Periodic modulation of repetitively elicited monosynaptic reflexes of the human lumbosacral spinal cord

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Running head: Periodic modulation of repetitively elicited spinal reflexes

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Abstract

In individuals with motor-complete spinal cord injury, epidural stimulation of the lumbosacral spinal cord at 2 Hz evokes unmodulated reflexes in the lower limbs, while stimulation at 22–60 Hz can generate rhythmic burst-like activity. Here, we elaborated on an output pattern emerging at transitional stimulation frequencies with consecutively elicited reflexes alternating between large and small. We analyzed responses concomitantly elicited in thigh and leg muscle groups bilaterally by epidural stimulation in eight motor-complete spinal cord injured individuals. Periodic amplitude-modulation of at least 20 successive responses occurred in 31.4% of all available data sets with stimulation frequency set at 5–26 Hz, with highest prevalence at 16 Hz. It could be evoked in a single muscle group only, but was more strongly expressed and consistent when occurring in pairs of antagonists or in the same muscle group bilaterally. Latencies and waveforms of the modulated reflexes corresponded to those of the unmodulated, monosynaptic responses to 2-Hz stimulation. We suggest that the cyclical changes of the reflex excitability resulted from the interaction of facilitatory and inhibitory mechanisms emerging after specific delays and with distinct durations, including post-activation depression, recurrent inhibition and facilitation as well as reafferent feedback activation. The emergence of the large responses within the patterns at a rate of 5.5 or 8 per second may further suggest the entrainment of spinal mechanisms as involved in clonus. The study demonstrates that the human lumbosacral spinal cord can organize a simple form of rhythmicity through the repetitive activation of spinal reflex circuits.

Key words: human, repetitive nerve stimulation, rhythm generation, spinal cord stimulation, spinal reflexes
Introduction

The soleus Hoffmann reflex (H reflex) is widely used in human neurophysiological studies that explore the organization of spinal reflexes and the sensorimotor integration in neural control of movement (Capaday and Stein 1986; Schiepatti 1987; Pierrot-Deseilligny and Mazevet 2000; Knikou 2008; Pierrot-Deseilligny and Burke 2012). It is evoked by electrical stimulation of large-diameter Ia afferents from muscle spindles in the posterior tibial nerve with subsequent recruitment of motor neurons largely through monosynaptic connections in the spinal cord (Magladery and McDougal 1950; Pierrot-Deseilligny and Burke 2012).

Changes in the H-reflex amplitude caused by conditioning inputs indicate modulation of the excitability of the reflex pathway by peripheral influence and central circuitry (Knikou 2008; Pierrot-Deseilligny and Burke 2012).

Each sensory input volley to the spinal cord elicited by stimulation of the tibial nerve not only evokes the H reflex, but also initiates central processes that modify the responsiveness to subsequent stimulus-induced input (Magladery and McDougal 1950; Crone and Nielsen 1989). When the tibial nerve is repetitively stimulated, there is a progressive fall in the amplitudes of the H reflexes (Ishikawa et al. 1966; Schindler-Ivens and Shields 2000). Conditioning inputs through stimulation of other peripheral nerves activate spinal circuits that are otherwise not engaged and lead to different patterns of H-reflex modulation (Mao et al. 1984; Meunier et al. 1993; Pierrot-Deseilligny and Burke 2012). Such conditioning-test paradigms have described various time-dependent facilitatory or inhibitory processes, based on the widespread spinal inter-connections between the reflex circuits of different motoneuron pools (Delwaide et al. 1976).

