fixed
individually, depending on the spontaneous rate and variability. For most units, the criterion fell between 10 and 30 spikes/s above spontaneous rate.

**Test for phase locking:** Tone bursts (50 ms total duration, 5 ms rise-fall time, fixed starting phase, 200 ms cycle) at the estimated CF and a level at least 20 dB above threshold were presented 100 times. These parameters were chosen to estimate the maximal vector strength (VS) a unit could produce. Only statistically significant VS were accepted (Rayleigh test, p<0.01).

**Peri-stimulus-time histograms (PSTH), latency, regularity analysis:** Tone bursts (50 ms total duration, 1.6 ms rise/fall time, variable starting phase, 200 ms cycle) at the estimated CF, usually 20-30 dB above threshold, were presented 300 times. For stimuli below 1 kHz, rise/fall time was routinely changed to 5 ms, to minimize spectral splatter. Also, in some early experiments, stimulus rise/fall times of 5 ms and 500 or 100 repetitions were generally used. For many units, several levels of PSTH were recorded, usually 20 dB apart. PSTH with a standard bin width of 0.1 ms were used to judge the general shape of the PSTH and to derive the minimal response latency (first two consecutive bins to exceed the maximum encountered outside the response). In addition, first-spike analyses (after Young et al. 1988) were carried out, using the first spike in each run after the minimal latency. Interspike interval distributions for all spikes during the response were calculated and regularity analyses (after Young et al. 1988) were carried out, using a time window of 12-20 ms after the minimal response latency for the calculations of mean interspike interval, mean standard deviation and mean coefficient of variation (CV). Mean regularity values were only used if the mean discharge rate was at least 100 spikes/s. These parameters and procedures closely followed the standard methods of many mammalian studies.

**Rate-level functions, spontaneous rates:** Tone bursts (50 ms total duration, 5 ms rise-fall time, 330 ms cycle) of different frequencies and levels were presented in random order. Levels varied in 3 dB steps from below the CF threshold to at least 60 dB SPL and usually about 10 frequencies (surrounding and including CF) were tested. Average rates over 10 repetitions of each stimulus, within the stimulus window (not corrected for neural latency) were calculated. Spontaneous discharge rates were derived from no-stimulus trials also included randomly. These data provided more detailed response maps, as well as the rate-level function at CF, with parameters identical to those used on auditory-nerve fibres in the barn owl (Köppl and Yates 1999). Saturation discharge rates were estimated as the average rate within the saturated part of the rate-level function, as judged subjectively. If no saturation was apparent and the function covered at least 40 dB above threshold, the rate at the highest level tested was taken. In the case of non-monotonic rate-level functions (defined as showing a decline of more than 20% below the highest rate), the highest driven rate, irrespective of level, was taken. Dynamic ranges were only calculated for monotonic rate-level functions, as the range covering from 10% to 90% of the increase above spontaneous rate to saturation rate. The 10% and 90% points were estimated from linear regressions through adjacent data points.

In 20 units (including all response types), the rate-level function at CF was repeated for contralateral stimulation. In all those cases, the threshold to contralateral stimulation was higher than to ipsilateral stimulation and the difference was consistent with physical cross-talk between the middle ears under similar conditions (Köppl 1997a), i.e. consistent with exclusively monaural input.

**PSTH and rate-level function for noise stimuli:** These data were collected analogous to the paradigms described above for pure-tone stimuli, except that band-limited noise (200 Hz to 12
kHz; synthesized freshly for each run) was presented. The level of noise stimuli across frequencies was equalized online, i.e. all noise levels are given as spectrum levels.

**Electrolytic lesions and histology:**

At selected recording sites, small lesions were induced by passing pulsed, positive current of 1-4 µA for 10 s to 8 minutes through the electrode. After a survival time of several hours to 13 days, the owl was sacrificed by an anaesthetic overdose and perfused transcardially with saline, followed by fixative (various buffered aldehyde mixtures). The brain was dissected out and cryoprotected by incubation in 30% buffered sucrose until it sank. Frozen sections were cut in the same (approximately transverse) plane as the electrodes had penetrated. Sections were mounted on gelatine-coated slides, stained with cresyl violet and coverslipped. All sections surrounding and including NA were examined at low magnification, documenting the extent of NA and any damage or glial accumulation potentially associated with lesions.

**RESULTS**

**Verification of recording sites**

Electrodes were aimed stereotactically or using landmarks and the decision whether NA was actually penetrated in a given track was based on physiological criteria. The physiological responses of auditory areas adjacent to NA, i.e. the auditory nerve, NM and NL are well known (e.g. Köppl 1997a; Pena et al. 1996; Sullivan and Konishi 1984). In addition, the well-known tonotopic organizations of the different nuclear areas (Carr and Konishi 1990; Köppl 2001; Takahashi and Konishi 1988) served as a further guideline.

Six recording sites were marked by electrolytic lesions. Five of these were later identified in histological sections of NA (examples in Fig. 1). The 6th lesion could not be found, presumably because the animal did not, as intended, recover from the experiment and the survival time was too short to reveal the characteristic glial accumulation. One further track was approximately confirmed by an ink mark placed on the surface of the brainstem. The tonotopic position of lesion sites corresponded with the best frequencies estimated immediately before current application. The tracks where the 5 successful lesions were placed accounted for 12 of the units reported here; all of those were classified as NA and all response types (except the rare chop-S) were represented. Using the lesioned sites as calibration points and the stereotaxic coordinates of electrode penetrations in the same individual as guidelines, we could reconstruct the approximate rostrocaudal position for about 70% of all recorded units. These recordings covered the rostral two-thirds of NA. The remaining units either came from brainstem hemispheres without any anatomical confirmation of recording sites or from penetrations aimed using landmarks. In the mediolateral axis, a detailed reconstruction of anatomical position was not possible. In this axis, the nucleus is much narrower and electrode angulation introduced an additional variable which could not be measured and reproduced as precisely as the electrode’s X and Y positions. However, characteristic frequency changes along the mediolateral axis of NA in the barn owl (Köppl 2001), such that the lowest CFs can only be found at the most medial positions and the
highest CFs at the most lateral extreme. Judging by this criterion, we have sampled the entire mediolateral extent of NA (see data below).

