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Title: The roles of monkey premotor neuron classes in movement preparation and execution

Abbreviated title: Characterizing interneurons in monkey PMd

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Abstract

Dorsal premotor cortex (PMd) is known to be involved in the planning and execution of reaching movements. However, it is not understood how PMd plan activity – often present in the very same neurons that respond during movement – is prevented from itself producing movement. We investigated whether inhibitory interneurons might ‘gate’ output from PMd, by maintaining high levels of inhibition during planning and reducing inhibition during execution. Recently-developed methods permit distinguishing interneurons from pyramidal neurons using extracellular recordings. We extend these methods here for use with chronically-implanted multi-electrode arrays. We then applied these methods to single- and multi-electrode recordings in PMd of two monkeys performing delayed-reach tasks. Responses of putative interneurons were not generally in agreement with the hypothesis that they act to gate output from the area: in particular it was not the case that interneurons tended to reduce their firing rates around the time of movement. In fact, interneurons increased their rates more than putative pyramidal neurons during both the planning and movement epochs. The two classes of neurons also differed in a number of other ways, including greater modulation across conditions for interneurons, and interneurons more frequently exhibiting increases in firing rate during movement planning and execution. These findings provide novel information about the greater responsiveness of putative PMd interneurons in motor planning and execution, and suggest that we may need to consider new possibilities for how planning activity is structured such that it does not itself produce movement.
**Introduction**

Dorsal premotor cortex (PMd) is known to be involved in both the planning and execution of reaching movements. Frequently, both planning and movement-related activity are exhibited in the very same neurons (Tanji and Evarts 1976; Weinrich and Wise 1982). We therefore ask a basic question: why does PMd plan activity not drive movement?

When a monkey is cued about the path of an upcoming reach but required to withhold it until a go cue, PMd activity exhibits tuning for parameters of the reach during the plan period (Churchland et al. 2006b; Godschalk et al. 1985; Hocherman and Wise 1991; Messier and Kalaska 2000; Riehle and Requin 1989). PMd activity also predicts reaction time (RT; Churchland et al. 2006c; Riehle and Requin 1993) and variability in the upcoming movement (Churchland et al. 2006a). Further arguing that PMd is specifically involved in movement planning, disruption of PMd activity near the time of the go cue delays movement onset (Churchland and Shenoy 2007). Additionally, it is known that PMd sends projections down the spinal cord to motor interneurons (Dum and Strick 1991), and microstimulation in PMd causes arm movements (e.g., Weinrich and Wise 1982). Hence, movement-epoch PMd activity presumably contributes to actually driving the movement. Theoretical treatments thus often assume a ‘gate’ between plan and movement-related neurons (e.g., Bullock and Grossberg 1988; Cisek 2006a), and pharmacological experiments have suggested that reduction of inhibition might impair the ability to withhold premature movements (Sawaguchi et al. 1996).
In the oculomotor system, this gating process is fairly well understood. During saccade preparation, cortical areas such as the frontal eye fields (FEF) generate a representation of the upcoming saccade. A downstream subcortical area (nucleus raphe interpositus) contains ‘omnipause neurons’ (Cohen and Henn 1972), which tonically inhibit saccade-generating ‘burst neurons’ in the brainstem (paramedian pontine reticular formation and rostral interstitial nucleus of the median longitudinal fasciculus; for review, Fuchs et al. 1985). Omnipause neurons stop firing during the saccade, releasing their inhibitory gate of burst neuron output (Evinger et al. 1982). A hypothetical mapping of this mechanism to PMd is shown schematically in Fig. 1. In this mapping, corticospinal neurons might correspond to burst neurons, local interneurons to omnipause neurons, and local pyramidal neurons to upstream areas such as FEF.

While PMd sends axons down the spinal cord, analogously to burst neurons, it is a cortical area, like FEF. There is also a small amount of preparatory activity even in the spinal cord (Prut and Fetz 1999). Thus, it is not clear whether PMd should be more closely analogous to FEF or brainstem saccade areas, or use some other mechanism entirely to prevent premature movements. It is known that PMd can exert inhibitory as well as excitatory effects on primary motor cortex (M1; Ghosh and Porter 1988; Keller and Asanuma 1993; Tokuno and Nambu 2000), which might point towards a feedforward-inhibition gating mechanism.

A recent study examined whether an oculomotor-like output-gating mechanism might be at play in forelimb movements in rats (Isomura et al. 2009). They found evidence against such a mechanism, but they also note that rats do not have a clear PMd-M1 separation and found that interneurons were only weakly tuned, in contrast to known
interneuron tuning in monkey M1 (Merchant et al. 2008). Thus, it remains unclear what mechanism is at work in preventing PMd plan activity from driving movement in the monkey.

Different hypotheses about movement gating make different predictions regarding the relative activity patterns of interneurons and pyramidal neurons. Examining differences in their patterns of activity could thus be informative regarding the gating mechanism. We therefore wished to identify interneurons and pyramidal neurons in our recordings. Since interneurons have briefer action potentials than pyramidal neurons (Connors and Gutnick 1990; McCormick et al. 1985), and the extracellular waveform reflects the intracellular waveform (Henze et al. 2000), the extracellularly-recorded waveform duration can be used to distinguish interneurons from pyramidal neurons with substantial reliability (Bartho et al. 2004). This technique has previously been used in primary somatosensory cortex (Simons 1978; Swadlow 2003), prefrontal cortex (Diester and Nieder 2008; Johnston et al. 2009; Rao et al. 1999; Wilson et al. 1994), V4 (Mitchell et al. 2007), FEF (Cohen et al. 2009), and M1 (Merchant et al. 2008).

We first replicate prior findings of a bimodal distribution of spike-waveform durations for single-electrode recordings. We then extend this method for use with chronically-implanted electrode arrays. We observed consistent differences in the activity patterns of putative interneurons versus putative pyramidal neurons. Those differences did not follow the predictions of the output-gating ‘omnipause’ hypothesis; interneuron activity was highest, not lowest, around movement onset. Nor were our results consistent with other, more refined variants of the output-gating hypothesis. These results imply that either gating is not accomplished through inhibition, or that the gate is located
116 downstream of PMd.