Applying sensory inputs to several lumbar and upper sacral spinal cord segments simultaneously became possible with the application of epidural spinal cord stimulation (SCS)
for the treatment of upper motor neuron disorders (Cook and Weinstein 1973), and the specific electrode placement required for the control of lower-limb spasticity (Pinter et al. 2000). When repetitively applied, such simultaneous, bilateral stimulation of multiple posterior roots produces motor outputs in the lower limbs of individuals with motor-complete spinal cord injury (SCI) that cannot be evoked by stimulation of a single peripheral nerve alone (Dimitrijevic et al. 1998; Shapkova and Schomburg 2001; Gerasimenko et al. 2002; Jilge et al. 2004; Danner et al. 2015). The SCS-elicited motor outputs are comprised of series of stimulus-pulse related posterior root-muscle (PRM) reflexes, i.e., large-afferent-induced spinal reflexes initiated in the posterior roots and recorded by surface-electromyography (EMG) from many lower-limb muscles (Minassian et al. 2004). Stimulation at 2 Hz elicits unmodulated, monosynaptic PRM reflexes (Murg et al. 2000), while 22–60 Hz can lead to rhythmic burst-like motor activity with waxing and waning of the PRM reflexes, modulated by concomitantly activated spinal interneuronal mechanisms (Dimitrijevic et al. 1998; Minassian et al. 2004; Danner et al. 2015; cf. Wenger et al. 2014).

In the present work, we elaborate on an output phenomenon encountered when applying intermediate SCS frequencies of 5–26 Hz. Instead of leading to a progressive reflex depression or random variability in reflex size as would be suggested by H-reflex studies, the repetitive stimulation often produced a pattern with the amplitudes of the successive responses alternating between large and small. Here, we provide a detailed description of this modulation pattern, including its prevalence, the dependence on the applied stimulation parameters, and observations hinting on the coupling of reflexes evoked in pairs of muscle groups. The cycle frequency of this periodic reflex modulation is well above the range of burst frequencies adequate for locomotion (Danner et al. 2015) and hence renders the involvement of the spinal locomotor rhythm generator unlikely. This study therefore adds to
the understanding of how the human lumbar spinal reflex circuits can process repetitive inputs to regulate and sustain a simple form of oscillating motor behavior.

Materials and Methods

Subjects

Data derived from eight individuals with traumatic, clinically motor-complete SCI in the chronic stage of recovery were analyzed (Table 1). The subjects’ neurological status was evaluated according to the International Standards for Neurological Classification of Spinal Cord Injury (Kirshblum et al. 2011). They had epidural SCS systems implanted for the control of spinal spasticity affecting the lower extremities (Pinter et al. 2000). Informed, written consent was obtained from the subjects prior to their participation. All procedures were approved by the Ethics Committee of the City of Vienna and conformed to the Declaration of Helsinki.

Epidural spinal cord stimulation

Epidural SCS was applied via a cylindrical electrode lead (Pisces-Quad electrode, Model 3487A, Medtronic, Minneapolis, MN, USA) that was connected to a programmable pulse generator (Itrel 3, Model 7425, Medtronic) placed subcutaneously in the abdominal wall. The lead carried four electrodes, referred to as 0 to 3 from rostral to caudal direction, each with a diameter of 1.3 mm, a length of 3 mm, and an inter-electrode distance of 6 mm. The electrodes were positioned longitudinally in the dorsal epidural space, targeting the lumbosacral spinal cord. Vertebral levels ranged from the lower half of T11 to the lower third of L1 across the eight subjects. Each electrode could be independently set to +, −, or ‘off’, allowing for various bipolar electrode combinations. Monopolar stimulation was carried out
by selecting one of the epidural electrodes as cathode and the active area of the implanted pulse-generator case (‘c’) as anode. The pulse generator delivered capacitively decoupled monophasic rectangular constant-voltage pulses of 210 µs width, each followed by an exponentially decaying phase to adjust charge balance for each stimulus and avoid delivery of direct current (Rattay et al. 2000). Programmable stimulation frequencies and intensities were 2–130 Hz and 0–10.5 V.

The rostrocaudal position (Pos) of the selected cathode relative to the spinal cord segments was physiologically estimated based on the thresholds of the PRM reflexes elicited at 2 Hz in the L2–L4 innervated quadriceps (Q) and the L5–S2 innervated triceps surae (TS), respectively (Murg et al. 2000; Minassian et al. 2007a): Pos 1, Q but not TS recruited with maximum applied stimulation intensity, active cathode position estimated to be rostral, yet close, to the L2-spinal cord segment; Pos 2, lower thresholds for Q than TS responses, cathode position over L2–L4 segments; Pos 3, same thresholds for Q and TS, cathode position over L5–S2 segments; Pos 4, lower thresholds for TS than Q, cathode position caudal to the S2 segment.