In summary, all recording sites that could be confirmed anatomically were confirmed to have been within NA, corroborating our judgements based on physiological criteria during the experiment.

**Classification of response types**

Responses of a total of 76 single units recorded in the NA are reported here, covering a range of CFs from 0.45 to 10.2 kHz. The responses clearly were not uniform, but could be classified into several different categories. Since they appeared to be comparable to types previously defined in the mammalian cochlear nucleus, we have adhered to the established nomenclature. Table 1 summarizes the types, their relative abundance and several salient statistics of their response behaviour. Figure 2 presents the decision tree that was developed, using a range of criteria, including the traditional PSTH parameters. The most common response of NA neurones was of the primary-like variety, followed by chopper-transient (chop-T), typeIV, onset and, rarest, chopper-sustained (chop-S) responses.

Note that in addition to the sample of NA units, there are 26 units that were classified as “auditory nerve” or “uncertain” (Table 1). This reflects some difficulty in separating potential auditory-nerve responses from primary-like NA units. We assumed that our sharp tungsten electrodes were capable of recording from both cell bodies and large axons or dendrites (see e.g. Joris 1998) and that all of these could in principle be encountered anywhere within the nucleus. Therefore a conservative interpretation was used, classifying all units whose values for a list of parameters fell within the known range of auditory-nerve fibres (Köppl 1997a, b; Köppl and Yates 1999) as “auditory nerve”. Based on previous studies on the avian NA (Sachs and Sinnott 1978; Sullivan and Konishi 1984; Warchol and Dallos 1990) and our own auditory-nerve data from the barn owl (Köppl 1997a, b), a relatively low vector strength of phase locking and/or a relatively low spontaneous discharge rate were considered decisive parameters for separating NA units from auditory-nerve inputs. Note that no specific criterion values can be universally used for this distinction, since both parameters are strongly CF-dependent. For example, a unit with a CF of 5 kHz and a vector strength of 0.5 would have been classified as auditory nerve, whereas a unit showing the same vector strength at a CF of 2 kHz would have been classified as NA. Cases where both the vector strength and the spontaneous discharge rate were consistent with auditory-nerve responses, but other characteristics did not fit an auditory-nerve fibre (e.g. a high maximal discharge rate) were classified as “uncertain”. Latency was, unfortunately, of little use in unit classification (see below, after introducing the response types).

We will first describe and illustrate with examples the typical features of each NA response type. Then the different types will be compared and salient differences and similarities highlighted. It should be emphasized that most of the response types did not appear to be sharply distinguished, i.e. their characteristics overlapped to some degree for individual parameters. In addition, many parameters showed an overall CF-related variation that we have tried to separate and disregard in our classification of response types. A number of scatter plots will be shown in addition to individual examples to illustrate the range of responses and the extent of overlap between types.
Response type “Primary-like”

Figure 3 illustrates an example of a primary-like response. The characteristic feature of primary-like units was, of course, a primary-like PSTH, showing a vigorous discharge at stimulus onset that gradually adapted to a steady, lower discharge (Fig. 3C). Occasionally, the PSTH showed a clear notch after a peak at response onset. However, this was confined to high levels of 40dB or more above threshold and intermediate forms were seen. Also, such notches may sometimes be observed in auditory-nerve responses at high levels (own unpublished observations). Therefore, we do not feel there is evidence for a distinct response type “primary-like with notch” (however, see also “onset units” below). Interspike interval distributions of both spontaneous and evoked discharges were Poisson-like (Fig. 3F). Mean CVs were around 0.8-0.9 (median 0.84), except in low-frequency units where cycle-by-cycle phase-locking produced a more regular response.

All primary-like units with a CF below 5 kHz showed significant phase locking. Although their vector strengths were generally lower than those of auditory-nerve fibres of comparable CF, they could approach auditory-nerve values at CFs below 1.5 kHz (Fig. 4). Above 5 kHz, vector strengths were very low or not statistically significant. Spontaneous discharge rates were mostly below those of auditory-nerve fibres of comparable CF (Fig. 5A and Table 1).

Tuning curves of primary-like units were comparable to auditory-nerve tuning curves, showing a similar range of thresholds, Q10dB and Q40dB values. Four of 33 primary-like units showed evidence for side-band inhibition along the high-frequency flank of the excitatory tuning curve.

Rate-level functions of primary-like units were monotonic (Fig. 3B), with a median saturation discharge rate of 257.5 spikes/s and a median dynamic range of 30 dB (Table 1). Responses to noise were similar to pure-tone responses, with respect to PSTH (Fig. 3G), regularity of discharge and saturation rates.

Response type “Chopper-transient”

Figures 6 and 7 illustrate two examples of chop-T responses. Chop-T units showed a regular discharge pattern at stimulus onset that produced several distinct peaks in the PSTH (Fig. 6C, 7C). This did not depend on the stimulus rise time in 3 cases tested (rise time varied between 1 and 5 ms; example in Fig. 7C). Interspike intervals always increased within the first 10-20 ms of the stimulus (Fig. 6D, 7D); the CV mostly decreased, but could also remain steady or increase slightly (Fig. 6E, 7E). Mean CVs fell between 0.38 and 0.84 (median 0.53). At the upper end of this range, there was some overlap with primary-like units. Besides the characteristic peaks in the PSTH, chop-T units also typically did not show a Poisson-like distribution of interspike intervals in their sound-evoked discharge. Their distributions were more skewed, with an increased proportion of short intervals (Fig. 6F, 7F).