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Materials and Methods

Subjects. Animal protocols were approved by the Stanford University Institutional Animal Care and Use Committee. Subjects were two adult male macaque monkeys (Macaca mulatta) trained to perform variants of the delayed reach task for juice reward. After initial training, we performed a sterile surgery during which the monkeys were implanted with a head restraint and either a 96 electrode silicon array (monkey H) or a standard recording cylinder (monkey J). The electrode array (Blackrock Microsystems, Salt Lake City, UT) was implanted in caudal PMd (adjacent to primary motor cortex), as estimated visually from local anatomical landmarks (Fig. 2A). Array recordings yielded strong, well-tuned plan-period activity for arm movements.

The cylinder (Crist Instruments, Hagerstown, MD) was centered over caudal PMd, initially estimated using stereotaxic coordinates (13-17 mm anterior to stereotaxic zero, the intermeatal “ear bar” line) and from previous surgeries and MRIs in other monkeys. The cylinder was placed surface normal to the skull, which was left intact and covered with a thin layer of dental acrylic. To accommodate recording, 3 mm holes were drilled later under ketamine/xylazine anesthesia.

We confirmed the location of our cylinder with a subsequent craniotomy (Fig. 2B) as part of a later array implantation surgery. The single-electrode recordings reported here appear to be in PMd proper just lateral of the precentral dimple, though potentially near the PMd/M1 ‘transition zone’ (Keller 1993; Weinrich and Wise 1982; Wise et al. 1986). While we cannot definitively localize PMd without histology, consistent with the PMd classification plan-period activity was common and robust for PMd recordings, but
not for surface or sulcal M1 sites (recorded separately, not otherwise reported here). Also, microstimulation thresholds for our PMd recordings were higher (median 50 μA) than in M1 (median 25 μA). Microstimulation evoked movements of the shoulder and upper arm, or (much less often) of the wrist. A number of our recordings were made deeper (> 4 mm) than the typical depth of cortex, consistent with the thickening of cortex near the precentral dimple.

Task apparatus. We used the same task apparatus as described previously (Churchland et al. 2006c). Briefly, during experiments monkeys sat in a customized chair (Crist Instruments) with the head restrained. The left arm of monkey J was restrained loosely using a tube and a cloth sling; monkey H did not have an arm restraint, but seldom moved the non-reaching arm from his side. Stimuli were back projected onto a frontoparallel screen ~27 cm from the eyes (the exact distance depended on the size of the monkey). A photodiode was used to record the timing of video frames with 1 ms resolution. The position of a reflector taped to the fingers was tracked optically in the infrared (Polaris system; Northern Digital, Waterloo, Ontario, Canada). The eyes were also tracked in the infrared (Iscan, Burlington, MA). A clear acrylic shield prevented the monkey from touching the dichroic eye-tracking mirror or from bringing the reflector to his mouth. A tube fixed to this shield dispensed juice rewards.

Task design. The tasks for both monkeys were variants of the center-out delayed reach task (Fig. 3), described previously (Churchland et al. 2006c). Experiments consisted of trials, each a few seconds long, which ended in a juice reward if successful. The animal began a trial by fixating and touching (for at least 400 ms) a fixation spot, after which a target appeared. After a plan period, a go cue was given, and reaches were
rewarded if they were brisk and accurate. Reward was delivered after the target was held
for 300 ms (monkey H) or 450 ms (monkey J), with the next trial beginning a few
hundred milliseconds later.

For monkey H, the fixation spot was central and the plan period was 200-1000
ms. Only trials with delay periods >400 ms were analyzed. The go cue was a slight
enlargement of the target and the disappearance of the fixation spot. RTs were required to
fall between 150 and 500 ms. Two rings of 8 targets were used, for a total of 16
conditions. For this monkey, each target location is hereafter referred to as a ‘condition.’

Monkey J performed a novel variant of the delayed-reach paradigm, called the
maze task. This task also required delayed reaches (0-900 ms plan period) to targets.
Again, only trials with delay periods >400 ms were analyzed. The maze task includes a
number of additional complexities (explained below), but here we use it simply as a 24-
condition delayed reach task. In the maze task the monkey touched the screen, but
contacted the targets with a virtual cursor floating 2.5 cm above his hand. Additionally,
we required that the cursor path not pass through a set of virtual barriers which varied
trial to trial. Thus, the monkey was required to make curved reaches on most trials.
Cursor contact with a barrier resulted in an aborted (un-rewarded) trial. Barriers appeared
at the same time as the target, and trials with barriers were interleaved with no-barrier
trials. Mazes varied in start point, end point, and barrier positions. Different mazes were
used for different neurons, with 24 stimuli (including those with and without barriers) for
each neuron. Effort was made to span the major reach directions and curve shapes for
each neuron recorded. The target jittered slightly (2 mm) when it appeared, and the
cessation of this jitter and disappearance of the central fixation spot comprised the go
cue. RTs were required to fall between 150 and 600 ms. For this monkey, each unique
maze (i.e., start point, end point, and reach shape) is hereafter referred to as a ‘condition.’

Neural recordings, classification of neuron types, and EMG. Neural and EMG
recordings were made using previously described techniques (Churchland et al. 2006c).
Neurons were classified as narrow- or broad-spiking using waveform analysis techniques
similar to those in Mitchell et al. (2007). Full details are given in the Appendix.

Neural analyses. We operationally define each neuron’s ‘cross-condition
modulation pattern’ as simply its mean response (in some time window) across
conditions (16 or 24 conditions, depending on the monkey). This is effectively a tuning
pattern, but no parametric tuning model (e.g., for direction, endpoint, etc.) is assumed. To
obtain the cross-condition modulation pattern, we took the mean firing rate over the
relevant epoch for each condition, resulting in a 16- or 24-element vector. We define the
‘cross-condition modulation’ as the maximum element of the vector minus the minimum
element. For analyses of tuning consistency, we produce one such vector for the baseline
period, one for the plan period, and another for the peri-movement period. We subtract
the baseline vector from each the plan and peri-movement vectors, and correlate the two
resulting vectors.
Results

Separation of narrow- and broad-spiking neurons

Previous work has established the trough-to-peak duration (TTP) of the spike waveform as among the more reliable methods for distinguishing putative interneurons from pyramidal neurons (Bartho et al. 2004; Mitchell et al. 2007). Specifically, the distribution of TTPs is often bimodal, with the briefer mode thought to correspond with inhibitory interneurons and the second mode corresponding predominately with pyramidal neurons (Krimer et al. 2005). The only known mammalian narrow-spiking neurons that are excitatory, spiny stellate cells, are not thought to be common outside of Layer IV of primary sensory cortex (Okhotin 2006). Additionally, previous studies in M1 and elsewhere have used spike-triggered averages to demonstrate that neurons with narrow spike waveforms are inhibitory with substantial reliability (Bartho et al. 2004; Merchant et al. 2008). Nonetheless, since we cannot know for certain that every neuron with a narrow waveform is an interneuron, or that every neuron with a broad waveform is not, we refer to them as narrow-spiking neurons and broad-spiking neurons to be explicit that we are not directly measuring whether each neuron is inhibitory or excitatory.