Surface poly-electromyography

EMG activity was recorded from Q, hamstrings (Ham), tibialis anterior (TA), and TS bilaterally with pairs of silver-silver chloride surface electrodes (Intec Medizintechnik GmbH, Klagenfurt, Austria) with an inter-electrode distance of 3 cm (Sherwood et al. 1996). Additional EMG recordings were taken from the lumbar paraspinal trunk muscles to monitor stimulation artefacts. Abrasive paste (Nuprep, Weaver and Company, Aurora, CO, USA) was used for skin preparation to reduce contact resistance. EMG signals were amplified using a Grass 12D-16-OS Neurodata Acquisition System (Grass Instruments, Quincy, MA, USA) adjusted to a gain of 2000 over a 3-dB bandwidth of 30–700 Hz. Data were digitized at 2002
samples per second and channel using a Codas ADC system (Dataq Instruments, Akron, OH, USA).

Stimulation protocol

For a given selection of active electrodes, stimulation was initially applied at 2 Hz and intensity gradually increased in 1-V increments until PRM reflexes were elicited. At this intensity, the stimulation frequency was increased step-wise, including 5, 11, 16, 22, and 26 Hz. The frequency variation was then repeated for intensities up to 10 V. Stimulation intensity and frequency variation were subsequently repeated for different epidural electrode combinations. All recordings were conducted with the subjects relaxed, lying comfortably supine.

Data analysis and statistics

Data were analyzed offline using Matlab R2012a (The MathWorks, Inc., Natick, MA, USA). Statistical analyses were performed with SPSS 22 for Mac OS X (10.8.5), and α-errors of P<.05 were considered significant. For Student’s t-tests, the assumptions of normality and equality of variances were verified.

Recordings obtained during SCS at 2, 5, 11, 16, 22, and 26 Hz were selected for analysis. The stimulation elicited PRM reflexes in Q, Ham, TA, and TS that were electromyographically recorded as stimulus-pulse related reflex-compound muscle action potentials (Minassian et al. 2004). The number of PRM reflexes collected with unchanged parameter settings ranged from 20 to approximately 1600 per muscle group studied, also depending on the applied stimulation frequency. The EMG of individual PRM reflexes was analyzed for 50 ms post-stimulus. Peak-to-peak amplitudes of the PRM reflexes were calculated.
For the detection of patterns characterized by an amplitude alternation of successively elicited reflexes between large and small, henceforth referred to as periodic reflex modulation, EMG sections under unchanged stimulation conditions were sequenced into shorter segments. The first segment contained the responses to the first 20 stimuli, and the respective next segments were each shifted by one stimulus (i.e., segment 2, responses to stimuli 2–21; segment 3, responses to stimuli 3–22, etc.). For each muscle group and recording segment, the first order differences of the peak-to-peak amplitudes of pairs of consecutive responses were calculated (response$_2$-response$_1$, response$_3$-response$_2$, …, response$_{20}$-response$_{19}$) to determine the signs of the 19 resulting differences. A steady pattern of periodic modulation was identified for a given 20-response segment, if the complete series of 19 signs was alternating, i.e., either +,−,+,−, etc. or −,+−, +, etc. The upper limit of the α-error considering the largest sample size of 1600 responses per muscle is 0.003. To compare the prevalence of periodic reflex modulation between muscle groups, a Pearson’s $\chi^2$ test was used. We introduced the consistency of modulation to describe the prevalence of the periodic modulation within each recording section with given SCS parameters per muscle. It was calculated as the number of 20-response segments with steady pattern relative to the total number of segments into which the recording section had been sequenced. A consistency of e.g. 50% would indicate that half of the 20-response segments of a recording section had steady patterns. To calculate the attenuation, the ratios of the mean peak-to-peak amplitudes of the 10 smaller and the 10 larger PRM reflexes within each patterned segment were calculated and averaged for each muscle group and recording section. Following the hypothesis that the attenuation correlated with the consistency, a Spearman's Rank Order Correlation test was performed for each muscle group and stimulation frequency.