Spontaneous rates of chop-T units were typically low, with many units showing no spontaneous discharge at all (Fig. 5A and Table 1). Chop-T units were also characterized by inferior phase-locking. Although about half of them did show significant phase-locking, their vector strengths remained at the low end of those of primary-like units of comparable CF (Fig. 4).
Chop-T units had a median saturation discharge rate of 316 spikes/s and a median dynamic range of 23 dB, but showed large ranges for both parameters (Table 1). There was no correlation between saturation rate and dynamic range. Discharge rates and dynamic ranges in response to CF-tones and to noise, respectively, showed no systematic differences across the population. However, the PSTH of noise responses never showed clear chopping (Fig. 6G).

Chop-T units at the low end of the saturation discharge range showed PSTH with little sustained activity after the initial chopping peaks. Therefore we earlier contemplated a separate response type, “onset-chopper”. However, a more detailed analysis did not produce any evidence for this in the owl. Onset-choppers in cats and guinea pigs are characterized by their large dynamic range, small difference in threshold to tones and noise, and often more vigorous response to noise (Rhode and Smith, 1986; Winter and Palmer, 1995; Joris and Smith, 1998). While these individual characteristics were occasionally observed in our chop-T units (e.g. small threshold difference in Fig. 7B), none displayed all of them. Also, the PSTH in response to noise never showed chopping, while the PSTH of onset-choppers are very similar in response to tones and noise (Winter and Palmer, 1995).

An unexpected phenomenon was a decrease in spike size during medium to high-level stimulation, observed in about one quarter of all our chopper-type neurones (including chop-S, an example of which is shown in Fig. 8H, I). In early experiments, we may have rejected some units like these, mistaking the non-uniform spike size as indication for a multi-unit recording. However, the presence of a refractory period in the interspike-interval histogram confirmed that the spikes were from single units, despite the variable spike height.

**Response type “chopper-sustained”**

Chopper-sustained (chop-S) responses were rare in our sample and are only represented by two units (from different animals). One example is shown in Fig. 8. They had the longest response latencies and were clearly above all other unit types in this respect (Fig. 9). This was our main criterion for placing them in a category of their own, instead of interpreting them as one extreme of the chopper-type range.

Chop-S units showed the lowest mean CVs of all types and their CV remained constant throughout the stimulus duration (Fig. 8E). Also, in one case the unit chopped to both pure-tone and noise stimuli (Fig. 8 C and G; in the other case, the noise spectrum level was not sufficiently above threshold). Both chop-S units had no spontaneous activity and reached only moderate saturation discharge rates around 200 spikes/s. Their dynamic ranges were rather different, at 16 and 35 dB, respectively.

**Response type “onset”**

Units that showed only a single prominent, initial peak in their PSTH and no evidence for chopping, were classified as “onset”. A second characteristic was that the maximal discharge rate to noise was higher than that to tones at CF. In other aspects, this group was heterogeneous, however, considering the overall rarity of onset units and thus small sample size (n = 6), we refrain from further subdividing. Figures 10 and 11 illustrate the range of responses. The unit shown in Fig. 10 represents an extreme case of onset response, with virtually no spiking during
the remainder of the stimulus. The unit illustrated in Fig. 11 showed a robust sustained response after the onset and was in some aspects reminiscent of primary-like-with-notch responses in mammals. However, the distinction between onset and primary-like-with-notch is not always sharp in mammals as well (e.g. Rhode and Smith, 1986; Blackburn and Sachs, 1989). Our final criterion for classifying this (and another similar unit) among onset responses was the clearly more vigorous response to noise. The standard deviation of the first spike, a measure for the synchrony of the initial discharge, was extremely low for the unit shown in Fig. 10, but, as a population, actually larger in onset units than in all other types. However, this mainly reflects the fact that the onset spike occasionally failed, producing a distribution of first-spike times with extreme outliers and thus making standard deviation an inappropriate measure of dispersion. If an onset spike was fired, it was temporally precise, summing up, over many stimulus repetitions, to the characteristic sharp peak in the PSTH. The PSTH in response to noise was more primary-like in all cases tested, i.e. showed less synchronization to the stimulus onset (Fig. 10D, 11G). Onset units did not appear to phase-lock well (Fig. 4), however, our sample is small and restricted in CF.

**Response “Type IV”**

Eleven units were classified as type IV, with CFs between 0.6 and 7.7 kHz. Their defining characteristic was a pronounced non-monotonic behaviour of the discharge rate across different levels at CF, typically showing little to moderate excitation at low levels and inhibition at higher levels (Fig. 12B). The PSTH showed a clear onset response, followed by varying degrees of inhibition, depending on the unit and the stimulus level; we call this pattern onset-inhibitory. Units that responded with net excitation at low levels showed a sustained excitatory response at those levels, i.e. the PSTH changed with stimulus level (Fig. 12C-E, 13C-E). At levels near the transition between net excitation and net inhibition, the PSTH could also look pauser-like. When testing a range of frequencies and levels, complex response maps resulted, with interleaving excitatory and inhibitory areas (Fig. 12A). CF and threshold were defined in these cases as the most sensitive point, regardless of whether the response was net excitatory or inhibitory. All type IV units had high spontaneous discharge rates, typically above 100 spikes/s (Fig. 5A and Table 1).

Most type IV units were also tested with noise stimuli. The majority showed purely excitatory responses and a monotonic increase in discharge rate with level (Fig. 13B). This was accompanied by a primary-like pattern in the PSTH (Fig. 13F). Three units gave similar responses to noise and tones, i.e. a net inhibition at higher levels, with an onset-inhibitory PSTH (Fig. 12B, F).