In monkey J, we recorded 163 neurons using single electrodes. We replicated the bimodal distribution of TTPs (Fig. 4A) found by Mitchell et al. (2007). The bimodality of our TTP distribution was significant with $p = 0.011$ using Hartigan’s dip test with a bootstrap (Hartigan and Hartigan 1985; Mechler and Ringach 2002).

For monkey J, 34 neurons were identified as narrow-spiking (putative interneurons) and 79 were identified as broad-spiking (putative pyramidal neurons).
excluded from analysis 14 neurons with flat-topped after-potentials whose peak time could not be reliably measured. An additional 36 had non-canonical waveforms (e.g., missing a post-trough peak); these neurons are analyzed separately in a subsequent section. At most of the recorded depths, a consistent proportion of narrow-spiking neurons was found, though deeper recordings yielded mostly non-canonical waveforms (Fig. 2D). The fraction of narrow-spiking neurons found, 30% of identified neurons (21% of all recorded neurons), is consistent with the proportion found by others (26%: Merchant et al. 2008; 27%: Mitchell et al. 2007; 27%: Rao et al. 1999) and with the estimated fraction of interneurons in cortex, 20-30% (Connors and Gutnick 1990).

In monkey H, we recorded 71 single units using a chronically-implanted multi-electrode array. 20 neurons were identified as narrow-spiking and 31 as broad-spiking (Fig. 4B). This corresponds to 39% of identified neurons being narrow-spiking. This modest over-representation of narrow-spiking neurons may occur because multiple broad-spiking neurons were commonly present on a single channel of the array recordings. High-quality spike sorts were thus achievable for a greater fraction of the (relatively isolated) narrow-spiking neurons, and they were therefore included at a higher rate.

The precise depths of the implanted array tips are not known, but the electrodes are 1 mm long and were fully inserted immediately after surgical implantation. The electrode tips were therefore likely shallower on average than our single-electrode recordings. Thus, the array recordings were presumably primarily from the shallower cortical layers, while the single-electrode recordings sampled all layers. To our knowledge, narrow- and broad-spiking neurons have not previously been identified using
this recording technology. Although special steps were required to address the lack of two clear peaks in the TTP distribution (see Appendix), subsequent analyses yield very similar results for both our single-electrode data and our electrode array data.

Comparison of the responsiveness of the two neuron classes

Fig. 5 shows example PSTHs for one broad-spiking neuron, one narrow-spiking neuron, and one deep non-canonical neuron (to be discussed later). Though there was wide variation in the response patterns of different neurons within each class, these examples exhibit the most common patterns recorded. During the plan period, the broad-spiking neuron (blue) shows increases in firing rate for some conditions and decreases for others. The narrow-spiking neuron (red) is also strongly tuned during the plan period, but most of its firing rate changes are positive. For this neuron, firing rates rise even further shortly before movement onset, and drop back to baseline several hundred milliseconds after movement onset.

Fig. 6 (top row) plots population PSTHs for narrow-spiking neurons and for broad-spiking neurons, collapsing over all conditions. There are substantial average differences between narrow- and broad-spiking neurons. Narrow-spiking neurons tend to increase their firing rates more strongly during the plan period, and to show a much greater rise in firing rate before movement onset (as in the examples). Firing rates do not return to baseline until after movement onset. Surprisingly, the mean rate for broad-spiking neurons is almost unchanged between the baseline period and the plan period. That is, on average pyramidal neurons do not fire at higher rates during motor planning than at baseline. The flatness of the aggregate PSTH for broad-spiking neurons does not imply that they have weak cross-condition modulation, however. Instead, it indicates that
on average their firing rates decrease as often as increase, as in the example neuron (Fig. 5, left).

The bottom row of Fig. 6 plots mean cross-condition modulation, defined as the range of firing rates over all conditions at each time point independently. For both narrow- and broad-spiking neurons, cross-condition modulation rises substantially at target onset and stays high through movement onset. However, cross-condition modulation is greater for narrow-spiking neurons both during the plan period and around movement onset. Together with the mean PSTHs, this means that both classes of neurons show substantial cross-condition modulation, but cross-condition modulation was stronger for narrow-spiking neurons and their firing rates were more likely to increase than decrease. This contrasts sharply with results from rat forelimb motor cortex, in which interneurons show cross-condition modulation almost exclusively during the movement epoch, and show little tuning for movement direction (Isomura et al. 2009). Merchant et al. (2008), however, found a generally similar pattern of firing rate changes in monkey primary motor cortex.

To see whether the structure of the population PSTHs was representative, we performed a cell-by-cell analysis. For each neuron, we took the mean plan-period firing rate (50 to 400 ms after target onset) minus the mean fixation-period baseline firing rate. The histogram over neurons is plotted in the top row of Fig. 7. Consistent with the population PSTHs, the distribution for narrow-spiking neurons is shifted slightly to the right of that for broad-spiking neurons in both monkeys (monkey J, narrow vs. broad medians: 4.9 vs. 0.8 spikes/s, p < 0.02; monkey H: 0.5 vs. 0 spikes/s, p < 0.001, Mann-Whitney U tests). The second row plots movement-period activity (-100 to +200 ms from
movement onset) minus the baseline, and here the rightward shift for narrow-spiking neurons is more pronounced (J: 12.6 vs. 0.5 spikes/s, p < 0.002; H: 5.8 vs. 1.3 spikes/s, p < 0.03), again consistent with the population PSTHs. Results were similar when the analysis was performed over neuron-conditions instead of neurons.

We also tested whether neurons exhibited a ‘pause-like’ pattern of activity, as was hypothesized for interneurons in the output-gating model (see Fig. 1B). We took the mean plan-period activity minus the mean movement-period activity for each neuron, and plot the distributions in the bottom row of Fig. 7. A neuron with a tendency to be very active during plan and only weakly active during the movement has a large value for this measure. No significant difference was observed between cells classes in either monkey for this metric, and the trends present were not in the hypothesized direction (J: -3.6 vs. 0.8 spikes/s, p > 0.14; H: -5.3 vs. -0.8 spikes/s, p > 0.07). Thus, it was not the case that one class of neuron tended to have a high firing rate during the plan period and pause during the peri-movement period. Additionally, note that these distributions appear to be unimodal; there does not appear to be a subset of ‘pause-like’ neurons. Results were again similar when the analysis was performed over neuron-conditions instead of neurons.