The phase-relation of periodic reflex modulation concomitantly occurring in 2 or 4 muscle groups unilaterally or in pairs of muscle groups bilaterally was determined for each patterned
20-response segment by comparing the signs of the respective 19 first order differences calculated. Synchronous modulation was identified if the series started with the same sign, and reciprocal modulation otherwise. Fluctuations were detected if at least one 20-response segment each with synchronized as well as reciprocal modulation occurred within a recording section. Consistency and attenuation of periodic modulation occurring in one muscle group only were compared to the respective values when these patterns occurred concomitantly with modulation in its antagonist or in the same group on the contralateral side using a Student’s t-test.

For the responses of each muscle group to 2-Hz SCS, the occurrence of post-activation depression was tested by comparing the amplitude of the PRM reflex elicited by the first stimulus to the mean amplitudes of the responses to the 5th–14th stimulus pulses using a paired Student’s t-test.

Onset latencies were determined for PRM reflexes elicited by 2-Hz SCS as well as for those constituting patterns with periodic reflex modulation as the times between the stimulus artefact and the first deflection from baseline exceeding 5% of the respective peak-to-peak amplitude. For each muscle group, mean onset latencies were calculated from all responses within a recording section with unchanged stimulation conditions. For each subject, these mean values were then averaged per muscle group, separately for the responses to 2-Hz SCS and for the periodically modulating ones. Group results were then obtained by averaging the corresponding mean values of the eight subjects. Mean latencies of responses to 2-Hz SCS and of the periodically modulating ones were compared using Student’s t-tests. Further, for each muscle group and epidural electrode combination, threshold intensities were defined as the minimum intensity required to consistently elicit responses at 2 Hz with amplitudes greater than 50 µV.
Results

Epidural SCS at 2–26 Hz evoked series of stimulus-locked PRM reflexes in the lower-limb muscle groups of the eight subjects studied. Stimulation at 2 Hz elicited highly repeatable responses (Fig. 1A, 2 Hz) with no significant change in reflex size within the series of PRM reflexes (paired Student’s t-test; Q, difference: 31.02±201.10 µV, t_{101}=1.550, P=.124, r=.15; Ham, difference: 6.40±301.17 µV, t_{101}=0.215, P=.830, r=.02; TA, difference: -1.986±104.73 µV, t_{72}=0.162, P=.871, r=.02; TS, difference: -30.890±349.84 µV, t_{85}=0.819, P=.415, r=.09).

When SCS was applied at 5–26 Hz, patterns emerged in which PRM-reflex amplitudes alternated between large and small (Fig. 1A, 22 Hz). This periodic reflex modulation could occur either in a single or in several uni- or bilateral muscle groups. Of the 1374 recording sections available (each with unchanged stimulation parameters) at 5–26 Hz, 431 (31.4%) included segments with periodic amplitude modulation. Characteristically, these patterns had short build-up phases (Fig. 1B), with 26.2% already established with the first pair of stimuli and additional 32.5% starting within the first ten stimuli applied. Another 16.9% of the patterns were established within the 11th to 30th stimulus. The remaining 24.4% cases appeared after the 30th stimulus pulse. Across all patterned EMG segments of the various muscle groups and subjects, even the larger of the alternating responses (0.84±0.85 mV) were significantly smaller than the unconditioned initial responses to the first stimulus of the train of stimuli applied during a given recording section (1.44±1.12 mV; Student’s t-test; difference: -0.6±0.69 mV, t_{399}=-17.314, P<.00001, r=.65); muscle-specific ratios were Q, 0.55±0.36; Ham, 0.75±0.35; TA, 0.54±0.31; and TS, 0.46±0.30.