Inhibition was seen to varying degrees in the different type IV units. At one extreme were units that showed little or no net inhibition, i.e. whose discharge rate never clearly decreased below the spontaneous rate (example in Fig. 13). At the other extreme were units that showed no clear excitation, i.e. whose only response appeared to be a net decrease of the discharge below spontaneous rate. However, the onset response in the PSTH was always present (example in Fig. 14). This was our final criterion for not placing such units into a (purely inhibitory) category of their own. It was also our impression that the responses recorded from type IV units had a snapshot character, meaning that the degree of inhibition seen in an individual neurone could change over time. Significant changes in the discharge behaviour over time could be documented...
for three single units. One example where inhibition became more pronounced with time is shown in Fig. 15. Stability or instability of the inhibitory response component did not appear to be related to the administration of anaesthetic agents.

**Comparison of types**

Our sample covered the full range of CFs expected for the barn owl, although only primary-like units were found throughout the whole range. Especially conspicuous was the absence of any type other than primary-like among the highest CFs, above 8 kHz. Chop-T units, although the second most frequent type, were also restricted at the low-frequency end and were only encountered at CFs between about 2 and 8 kHz. Anatomically, we did not observe any differences in where the various response types were found.

Perhaps surprisingly, latency was not clearly different between units classified as auditory nerve or NA, respectively, or between most of the NA response types. A number of variables influence latency, most importantly the CF and, using a fixed rise time, both the absolute and relative level (above threshold) of the stimulus (e.g. Heil and Neubauer 2001). It seemed that the scatter introduced by those variables largely obscured the small latency differences between auditory-nerve fibres and NA units (Fig. 9). The only exception with clearly different latencies were the chop-S units. Higher sound levels or clicks might have provided more consistent information, but unfortunately unit isolation could be compromised with such stimuli. We also evaluated mean and median first-spike latency, and these showed greater variability than the minimal latency shown in Fig. 9.

Except for the type IV, the different response types appeared to grade into each other. The extremes of the primary-like and chop-T responses, for example, were clearly different, however, a few units could only be classified by defining an arbitrary borderline value for the mean CV and subjectively classifying their PSTH shape. Similarly, chop-T units with low saturation discharge rates may form a continuum with onset units.

A number of parameters did not differ at all between most types of units. Tuning curves largely reflected the auditory-nerve inputs in terms of tuning and thresholds (Fig. 16). Only a minority of primary-like and chop-T units (4 of 33 and 2 of 17, respectively) showed evidence for off-CF inhibitory inputs along the high-frequency flank of their excitatory tuning curves. Type IV units tended to fall among the most sensitive thresholds, however, there was no statistically-significant difference in threshold between the response types (Kruskal-Wallis H-test, p = 0.06; only units with a CF below 8 kHz tested, to minimize the variation of threshold with CF). Dynamic ranges were not significantly different between auditory-nerve units and the different NA response types (Kruskal-Wallis H-test, p = 0.22; type IV units excluded, since no dynamic range was defined for non-monotonic rate-level functions). For the driven discharge range, i.e. the difference between spontaneous and saturation rate, significant differences were revealed. However, it was only the onset group that differed from all others (Kruskal-Wallis H-test with subsequent pairwise Mann-Whitney U-tests; type IV units excluded). This is probably entirely due to the low saturation discharge rates of onset units, which similarly differed from those of all other types (see also Table 1).
DISCUSSION

The two main findings of this study are that there are 5 main physiological types in the barn owl NA and that these show basic similarities to responses in the mammalian cochlear nucleus. Since our results appear to be at variance with previous studies in the owl and chicken, we will first attempt a new synthesis of NA physiology in birds, showing that technical issues are a major concern and that our results may in fact be representative for all birds. We then highlight the quantitative similarities in the response types of the barn owl and mammals and discuss why these similarities are remarkable and what their implications are for the evolution of the auditory system. Finally, we discuss the implications for amplitude coding in birds.

Comparison with other bird studies – how species-specific is NA?

There are several previous studies of NA physiology in different bird species (Hotta 1971; Sachs and Sinnott 1978; Sullivan 1985; Sullivan and Konishi 1984; Warchol and Dallos 1990). They differed substantially in their findings and our new data seem to add yet more variety and conflict with earlier reports on the same species, the barn owl. We argue, however, that the observed differences between species and studies may be to a large extent methodological in nature. Our study had two important advantages over previous work. First, increasingly standardized methods exist for analyzing and classifying unit responses in the mammalian cochlear nucleus. While we did not assume a priori that our data would fit existing mammalian categories, we made use of the quantitative methods of analysis (e.g. regularity analysis, Young et al. 1988) as a basis for classification. Second, a large database was available about the responses of auditory-nerve fibres in the same species (Köppl 1997a, b; Köppl and Yates 1999), which proved essential for recognizing primary-like NA units.

Sullivan (1985) found a large majority (92%) of chopper units in the barn owl NA, with the remaining 8% being onset units. We believe that having no auditory-nerve data available, Sullivan conservatively interpreted any primary-like units he encountered as auditory nerve. This is consistent with the data shown in Sullivan and Konishi (1984) for phase-locking of presumed auditory-nerve fibres and NA units. The vector strengths for their NA units are largely overlapping with those of our chop-T units and many of their auditory-nerve values fall within the range of our primary-like units. Furthermore, “large, easily-isolated spikes” were used as one criterion to distinguish NA units from auditory-nerve fibres (Sullivan and Konishi 1984). This may have biased the sample in a different way compared to our set of data. We often found it difficult to obtain good unit isolation in NA and had best success with relatively high-impedance tungsten electrodes. Unit isolation was especially critical in the case of typeIV units, where vigorous background discharges could sometimes be observed during inhibition of the isolated unit. Several distinct morphological cell types are known in the barn owl NA, with different soma sizes and varying extents of dendritic arbours (Soares and Carr 2001). Should there be a correlation between morphology and physiology, it is possible that the probability and quality of extracellular recordings differs for the various response types. TypeIV units may also be mistaken for non-auditory neurones if CF and threshold are determined audiovisually (Sullivan 1985), instead of more extensively probing for the response map. Their weak excitatory responses are difficult to discern above the relentless spontaneous discharge and inhibitory responses may be partly masked by background excitatory discharge. We thus suggest that a
combination of technical factors could explain the substantial differences between our data and previous work on the barn owl.