From the output-gating hypothesis illustrated in Fig. 1, one might have expected a particular pattern of activity from interneurons: high and perhaps un-tuned firing rates during the plan period, and a pause in firing around movement onset. This is not what was observed for the narrow-spiking neurons (putative interneurons). Instead, such neurons generally showed strong cross-condition modulation during the plan period, followed by a rise in activity around movement onset. Nor was there a clear subset of
narrow-spiking cells with a pause-like response. These patterns do not appear consistent
with a straightforward formulation of the output-gating hypothesis, for which inhibition
should decline around movement onset.

Additional controls for neuron classification

We considered whether the observed effects could be a result of neuron-selection
bias. For the single electrode recordings, we attempted to isolate every neuron
encountered. Recordings were abandoned only if the isolation was lost, the neuron died,
or the neuron was completely unresponsive during the plan period. Few neurons were
abandoned due to a lack of responsiveness, perhaps because the novel task used with
monkey J evoked particularly strong responses. While our selection criteria could lead to
a general bias towards responsiveness, they should not lead to a systematic difference
between the classes. Further, since we recorded as many narrow-spiking neurons as
expected from the percentage of interneurons in cortex, our criteria probably did not
strongly bias us towards or away from recording interneurons. For the array recordings,
neuron selection was made purely on quality of isolation. Again, this should not produce
response differences between neuron classes, and similar results were found with the
array recordings as with single electrodes.

We also tested whether our effects were sensitive to the precise threshold for
separating narrow- and broad-spiking neurons. We re-divided the population using a
range of thresholds from 170-300 μs and calculated the mean difference between the
population PSTHs for the two classes. The differences varied smoothly with the threshold
value in both monkeys. Thus, the observed difference in firing rate changes is largely
insensitive to the precise threshold value.
We also considered that inhibitory interneurons might change their preferred condition between the plan period and the movement period (as observed by Rao et al. (1999) in prefrontal cortex). During planning of a movement, one set of interneurons could prevent premature execution while pyramidal neurons achieved the needed pattern of rates. During execution of the movement, these interneurons might decrease their inhibition, while another set of interneurons inhibited competing movements. Instead of an overall pause in firing for interneurons, this version of the output-gating hypothesis predicts that interneurons would invert their preferred conditions between the plan and movement epochs. In contrast, pyramidal neurons would be expected to maintain consistent preferred conditions. Thus, what was inhibited during plan would be released during movement.

We did not find this pattern in our data. We frequently observed changes in the preferred condition between the plan and movement periods. However, narrow-spiking neurons did not tend to fully invert their preferred conditions, and such preference changes were just as prevalent for broad-spiking neurons as for narrow-spiking neurons. PSTHs for three example neurons, all broad-spiking, are shown in Fig. 8 (top). These neurons are all well-modulated across conditions at nearly every time point. One of these neurons had consistent preferred conditions between the plan and movement periods (Fig. 8, top right). The other two examples’ patterns of cross-condition modulation change dramatically between the plan period and the peri-movement period: some most-preferred conditions during planning became least-preferred during movement, and vice versa. Such shifts are not visible in the population plots (Fig. 6), where cross-condition
modulation was computed as the most-preferred minus least-preferred condition at each
time point independently.

To quantify the extent of such changes in cross-condition modulation, and to ask
whether changes were more common for narrow-spiking neurons, we determined the
consistency between plan modulation and movement modulation for each cell. For each
well-modulated neuron ( > 5 spikes/s cross-condition modulation for plan and move), we
computed its “cross-condition modulation pattern”: a vector containing the mean rate for
each condition over an epoch, minus the baseline firing rate for that condition. We then
correlated the cross-condition modulation patterns for the plan period (50 to 400 ms after
target onset) with those from the peri-movement period (-100 to +200 ms from
movement onset). These correlations spanned a broad range (Fig. 8, bottom): being
sometimes near one (very similar modulation patterns during planning and movement)
and sometimes near negative one (pattern of preferences inverted). On average,
correlations were only slightly above zero (little relationship between the two patterns,
either consistent nor inverting). This was true for both monkeys for both narrow- and
broad-spiking neurons. These weak correlations using cross-condition modulation
patterns is consistent with previous findings using a cosine-tuning model (Crammond and
Kalaska 2000), with population statistics (Johnson et al. 1999), and with oscillatory
premotor neurons (Lebedev and Wise 2000). Results when fitting our data with cosine
tuning curves yielded similar patterns between narrow- and broad-spiking neurons as
well (Supplemental Fig. 1).

As a control, we considered whether this overall low correlation between plan-
and movement-period modulation patterns could be due to noise: if we poorly estimated
the mean firing rate due to low trial counts, this could reduce the correlation between patterns that were actually similar. To estimate the expected departure from unity correlation due to sampling noise, we re-sampled trials and calculated the re-sampled plan (move) modulation pattern against the original plan (move) modulation pattern; these re-samplings yielded values rather close to 1 (Fig. 8 gray arrows), implying that the wide range of correlations between the plan and move periods are not due to sampling noise. Finally, we considered whether having subtracted baseline activity to compensate for posture tuning might have reduced the correlations. However, when baseline activity was not subtracted, correlations were even lower.

**Non-canonical waveforms**

In classifying neurons into narrow- and broad-spiking, we excluded waveforms that did not conform to the classical pattern of a trough followed by a peak. In particular, this included many neurons that had only a single positive peak, or a positive peak followed by a small trough and with little post-trough peak (Fig. 9A). Computer simulations, in combination with systematically positioned extracellular recordings, suggest that such waveforms may be obtained from the distal portion of pyramidal neurons’ dendrites because of capacitative currents (Gold et al. 2006). In order to isolate a distal dendritic recording with other neurons nearby, presumably the neuron must be large. Since non-canonical waveforms were found almost exclusively deep in cortex (Fig. 2D), this may indicate that they belong primarily to large pyramidal neurons in layers V and VI, which are apt to project to other brain areas or down the spinal cord. Merchant et al. (2008) have recently argued for an identifiable subgroup of putative pyramidal neurons in primary motor cortex, characterized by having long duration waveforms,
having seemingly different connectivity, and being located primarily in Layer V. We suspect that the neurons we identify as having non-canonical waveforms may be a similar subgroup as that identified by Merchant et al. (2008), based on their depth. We found non-canonical waveform neurons almost exclusively in recordings from monkey J; the lack of non-canonical waveforms from the multi-electrode arrays is likely due at least in part to their having electrodes 1 mm long, and thus probably not recording from the deeper layers.