Stimulation-parameter dependence of periodic reflex modulation
Periodic reflex modulation was most likely to be elicited at 16 Hz across all muscle groups studied, and its prevalence rapidly decreased when lower or higher stimulation frequencies were applied (Fig. 2A). Stimulation frequencies producing the patterns ranged from 5–26 Hz for Q and Ham and from 11–22 Hz for TA and TS, respectively. Across muscle groups, the prevalence of recording sections with invariant stimulation parameters containing at least one patterned segment decreased with increasing stimulation intensities (Fig. 2B). There was no specific relationship between the site of stimulation and the occurrence of the patterns, except for stimulation from Pos 4 that was least effective (Fig. 2C). Overall, the prevalence of the periodic reflex modulation (Fig. 3A) was higher in the proximal muscles (Q: 27.7% of the available data sets; Ham: 29.1%) than in the distal muscles (TA: 18.2%; TS: 21.0%; \( \chi^2(3)=13.617, P=.0035, \text{odds ratio}=1.44 \)).

Also, the consistency of the periodic reflex modulation was higher in the thigh than in the leg muscles (Fig. 3B). In more than 50% of all recording sections of Q or Ham with periodic reflex modulation, more than half of the successively elicited PRM reflexes alternated between large and small, while this was not the case in the distal muscles. A consistency of even 100%, i.e., modulation occurring throughout a recording section, was found in all muscle groups studied and occurred most commonly in Ham.

The mean consistency was stimulation-frequency dependent and peaked at 16 Hz for Q, Ham, and TS, and at 11-Hz SCS for TA (Fig. 4A). The attenuation of the small-amplitude PRM reflexes relative to the larger ones ranged from 0.42–0.86 for the different stimulation frequencies, with the strongest attenuation found at 16-Hz SCS across muscles (Fig. 4B). Further, in Q and Ham, the attenuation correlated with the consistency of the patterns (Q, \( \rho=-.677, P=.00061 \); Ham, \( \rho=-.436, P=.039 \)). With other words, the stronger the relative attenuation of the smaller responses, the higher was the prevalence of patterned segments to
occur within a given recording section. No such statistical relationship was found for the leg muscle groups (TA, $\rho=-.381$, $P=.171$; TS, $\rho=-.364$, $P=.133$).

**Coupling of reflex modulation in pairs of related muscle groups**

Periodic reflex modulation could be generated in one muscle group only (Fig. 5A(i)), but had a 2.0-times higher prevalence in a given muscle when the amplitude modulation concomitantly occurred in its antagonist (cf. Table 2, top). The periodic reflex modulation in antagonistic muscle groups occurred with reciprocal (Fig. 5A(ii)) and synchronized relationship (Fig. 5A(iii)). Both output relationships were produced with similar prevalence in Q and Ham, while in TA and TS, synchronized modulation was almost exclusively elicited (Table 2, top). Fluctuations between reciprocal and synchronized modulation were exceptions.

Similarly, periodic reflex modulation in a given muscle occurred with a 1.7-times higher prevalence when such pattern was concomitantly produced in the same muscle group on the contralateral side (Fig. 5B). These patterns were most commonly inter-related by synchronous modulation in Ham, TA, and TS, whereas in Q, reciprocal and synchronous modulation had similar prevalence (Table 2, bottom). The occurrence of the different types of patterns in pairs of related muscle groups is described in Table 3 for each subject.

The consistency of the periodic reflex modulation within a recording section of a given muscle as well as the relative attenuation of the smaller responses were significantly higher when elicited in pairs of either antagonistic (Fig. 6A) or bilateral (Fig. 6B) muscle groups compared to those patterns occurring in one muscle of the pair only (all $P<.00001$).

**Phase-relation of reflex modulation across all four unilateral muscle groups**

Periodic reflex modulation across all four unilateral muscle groups studied was detected in 3 of the 8 subjects (Table 3) and 16.7% of the 431 recording sections with patterned EMG
segments. The most frequent output relationships were those of reciprocal modulation of the
Q responses with respect to Ham, TA, and TS (7 examples, 38.9%; Fig. 7A) and of reciprocal
reflex modulation of Ham with respect to the Q, TA, and TS activity (6 examples, 33.3%; Fig.
7B). Synchronous modulation of the PRM reflexes in the proximal muscles occurring
reciprocally to the synchronized modulation of the responses in the distal muscles was found
in another 3 examples (16.7%). One example (5.6%) was found with synchronized
modulation of Q and TA responses occurring reciprocally to the synchronized modulation of
Ham and TS responses and one other example (5.6%) with synchronized response modulation
across all four muscle groups. No other output relationship was detected.