Another study on the redwing blackbird, a songbird, found a large majority (about 81%) of primary-like responses, about 10% type IV units and some onset (5%) and pauser (4%) units (Sachs and Sinnott 1978). Interestingly, no chopper-like responses at all were observed. We believe that technical factors may have precluded a distinction between what we call primary-like and chop-T neurones. Sachs and Sinnott’s study predated the introduction of regularity analysis as a useful criterion for this distinction and the bin width of the PSTHs shown would have been too crude to visually reveal the characteristic fast chopping pattern of chop-T responses (unfortunately, no bin width was specified, but it can be estimated at 5-10 ms from the figures). Indeed, if our primary-like and chop-T categories in the barn owl are combined, the distributions of the 4 remaining response types are similar to the redwing blackbird. A further intriguing point is the detailed similarity of type IV responses in the redwing blackbird and the owl. In both species, type IV neurones had high spontaneous rates, although most other NA neurones had lower spontaneous rates than auditory-nerve units. Their response maps showed interleaving excitatory and inhibitory areas, which differed between individual neurones. The rate-level functions at CF showed varying degrees of inhibition below spontaneous rate and the PSTH showed an onset-inhibitory response at levels with net inhibition. Finally, Sachs and Sinnott (1978) also provided evidence that the inhibitory component in type IV responses is dynamic, by eliminating it through systemic administration of barbiturate.

Warchol and Dallos (1990) recorded in the chicken NA and, based on rate-level functions at CF and visual inspection of the PSTH at a fixed level above CF-threshold, also defined several response types. The majority (40%) were primary-like, 28% were chopper responses and 8% onset responses. This corresponds almost exactly to what we found in the barn owl. However, the remaining 24% of the chicken neurones showed the unusual combination of a non-monotonic rate-level function, suggestive of type IV responses, and a chopper-like PSTH. Although chopper PSTH can be associated with type IV response maps in mammals (see e.g. review by Romand and Avan 1997), this was never seen in the red-wing blackbird and the barn owl. A confounding factor may have been the use of barbiturate for the induction of anaesthesia in the chicken. Barbiturate has been shown to eliminate the inhibitory response component of type IV neurones (Sachs and Sinnott 1978) and, depending on the duration of this effect, may have altered the responses recorded by Warchol and Dallos (1990). Further studies are needed to determine whether type IV neurones are truly present in the chicken.

In summary, although the results from studies of the avian NA in different species and across different labs appear to differ substantially, technical issues may be responsible for a large part of these differences. Taking those into account, the available data can be reconciled into a fairly consistent scheme across species. There are 4 to 5 different response types in NA, which may partly grade into each other. The most common one in all species is probably the primary-like type, followed by a chopper-like response type. Further, more detailed studies are needed to clarify whether the chop-T classification found appropriate for most chopper responses in the barn owl is typical for birds in general. Although Sullivan (1985) classified his chopper units as transient, this was not backed by regularity analysis. Chopper responses of the sustained type were only clearly documented in two rare cases in the owl and their unusually long latencies may indicate that these are not NA neurones, but possibly afferents from the superior olive, known to project to NA (e.g. Yang et al. 1999). Onset responses are also typically present, in low
proportions of less than 10%. TypeIV responses were unambiguously documented in the owl and the redwing blackbird, in similar proportions of 10-15%. They are thus likely to be a typical feature of the avian NA.

Comparison with the mammalian cochlear nucleus – how far does the analogy go?

The great similarity of the physiological responses in the cochlear nucleus between birds and mammals is our most interesting result. There is firm evidence now for primary-like, chopper-transient, onset and typeIV responses in the NA, as well as another primary-like population in the NM (Sachs and Sinnott, 1978; Sullivan and Konishi, 1984; Sullivan, 1985; Warchol and Dallos, 1990). As in mammals (Blackburn and Sachs, 1989; Joris et al., 1994), primary-like units tend to show inferior phase-locking compared to auditory-nerve fibres at high frequencies, however there is no firm evidence for their superior phase-locking at frequencies below 1 kHz (Köppl, 1997b; this study). In addition, NA primary-like units show lower, NM units higher average spontaneous rates than auditory-nerve fibres. The primary-like-with-notch response type is probably absent in birds. Chopper responses in birds appear to be mainly of the transient type, with only tentative evidence for a rare sustained type (this study). This is in contrast to mammals, where chop-S units are always common and often the predominant type of chopper (e.g. Rhode and Smith, 1986; Young et al., 1988; Blackburn and Sachs, 1989; Winter and Palmer, 1990). However, the distinction of mammalian chop-T and chop-S units by their degree of regularity is somewhat arbitrary and Rhode and Smith (1986) have argued that there is a continuum. It is thus possible that birds simply show less variation or a more skewed distribution within the same basic response type. Onset units are also definitely present in the NA, but they were fairly rare in all studies. In mammals, onset units are commonly subdivided into 3 variants, with onset-chopper being the most common (e.g. Rhode and Smith, 1986; Winter and Palmer, 1995). Some onset units in birds (e.g. Fig. 10; Sachs and Sinnott, 1978, Fig. 9; Sullivan, 1985, Fig. 11) are similar to the onset-I type of mammals and no evidence was found so far for onset-chopper responses. However, any definite classification must await a larger sample. Finally, typeIV responses in birds resemble those of mammals in great detail, even showing the same variation in the inhibitory component between units and its anaesthesia sensitivity (reviews in Rhode and Greenberg, 1992 and Young and Davis, 2002; Sachs and Sinnott, 1978; this study).