We analyzed these neurons separately. The example neuron shown in Fig. 5 (right) is representative of this neuron class. Such neurons typically had modest changes in firing rate during the plan period, with much greater (and mostly positive) changes in firing rate during the movement. This movement-period activity seems to be maintained relatively later into the movement than for the other two neuron classes. The pattern illustrated in the example neuron can be seen in the population PSTH and cross-condition modulation plots for non-canonical waveform neurons (Fig. 9B, C). This might indicate the presence of a mechanism for limiting the activity of output neurons within PMd.
These results show that one can distinguish, based on extracellularly-recorded waveforms, two classes of neurons in PMd: narrow-spiking (putative interneurons) and broad-spiking (putative pyramidal neurons). These classes differ, as populations, in their patterns of activity during both planning and execution of movements. Compared with putative pyramidal neurons, putative interneurons have stronger cross-condition modulation, are more likely to have firing rates that rise during motor planning, and are more likely to have firing rates that rise around the time of movement onset.

Interneurons are more responsive than pyramidal neurons

The finding that interneurons are more strongly modulated across conditions than pyramidal neurons is consistent with their larger dynamic range (Connors and Gutnick 1990). More surprising was the finding that putative interneurons are more likely to have firing rates that rise during movement planning, while putative pyramidal neurons showed more symmetric firing rate changes. To put this another way, for interneurons most conditions (most reach directions / reach paths) caused an increase in firing rate, while for pyramidal neurons, conditions that caused suppression were almost as common as those that caused excitation. This is perhaps remarkable – one usually supposes that overall activity goes up during motor planning – yet our results indicated the average rate of a population of pyramidal neurons rises only slightly. Most of the increase in overall rate (and what is perhaps detected with lower-resolution methods such as fMRI) is contributed by interneurons. The functional significance of the rise in inhibition, but not in excitation, is unclear. One speculation is that as incoming excitation rises, the relative
rise in internal inhibition may serve to maintain a balance of excitation and inhibition (Shadlen and Newsome 1998; van Vreeswijk and Sompolinsky 1996). Physiologically, the asymmetry may arise because many interneurons have chloride reversal potentials near threshold (Martina et al. 2001). If such neurons are not readily hyperpolarized by GABA, they may be less likely to show declines in firing rate.

Lack of support for output gating by inhibition

Contrary to what is expected if inhibition served an output-gating function for PMd, putative interneurons were actually most active around movement onset. Indeed, of the two cell classes, interneurons showed the larger activity increase at that time. Thus, it does not appear that inhibition falls around movement onset, or that excitation outstrips inhibition. It was also not the case that interneurons tended to invert their direction / condition preference around movement onset, as might be expected if they switched from inhibiting the planned movement to inhibiting competing movements. Instead, interneurons had a broad distribution of correlations between their plan- and movement-period preferences across cells, with the distribution centered slightly above zero. That is, on average they neither maintained their preferences consistently nor switched their preferences consistently between planning and movement. Surprisingly, pyramidal neurons showed a similarly wide range of correlation values. This wide range of correlations is again inconsistent with the output-gating hypothesis, which predicts that the pattern of pyramidal cell plan-period activity should resemble the movement-period activity (merely at sub-threshold levels). Thus, it does not appear that premotor cortex simply creates a pattern of activity in the excitatory neurons while output cells are kept inhibited until the go cue.
Alternative mechanisms for preventing plan activity from driving movement

Some mechanism would appear to be necessary to prevent plan period activity from causing undesired movements. One possibility is that a small subset of interneurons might act as gates, while most have some other function. However, we did not see support for this in the form of a subset of cells with a pause-like activity pattern (high plan-period firing rates and low movement-period firing rates). Instead, regarding their tendency to pause during movement, we found a unimodal distribution across neurons (Fig. 7 bottom row).

A second possibility is that PMd may make feedforward connections primarily on inhibitory neurons in M1, essentially making the rest of M1 the ‘output’ neurons in Fig. 1. There is physiological evidence that some pyramidal neurons in PMd synapse primarily on inhibitory cells in M1, producing feedforward inhibition (Ghosh and Porter 1988; Keller 1993; Tokuno and Nambu 2000). However, we did not find a clear subset of pyramidal neurons with pause-like activity patterns. The feedforward inhibition hypothesis also makes the concrete prediction that we should find pause-like neurons in M1, which presents a good target for future study.

Another possibility is that gating may occur purely downstream of PMd. Local inhibition in M1 could prevent it from responding during planning, and then be released before movement. Other work has suggested that cancellation of movements, a form of gating, could involve lateral inhibition in M1 (Riehle et al. 2006). However, recent work has shown that in M1, as in PMd, inhibition rises rather than falls around the time of movement onset (Merchant et al. 2008).
An intriguing alternative is that the motor system does not use a straightforward mechanism of inhibitory gating at all. Indeed, a blanket output-gating mechanism might be ill-suited to real-world reaching behavior. One must typically plan one movement while executing another, or while maintaining a posture (e.g., in our task the arm had to be supported against gravity). If a feedforward gate exists, it seems it must therefore be movement-specific. The connectivity of PMd also argues against the presence of a blanket output gate: PMd projects to primary motor cortex (M1) and spinal interneurons but not to lower motor neurons (Dum and Strick 2002). Because both M1 neurons and spinal interneurons show some plan-period activity (Prut and Fetz 1999), PMd should be expected to send some signals down the spinal cord and into M1 even during the plan period. Such activity presumably modulates reflexes or otherwise prepares the downstream motor system. Lastly, the mean firing rate across pyramidal neurons barely changed during the planning period. It is thus possible that planning occurs in such a way that there is no ‘excess’ output to gate.