Considering the fundamental morphological differences between the cochlear nuclei of birds and mammals, the great similarity of response types was somewhat surprising. NA has no immediately obvious equivalent in mammals. It is a heterogeneous nucleus with no prominent subdivisions and with several morphological cell types distributed across the tonotopic gradient (Häusler et al. 1999; Soares and Carr 2001). This is in contrast to the complexity of the mammalian cochlear nucleus with many subdivisions and most cell types characteristically concentrated within those (e.g. review in Cant 1992). In addition, while some of the morphological cell types in the NA bear obvious similarities to types found in mammals, e.g. the radiate and planar multipolar neurones, the most common avian type, termed “stubby” (Soares and Carr 2001), has no equivalent in mammals.

This raises the very interesting question of how birds implemented those same physiological types. TypeIV units represent an especially intriguing case, since this response type, in mammals, is typically found in the DCN (recent review in Young and Davis 2002), a part of the cochlear
nucleus with cerebellum-like organisation that birds do not have. An elaborate circuit, including
direct projections from the auditory nerve and a multitude of both excitatory and inhibitory
connections from other cochlear-nucleus neurones (both DCN and VCN) shapes the type IV
response. It appears that the neuronal morphologies and circuits underlying these responses may
well be different, representing an intriguing case of independent evolution in birds and mammals,
similar to that pointed out for the avian visual wulst and mammalian visual cortex (Pettigrew and
Konishi 1976). Indeed, recent evidence from fossil studies on the middle ear (Clack 1997) and
comparative studies on the inner ear (Manley and Köppl 1998) suggests that the sensitive, high-
frequency hearing of airborne sound may be a relatively late development in vertebrate evolution
that happened independently in the major clades. Such dramatic changes in the auditory
periphery may have imposed powerful selective pressures on the central auditory system
(Wilczynski 1984), leading to the independent evolution of sophisticated auditory processing
capabilities. Consistent with this hypothesis, onset- and chopper-like responses were also found
in the dorsal medullary nucleus, the primary auditory brainstem nucleus of frogs (Hall and Feng,
1990), whose homology to the cochlear nucleus of other vertebrates is highly controversial (e.g.
review by McCormick, 1999). Future studies are needed that explore the links between cellular
morphology and physiology in the avian NA, as well as details of the neural circuitry and
pharmacology.

**Nucleus angularis – the starting point for intensity processing?**

A classic view of the avian cochlear nucleus is that of a clear dichotomy between its two main
subdivisions, with NM specializing in temporal coding and NA in sound level coding. This
interpretation is heavily based on the elegant experiments of Takahashi et al. (1984), which
showed that silencing NA eliminated the sound-level-based selectivity of midbrain “space-
specific” neurones involved in sound localization. Later, the data of Sullivan and Konishi (1984)
suggested a nearly homogenous neurone population in NA with large dynamic ranges, low
spontaneous and high saturation discharge rates, i.e. well suited for the coding of sound level at
frequencies near their CF. Our results, as well as studies in other avian species, however, have
established a much greater variety of responses and showed that the classic depiction of NA is
too simplistic. Instead, they suggest that NA, in contrast to NM, is the starting point of multiple,
distinct auditory processing streams. Since nothing is known about potentially different
projection targets of the different response types, we may draw inferences based on comparisons
with the mammalian literature.

Clearly, as earlier experiments (Takahashi et al., 1984) had shown, NA is involved in the
processing of sound level. However, sound level processing may involve more than coding for a
wide dynamic range via average discharge rate. Certainly, none of our response types appeared to
be specialized for large dynamic ranges or high driven rates. Although individual examples of
both were found, at the population level, the response types did not differ from each other or from
the auditory nerve in these respects. We suggest that instead, the major response types in NA may
all mediate different aspects of level coding. In addition to the rate information relayed by
primary-like units, the lower discharge variability of chopper-type responses may make these
units particularly suitable as inputs to interaural level comparison circuits, where accurate,
invariant information about the levels at both ears is important (Manley et al. 1988; Shofner and
Dye, 1989; Mogdans and Knudsen 1994). Finally, type IV units may encode spectral features, i.e. level information integrated over a wider band of frequencies (Yu and Young 2000).

In mammals, it is thought that type IV DCN neurones are involved in the detection of spectral notches, characteristic nulls in the spectrum that are caused by the acoustic filtering properties of the pinna and provide reliable cues to sound direction (May 2000; Young and Davis 2002). Could they also fulfil this role in birds? In the barn owl, localization cues in the form of monaural spectral notches do exist at high frequencies above 6 kHz (Keller et al. 1998). This is due to the specialized feather mask that effectively works like an immobile pinna. It has also been shown that owls are able to discriminate noisy signals with missing components relative to a learned reference, i.e. are principally able to recognize spectral-notch-like signals (Konishi and Kenuk 1975). However, there is little evidence that the animal uses spectral-notch cues in sound localization (Poganiatz and Wagner 2001). Moreover, we found type IV neurones in the owl’s NA with CFs down to frequencies below 1 kHz. At those frequencies, monaural spectral cues have not been shown for any avian species, but could theoretically be generated by a pressure difference receiver mechanism mediated by the interaural canal in many birds. Whether these cues could be of sufficient magnitude and could, e.g., take the form of sharp spectral notches, is still controversial (recent review in Klump 2000). In summary, there is little evidence to support the hypothesis that type IV neurones in birds are used in sound localization. They may instead serve the more general function of detecting sharp spectral features (notches or peaks) in communication or other environmental sounds (Young and Davis 2002).