Therefore, instead of a gated oculomotor-like system such as that illustrated in Fig. 1, we might consider PMd and M1 as forming a dynamical system for controlling the arm (Churchland et al. 2006b; Cisek 2006b; Fetz 1992; Scott 2004; Todorov and Jordan 2002). Since there are many more neurons than muscles, neural activity would likely be higher dimensional than the muscle activity it controls. If so, many directions in the high-dimensional ‘neural space’ would fall in the ‘null space’ of the muscles: changes in neural activity along those dimensions would not produce changes in muscle activity (Churchland et al. 2007; Yu et al. 2009). An output-gating mechanism would not be required. Such a system would also not be obliged to possess elements with consistent
‘tuning’ during the plan and movement epochs (or even within the movement epoch; Churchland et al. 2006b; Fu et al. 1995; Hatsopoulos et al. 2007). Our findings are consistent with this view, but they fall well short of an explicit test. Future work will have to determine how to generate specific predictions from the dynamical systems / feedback-control framework.

Summary

We found a number of differences between the responses of putative interneurons and putative pyramidal neurons. Two findings present a potential challenge to our current conceptualization of PMd function. First, we found no evidence that interneurons in PMd act to ‘gate’ outputs. It thus remains unclear how, in the absence of movement, plan-period activity can be so prevalent in neurons that also appear actively involved in generating movement. Second, we found that most neurons in PMd have cross-condition modulation that is inconsistent between the plan and movement epochs. It is possible that a dynamical systems / feedback-control framework can account for these findings, although it seems just as possible that the correct theoretical framework has yet to be developed.
Neural recordings. We made neural recordings using single electrodes in monkey J and an implanted multi-electrode array in monkey H. For monkey J, single-unit recordings were made using tungsten single electrodes with 3-7 MΩ starting impedance (Part # UEWLGCEEN1E, Frederick Haer Company, Bowdoinham, ME) driven by a hydraulic microdrive (David Kopf Instruments, Tujunga, CA), as described previously (Churchland et al. 2006c). Electrode impedances were reduced to ~1 MΩ after piercing the dura. An effort was made to isolate neurons that were active during the plan period. Neural signals were amplified, filtered, and sorted using a Multichannel Acquisition Processor (MAP; Plexon, Dallas, TX). The signal path was: unity-gain buffer (head stage), 154 Hz 1 pole high-pass filter, 100x amplifier, 8.8 kHz 1 pole low-pass filter, 10x amplifier, 30 Hz 1 pole high-pass filter, digital 400 Hz 2 pole high-pass filter, digital 6 kHz 6 pole low-pass filter (Fig. 10). Before (or occasionally after) recording each new neuron, waveforms were also collected with the digital filters disabled. For most neurons (137), the recording site depth was estimated. The point at which the electrode entered cortex was determined by listening to the noise characteristics while lowering the electrode. For analyses involving depth, we excluded penetrations where this point was uncertain. 163 neurons were recorded in total, with an average of 14 trials per condition. For monkey H, signals were recorded from the implanted array using the Cerebus system (Blackrock Microsystems, Salt Lake City, UT) and sorted with the Sahani sorting algorithm (Santhanam et al. 2004) as described previously (Zumsteg et al. 2005). Clustering plots generated by the sorting algorithm assisted in identifying single units by
hand; identified multi-units were discarded. During performance of the task, standard filter settings were used (Fig. 10). Before recordings, two minutes of broadband data were also recorded (0.3 Hz 1 pole high-pass filter, 7.5 kHz 3 pole low-pass filter) for use in waveform classification. A single day of recording (H20041119), which yielded 71 single units, was used.

**EMG recordings.** EMG activity was recorded from monkey J using hook-wire electrodes (44 gauge with a 27 gauge cannula; Nicolet Biomedical, Madison, WI) placed in the muscle for the duration of single recording sessions. EMG was recorded for a subset of the maze conditions. Recordings were made from trapezius, latissimus dorsi, pectoralis, triceps brachii, medial and lateral aspects of the biceps brachii, and anterior, medial and posterior aspects of the deltoid. Recordings were made one muscle at a time, after completion of neural recording. Electrode voltages were amplified, bandpass filtered (150 –500 Hz, four pole, 24 db/octave), sampled at 1000 Hz, and digitized. Off-line, raw traces were differentiated (to remove any remaining baseline), rectified, smoothed with a Gaussian (SD of 15 ms) and averaged.

We verified that plan activity was not producing substantial anticipatory changes in muscle activity. EMG activity was typically unmodulated from the baseline period to the plan period, or in rare instances, very weakly modulated during the plan period. This is consistent with our previous verifications in similar tasks with previous animals (Churchland et al. 2006b; Churchland et al. 2006c).

**Classification of neuron types using single-electrode recordings.** To find each neuron’s mean waveform, we took up to 300 waveforms, spline interpolated them to 2.5 μs precision, then aligned each. Alignment was generally performed at the mid-point of
the slope between trough and peak, to minimize jitter for both the trough and peak. For

the very broadest-waveform neurons, this yielded poor alignment. Thus, when the
downward slope of the trough was > 1.5 times that of the upward slope of the trough, we
instead used the mid-point on the initial downward slope (see Supplemental Fig. 2, 3).

We found that this procedure yielded alignment as good or better than that provided by
alternate methods based on the trough minimum or trough center of mass. Following
alignment, we selected the cleanest waveforms for purposes of obtaining the average
waveform (spikes were not discarded for other analyses). To do so, we took the standard
deviation (SD) of the waveforms at each time point, then accepted only waveforms where
(1) the absolute difference from the mean was < 2 SDs at every time point, and (2) the
absolute mean difference from the mean across time was < 0.4 SDs. These values were
chosen by hand as eliminating most noisy and vertically offset waveforms. We averaged
these clean waveforms to get the mean waveform that was analyzed for each neuron. We
then separated out by hand neurons with mean waveforms whose shapes were not
canonical (e.g., were lacking a trough or a post-trough peak; plotted in Fig. 9A of
Results) and rejected neurons whose mean waveform after-potential was very flat and
therefore difficult to measure reliably. Rejections were performed blind to the trough-to-
peak duration (TTP) distribution and were performed prior to subsequent analyses. We
found a bimodal distribution with a TTP threshold (200 μs) for separating neuron types,
esentially identical to that in Mitchell et al. (2007).