In addition to sound level processing being more varied that previously appreciated, it may not be the only function of NA. The presence of onset units argues for an additional involvement in temporal processing. In mammals, onset responses are found in several cell types in the cochlear nucleus (reviews in Rhode and Greenberg 1992; Romand and Avan 1997) and some are thought to encode temporal features such as broadband transients (e.g. Oertel et al. 2000). NA onset neurons may serve a similar function. This does not question the prominent and well-established role of NM in temporal processing. However, it emphasizes that NM is a very specialized nucleus, focussed on one particular aspect of temporal coding (i.e. phase locking), while NA is more versatile and probably the source of multiple ascending auditory pathways.

ACKNOWLEDGEMENTS

We acknowledge Chris Malek for programming support, Dr. J. Pena for assistance with Matlab and Dr. D. Soares for help with data acquisition and histology. Knowles Inc. generously donated samples of their miniature microphones. Geoff Manley and several anonymous reviewers kindly commented on an earlier version of the manuscript. The work was supported by a Heisenberg Fellowship of the Deutsche Forschungsgemeinschaft to CK, by NIH DCD00436 to CEC and by the University of Maryland Center for Neuroscience.
REFERENCES


FIGURE LEGENDS

Fig. 1: Two examples of lesions that were placed to mark recording sites and were subsequently found in NA. A and C: Drawings of the outlines of the brainstem and several auditory nuclear areas in a transverse section. The areas of lesions are drawn stippled, dashed outlines indicate the areas shown at higher magnification in B and D. NA = Nucleus angularis, NM = N. magnocellularis, NL = N. laminaris. B and D: Photomicrographs of NA at higher magnification. Note that the lesion shown in B was 8 days old and displayed the accumulation of glial cells typical after that time. For the lesion shown in D, more current was used and survival time was only a few hours with little time for repair processes; thus the lesion has a more vacuolated appearance. Scale bar applies to B and D.

Fig. 2: Decision tree developed for classifying the recorded units.

Fig. 3: Example for a unit with primary-like response. A: Tuning curve. B: Rate-level functions for stimulation at CF = 10 kHz and with noise. C: Peri-stimulus time histograms (PSTH) at CF, at 80 dB SPL = 38 dB above threshold, and (inset) 60 dB SPL = 18 dB above threshold. The stimulus was presented between 5 and 55 ms. D and E: regularity analysis for the PSTH at 80 dB SPL. The interspike interval (solid line) and its standard deviation (dotted line) are plotted as a function of time (in 0.1 ms bins) in D, the coefficient of variation (CV) in E. All lines in D and E represent 3-point weighted running averages. Mean CV was calculated for a time window of 12-20 ms after neural latency. F: Interspike interval distribution during stimulation at CF, at 80 dB SPL. G: PSTH for stimulation with noise at 70 dB SPL spectrum level.

Fig. 4: Vector strength for stimulation at CF, at least 20 dB above threshold, as a function of CF. Data for the different response types are drawn with different symbols as indicated; the dot data points and dashed line serve as a reference, displaying published data and median values for the auditory nerve (Köppl 1997b).

Fig. 5A: Spontaneous discharge rate as a function of CF. Data for the different response types are drawn with different symbols as indicated; the dot data points and dashed line serve as a reference, displaying published data and an exponential fit for auditory-nerve data (Köppl 1997a). B: Distribution of spontaneous rates, separately for units classified as NA and auditory nerve, respectively. Note that the population of NA units was skewed towards low rates.

Fig. 6: Example of a unit with a chop-T response, showing a high saturation discharge rate. The layout is the same as in Fig. 3. A: Tuning curve. B: Rate-level functions for stimulation at CF = 6 kHz and with noise. C: PSTH at CF, at 70 dB SPL = 34 dB above threshold and (inset) 50 dB SPL = 14 dB above threshold. D and E: regularity analysis for the PSTH at 70 dB SPL. F: Interspike interval distribution during stimulation at CF, at 70 dB SPL. G: PSTH for stimulation with noise at 60 dB SPL spectrum level.

Fig. 7: Example of a unit with a chop-T response, showing an average saturation discharge rate. The layout is similar to Fig. 3. A: Tuning curve. B: Rate-level functions for stimulation at CF = 7.4 kHz and with noise. C: PSTH at CF, at 40 dB SPL = 34 dB above threshold; the inset shows the PSTH to the same stimulus, but with 5 ms rise/fall times. D and E: regularity analysis for the PSTH at 40 dB SPL. F: Interspike interval distribution during stimulation at CF, at 40 dB SPL.

Fig. 8: Example of one of the rare chop-S units. The layout is similar to Fig. 3. A: Tuning curve. B: Rate-level functions for stimulation at CF = 6.3 kHz and with noise. C: PSTH at CF, at 50 dB SPL = 26 dB above threshold and (inset) 40 dB SPL = 16 dB above threshold. D and E: regularity analysis for the PSTH at 50 dB SPL. F: Interspike interval distribution during stimulation at CF, at 50 dB SPL. G: PSTH for stimulation with noise at 60 dB SPL spectrum level. H and I: Analog recordings of the response waveform for one particular presentation of the stimulus at 39 and 60 dB SPL, respectively. Both graphs are scaled identically. Note the decreasing amplitude of the spikes during the response to the higher stimulus level.

Fig. 9: Minimal response latency to stimuli at CF, at 20 to 35 dB above threshold, as a function of CF. Data for the different response types are drawn with different symbols as indicated. Each unit is only represented once. Note the extensive overlap between all response types, except for chop-S.
Fig. 10: Example of a unit with an onset response, showing little sustained discharge. **A**: Tuning curve. **B**: Rate-level functions for stimulation at CF = 4.7 kHz and with noise. **C**: PSTH at CF, at 75 dB SPL = 42 dB above threshold and (inset) 55 dB SPL = 22 dB above threshold. **D**: PSTH for stimulation with noise at 50 dB SPL spectrum level.