Classification of neuron types using multi-electrode array recordings. We
extended the TTP method for classifying neuron types to data collected with the
Blackrock multi-electrode arrays. To our knowledge, neuron classes have not previously
been distinguished using these arrays. To classify these waveforms, we simulated the filters used with our single-electrode recordings, which we knew could yield good separation of narrow- and broad-spiking neurons. To simulate the single-electrode signal path, we took advantage of the broadband array recordings made before some experiments (Fig. 10). We re-filtered the original broadband data with digital filters to simulate the (non-digital) single-electrode high-pass filters (thick border in Fig. 10). Spike sorting was still based on the original filtered waveforms, and the waveforms from broadband were classified by re-filtering them to match the standard array filtering. This produced sorted waveforms with filtering similar to that for monkey J, where we found bimodality. This process does not yield a distribution of waveforms from the array with two clear peaks, though the distribution does appear to be bimodal (see Results). This blurring is presumably because of electrode tip geometry, lower impedance, filtering properties of the electrode tips, or other unknown properties of the arrays. Use of even broader filters (100 Hz 4 pole high-pass) did not yield cleanly separable peaks either. We do not know what properties of the arrays are responsible for blurring the bimodality of the distribution. However, based on simulations using our single-unit data, we found that even relatively small amounts of additional filtering destroyed the two-peaked distribution shape. That exercise also revealed that even with such a blurred distribution it was still possible to properly classify neurons (classification after additional filtering always agreed with that before), so long as one was willing to use a small exclusion zone (30 μs). We thus applied this zone to the array-recorded TTP data, making our thresholds < 185 μs and > 215 μs. On the assumption that the filtering
properties of the two electrode types (and/or the chronic versus acute recording environment) are unlikely to be exceedingly different, this should be conservative. Importantly, we note that the key differences between neurons classified as narrow-spiking and neurons classified as broad-spiking were very similar between the single-unit dataset and the multi-electrode array dataset. This confirms that the choice of threshold and exclusion zone was reasonable. As a control, we subsequently repeated key analyses using different threshold choices, and found that the main effects were only weakly sensitive to the exact choice of threshold.
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**Figure legends**

**Figure 1.** The output-gating hypothesis. *A.* Cartoon of cortical connectivity. Neurons marked P are local pyramidal neurons, IN are interneurons, CS is a corticospinal neuron (also pyramidal). Blue synapses are excitatory, red are inhibitory. Gray connection exists, but is not central to the output-gating hypothesis. *B.* Hypothetical PSTHs. TARG indicates target onset, MOVE indicates movement onset. Colors correspond to neuron types above. Superficial pyramidal neurons might be expected to build up activity during planning that is amplified at movement onset through recurrence. Interneurons might inhibit output neurons during planning, and later permit activity to flow through to the spinal cord during movement. Corticospinal neurons might then be almost exclusively active during movement.

**Figure 2.** Relative locations of narrow- and broad-spiking neuron recordings. *A.* Location of the implanted electrode array in monkey H. Nearby sulci are shown with black lines. Red dots indicate narrow-spiking cells, blue dots indicate broad-spiking neurons. Note that some electrodes recorded two isolable neurons, indicated by two dots in the box for that electrode. *B.* Approximate location of the region of single-electrode recordings in monkey J. *C.* Expanded view of single-electrode penetration locations for monkey J. Blue and red are as in panel A, black dots indicate non-canonical waveform neurons. Dots are scattered < 0.1 mm to reveal overlapping recordings. *D.* Counts of each neuron type by
depth for monkey J. Red and blue are as in other panels; gray indicates neurons with non-
canonical waveforms. Neurons unclassifiable due to flat peaks not shown.

**Figure 3.** Delayed reach tasks. Each row illustrates a single trial, taking 2-3 seconds. Top
row illustrates task for monkey H, bottom row for monkey J. Monkey H touched targets
directly with his fingers; monkey J also touched the screen but contacted the targets with
a cursor floating 2.5 cm above his hand. The animals first fixated (eye and hand/cursor)
fixation cues (> 400 ms), after which a target appeared. After a plan period (H: 200-1000
ms; J: 0-1000 ms), the go cue was given. For monkey H, the go cue was indicated by a
slight enlarging of the target; for monkey J, the target jittered slightly during the plan
period and the cessation of jittering indicated “go.” In both cases the fixation point was
also extinguished at the time of the go cue. For monkey J, most trials also involved
barriers which appeared at the same time as the target, instructing a curved reach.

**Figure 4. A.** Histogram of the trough-to-peak (TTP) duration for monkey J. Inset shows
amplitude-normalized mean waveforms. Red indicates narrow-spiking neurons, blue
indicates broad-spiking neurons. **B.** Same for monkey H using simulated MAP filters on
broadband data from the chronic electrode array. Red and blue are as above, green
indicates neurons considered unclassifiable. Special steps were taken to address the lack
of two clear peaks in this distribution (see Appendix), and subsequent analyses yield very
similar results for both the single-electrode data and the electrode array data.
Figure 5. PSTHs of example PMd neurons (one neuron per panel). Each trace plots the average firing rate for one condition from that neuron. Insets show the output-gating model’s predictions from Fig. 1. All neuron classes were highly heterogeneous, but PSTHs pictured represent common response patterns. During planning, pyramidal neurons often increased their firing rates for some conditions and decreased their firing rates for others. Interneurons’ firing rates typically increased during the plan period, and they tended to have their highest firing rates around the time of movement onset. Deep neurons, which may project to other brain areas or the spinal cord, tended to be less strongly modulated across conditions during the plan period but were typically quite active during the movement. 20 ms SD Gaussian smoothing used. Neurons from monkey J (P17, P33, P145).

Figure 6. Comparison of response properties between narrow- and broad-spiking neurons. Changes in firing rate for monkey J (left); monkey H (right). Top row: mean firing rate across conditions. Red trace shows the average firing rate across all narrow-spiking neurons across all conditions (both preferred and non-preferred). Blue trace shows the same for broad-spiking neurons. These plots are essentially population PSTHs. Bottom row: Red trace shows the cross-condition modulation (most preferred minus least preferred condition at every time point) for narrow-spiking neurons. Blue trace is the same for broad-spiking neurons. These plots are effectively a non-parametric ‘depth of tuning’ over time. Note that treating time points independently causes baseline cross-condition modulation to be greater than zero due to noise. TARGET is target onset time; MOVE is movement onset time. Flanking traces are SEMs across neurons.
Figure 7. Cell-by-cell analysis of firing rates. Distributions for monkey J (left); monkey H (right). This analysis summarizes patterns from Fig. 6 for individual neurons. Top row: For each neuron, the mean plan-period firing rate (FR) is taken and the mean baseline-period FR is subtracted. The histogram over neurons is plotted. Red indicates narrow-spiking neurons, blue indicates broad-spiking neurons. Note that the distributions are unimodal and nearly symmetrical, and that the distribution for narrow-spiking neurons is shifted to the right of that for broad-spiking neurons. That is, narrow-spiking neurons tended to have higher FRs during planning than during baseline. This shift was significant for both monkeys (see Results). Middle row: Same for mean movement-period FR minus mean baseline-period FR. Note similar shift as for the plan period above. This shift was significant for both monkeys. Bottom row: mean plan-period FR minus mean movement-period FR. High values indicate a pause-like neuron. The broad- and narrow-spiking distributions were not significantly different for either monkey.

Figure 8. Relationship between plan and peri-movement cross-condition modulation patterns. Top: PSTHs for three example neurons, all broad-spiking (monkey J P59, P57, P23). For these examples, the correlations between the plan and peri-movement cross-condition modulation patterns were -0.53 (left), 0.17 (center), and 0.86 (right). Colors correspond to different conditions. Conditions selected are representative; not all are shown. Bottom: Distribution of correlation values. A near-zero correlation means that the neuron showed little relationship between its plan and peri-movement modulation patterns. A negative value implies inverted preferences during the two epochs.
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bottom: Histogram of correlations for narrow-spiking neurons; right bottom: for broad-spiking neurons. Neurons are pooled across monkeys: monkey J (H) contributed 33 (8) narrow-spiking neurons and 79 (4) broad-spiking neurons (note that many of monkey H’s neurons had shallow cross-condition modulation for one epoch and were thus excluded from this particular analysis; see Results). Black arrows are distribution means. For narrow-spiking neurons, the mean for monkey J (H) is 0.26 (0.16), and for broad-spiking neurons the mean is 0.17 (0.09). Gray arrows are bootstraps (see Results).

Figure 9. Analysis of ‘non-canonical waveform’ neurons (all from monkey J). A. Normalized waveforms. B. Mean firing rate across conditions and neurons over time (same format as Fig. 6 top left). C. Cross-condition modulation (preferred - least preferred condition, same format as Fig. 6 bottom left). Black lines show responses of non-canonical neurons. Flanking traces are SEMs.

Figure 10. Signal paths for single electrode (MAP) recordings (top) and multi-electrode array recordings (bottom). Key filters marked with heavy outline. HP: high pass. BP: band pass. ADC: analog-digital converter. Bracket under top waveforms illustrates how trough-to-peak waveform duration is measured.
References


Cisek P. Preparing for speed. Focus on "Preparatory activity in premotor and motor cortex reflects the speed of the upcoming reach". *J Neurophysiol* 96: 2842-2843, 2006b.


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A

To spinal cord

B

Firing rate

Pyramid

Interneuron

Corticospinal neuron

TARG

MOVE

TARG

MOVE

TARG

MOVE
Acquire touch
Acquire fix

Target cue
Plan period

Go cue

Movement period
Target acquired

Monkey H

Monkey J
monkey J
mean PSTH
(mean across all conds)
monkey H

narrow-spiking
broad-spiking

spikes/s
-400 TARGET 400 -400 MOVE 600
25
25
15

-400 TARGET 400 -400 MOVE 600
25
0

-400 TARGET 400 -400 MOVE 600
0

spikes/s
-400 TARGET 400 -400 MOVE 600
40
25
15
5

-400 TARGET 400 -400 MOVE 600
40
25
15
5

cross-condition modulation
(pref - least pref)
monkey J

monkey H

# of neurons

plan - baseline (spikes/s)

# of neurons

move - baseline (spikes/s)

# of neurons

plan - move (spikes/s)

# of neurons

plan - move (spikes/s)

-20 -10 0 10 20 30

-20 -10 0 10 20 30

-20 -10 0 10 20 30

-20 -10 0 10 20 30

-10 0 10 20 30

-10 0 10 20 30

-10 0 10 20 30

-10 0 10 20 30

-60 -40 -20 0 20 40 60

-60 -40 -20 0 20 40 60

-60 -40 -20 0 20 40 60

-60 -40 -20 0 20 40 60

narrow-spiking

broad-spiking

-60 -40 -20 0 20 40 60

-60 -40 -20 0 20 40 60

-60 -40 -20 0 20 40 60

-60 -40 -20 0 20 40 60

-40 -20 0 20 40 60

-40 -20 0 20 40 60

-40 -20 0 20 40 60

-40 -20 0 20 40 60

-20 -10 0 10 20 30

-20 -10 0 10 20 30

-20 -10 0 10 20 30

-20 -10 0 10 20 30

-10 0 10 20 30

-10 0 10 20 30

-10 0 10 20 30

-10 0 10 20 30

-40 -20 0 20 40

-40 -20 0 20 40

-40 -20 0 20 40

-40 -20 0 20 40

-20 0 20 40

-20 0 20 40

-20 0 20 40

-20 0 20 40

-10 0 10 20 30

-10 0 10 20 30

-10 0 10 20 30

-10 0 10 20 30

0 20 40

0 20 40

0 20 40

0 20 40

# of neurons

plan - baseline (spikes/s)

plan - baseline (spikes/s)
JHH J Bootstraps

narrow-spiking broad-spiking

$r = -0.53$

-400 TARG 400 -400 MOVE 600

$0$ $100$

$0$ $140$

Correlation (r)

$0$ $6$

$3$

# of neurons

spikes/s

$0$ $100$

$0$ $140$

Correlation (r)

$0$ $6$

$3$

# of neurons

r = 0.17

-400 TARG 400 -400 MOVE 600

$0$ $100$

$0$ $140$

Correlation (r)

$0$ $6$

$3$

# of neurons

spikes/s

$0$ $100$

$0$ $140$

Correlation (r)

$0$ $6$

$3$

# of neurons

r = 0.86

-400 TARG 400 -400 MOVE 600

$0$ $100$

$0$ $140$

Correlation (r)

$0$ $6$

$3$

# of neurons

spikes/s

$0$ $100$

$0$ $140$

Correlation (r)

$0$ $6$

$3$

# of neurons

narrow-spiking H J Bootstraps

broad-spiking H J Bootstraps
Single electrode

1x buffer

154 Hz 1-pole HP filter

100x amp

Broad BP filters, 10x amp

ADC

Digital BP filters (incl. 400 Hz 2-pole HP)

Single-unit waveforms

Head stage, incl. broad filters

ADC

Save broadband data

Cerebus "spike medium" filters: 250 Hz 4-pole HP

ADC

Save standard data

Simulate "spike medium" filters

Classify events (spike sort)

Sorted MAP-filtered waveforms

Simulate MAP filters (154 Hz 1-pole HP)