Fig. 11: Example of a unit with an onset response, showing a robust sustained discharge as well. **A**: Tuning curve. **B**: Rate-level functions for stimulation at CF = 4.0 kHz and with noise. **C**: PSTH at CF, at 40 dB SPL = 25 dB above threshold and (inset) 20 dB SPL = 5 dB above threshold. **D and E**: Regularity analysis for the PSTH at 40 dB SPL. **F**: Interspike interval distribution during stimulation at CF, at 40 dB SPL. **G**: PSTH for stimulation with noise at 50 dB SPL spectrum level.

Fig. 12: Example of a unit with a type IV response. **A**: Response map. The areas enclosed by solid black lines delineate excitatory stimuli, the gray line indicates the lower boundary of the inhibitory area. The 3 dots indicate the frequency and levels of the stimuli used for the PSTHs in C-E. **B**: Rate-level functions for stimulation at CF = 7.4 kHz and with noise. The dashed line indicates the spontaneous discharge rate. Both functions were clearly non-monotonic. **C-E**: PSTH at CF, at 10, 30 and 50 dB SPL = 5, 25 and 45 dB above excitatory threshold. Note the change from a sustained weak excitatory response to a pronounced onset response followed by inhibition. **F**: PSTH for stimulation with noise at 50 dB SPL spectrum level.

Fig. 13: Example of a unit with an extreme type IV response, showing no inhibition below spontaneous rate. **A**: Response map. There was a V-shaped band of excitation (shown by the solid black lines). The 3 dots indicate the frequency and levels of the stimuli used for the PSTHs in C-E. **B**: Rate-level functions for stimulation at CF = 7.2 kHz and with noise. The dashed line indicates the spontaneous discharge rate. Note that the CF-function was clearly non-monotonic, while the response to noise was monotonic. **C-E**: PSTH at CF, at 12, 30 and 50 dB SPL = 9, 27 and 47 dB above excitatory threshold. Note the change from a sustained weak excitatory response to a pronounced onset response followed by a brief, weak inhibition. **F**: PSTH for stimulation with noise at 35 dB SPL spectrum level.

Fig. 14: Example of a unit with an extreme type IV response, showing no excitation above spontaneous rate. **A**: Inhibitory tuning curve, drawn in gray. The 3 dots indicate the frequency and levels of the stimuli used for the PSTHs in C-E. **B**: Rate-level function for stimulation at CF = 4 kHz. The dashed line indicates the spontaneous discharge rate. **C-E**: PSTH at CF, at 20, 30 and 50 dB SPL = -1, 9 and 29 dB relative to the inhibitory threshold. Note the change from a sustained weak inhibitory response to an excitatory onset response followed by pronounced inhibition. **F**: PSTH for stimulation with noise at 50 dB SPL spectrum level.

Fig. 15: Example of a change in the rate-level function of a type IV unit. Two functions are shown that were recorded at CF (700 Hz) under identical conditions, 35 minutes apart. Note that the spontaneous rate (indicated by the dashed line) did not change. However, threshold shifted slightly and the relative amounts of excitation and inhibition changed.

Fig. 16A: Threshold at CF, as a function of CF. Data for the different response types are drawn with different symbols as indicated in B. The dashed line serves as a reference displaying a 2nd order polynomial fit to the values of 335 auditory-nerve fibres (own partly unpublished data). **B**: Q10dB of the tuning curve, as a function of CF. Note that type IV units have been omitted from this graph, since a conventional definition of a tuning curve is not possible for these units. The dashed line serves as a reference displaying a published power fit for the auditory nerve (Köppl 1997a).
TABLES

Table 1: Summary of salient characteristics of the different response types. In the first column, the types and the numbers found of each are listed. Columns 2-7 list characteristic frequency (CF), threshold at CF, the spontaneous discharge rate, the saturation discharge rate, the dynamic range and the driven discharge range (saturation rate – spontaneous rate). For each parameter, the median, minimum and maximum values and the number of units are given.
# Table 1: Summary of salient characteristics of the different response types

<table>
<thead>
<tr>
<th>Type</th>
<th>CF Threshold at CF</th>
<th>Spont. rate (spikes/s)</th>
<th>Saturation rate (spikes/s)</th>
<th>Dyn. range (dB)</th>
<th>Driven rate (spikes/s)</th>
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<td>22.5 (dB SPL)</td>
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<td></td>
<td>n</td>
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<tr>
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<td></td>
<td>n</td>
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FIG. 1
Rate-level function non-monotonic
PSTH onset-inhibitory

YES

"Type IV"

NO

VS within auditory-nerve range

YES

All other information consistent with auditory nerve (spont. rate required)

YES

"Auditory nerve"

NO

Spont. rate within auditory-nerve range

NO

"NA unclassified"

YES

Primary-like

"Uncertain"

NO

"Primary-like" (only low CF)

PSTH available

YES

Primary-like

NO

Onset

PSTH primary-like
Mean CV > 0.7

YES

"Chopper-transient"

NO

"Chopper-sustained"

NO

PSTH with one sharp onset peak

NO

PSTH chopper-like
(minimum two peaks)
Mean CV > 0.4

YES

Latency above range of all other types
Mean CV < 0.4

NO

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FIG. 2
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FIG. 3
FIG. 5

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FIG. 6
FIG. 7
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FIG. 8
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FIG. 9
Köppl and Carr: Computational diversity in nucleus angularis

FIG. 10
Köppl and Carr: Computational diversity in nucleus angularis

FIG. 11
Köppl and Carr: Computational diversity in nucleus angularis

FIG. 12
Köppl and Carr: Computational diversity in nucleus angularis

**FIG. 13**
FIG. 14

Köppl and Carr: Computational diversity in nucleus angularis
Köppl and Carr: Computational diversity in nucleus angularis

FIG. 15
Köppl and Carr: Computational diversity in nucleus angularis

FIG. 16