EMG waveforms and latencies of the modulated PRM reflexes

The vast majority of the amplitude-modulated PRM reflexes elicited at 5–26 Hz closely
resembled the unmodulated responses at 2 Hz. The responses to 2-Hz SCS had muscle-
specific EMG waveforms and short onset latencies amounting to Q, 9.5±0.9 ms; Ham,
10.0±0.6 ms; TA, 18.2±0.9 ms; and TS, 17.9±1.0 ms across subjects (Fig. 8A). The PRM
reflexes within the patterns with periodic modulation had the same waveforms in Q, in 99.3%
of all examples; Ham, 85.1%; TA, 63.9%; and TS, 91.3% (Fig. 8B). The onset latencies of
these PRM reflexes were Q, 10.2±0.7 ms; Ham, 11.1±1.2 ms; TA, 18.5±1.1 ms; and TS,
18.7±1.9 ms and did not differ from those of the responses to 2-Hz SCS (Student’s t-test; Q,
t7=1.928, P=.095, r=.59; Ham, t7=2.318, P=.054, r=.66; TA, t6=.533, P=.613, r=.21; TS,
t6=.800, P=.454, r=.31).

Discussion
Epidural stimulation of the lumbosacral spinal cord at 2 Hz evoked stimulus-pulse locked PRM reflexes in the lower-limb muscle groups that did not demonstrate post-activation depression, nor periodic modulation. When increasing the stimulation frequency to 5–26 Hz, patterns emerged after short build-up phases within the series of PRM reflexes with an alternation between larger and smaller response amplitudes. The prevalence of the periodic reflex modulation, its consistency and the relative attenuation of the smaller-amplitude responses depended on the applied stimulation frequency, maximizing at 16 Hz. While the amplitude modulation could be elicited in one muscle group only, its consistency and the relative attenuation were significantly higher when generated in pairs of related muscle groups. Patterns occurring in pairs of related muscle groups were coupled by synchronous modulation of the responses in both rather than reciprocating.

*Nature of the periodically modulated PRM reflexes*

The neural structures electrically stimulated by the longitudinal, cylindrical electrode type as employed here, placed in close distance to the posterior aspect of the lumbosacral spinal cord, are sensory fibers, while the immediate stimulation of motor fibers within the anterior roots can be ruled out even with the maximum available stimulation intensity (Rattay et al. 2000; Minassian et al. 2004, 2007a). Here, the vast majority of the periodically modulating responses had the same EMG waveforms and short onset latencies like the PRM reflexes evoked by 2-Hz SCS (cf. Murg et al. 2000; Minassian et al. 2004). These responses were previously suggested to be initiated in large-diameter afferents within the posterior roots/rootlets (Rattay et al. 2000) and transmitted via monosynaptic reflex arcs (Minassian et al. 2004, 2007a; cf. Capogrosso et al. 2013). They also closely resemble the lower-limb responses evoked by transcutaneous posterior-root stimulation (Minassian et al. 2007b, 2011; Hofstoetter et al. 2008, 2014; Ladenbauer et al. 2010). Physiologically, PRM reflexes elicited by single stimuli share similarities with the H reflex (Maertens de Noordhout et al. 1988),
such as attenuation by sustained tendon vibration (Minassian et al. 2007b, 2011) and motor-
task specific amplitude modulation (Courtine et al. 2007; Minassian et al. 2007b; Hofstoetter
et al. 2008). Also, when pairs of stimuli are delivered with an inter-stimulus interval of 50 ms,
both, the H reflex (Paillard 1959; Schiepatti 1987) as well as the PRM reflexes (Minassian et
al. 2004, 2007b; Roy et al. 2014) evoked by the second pulse are attenuated or completely
suppressed. Differences between these two types of reflexes become obvious when evoked by
trains of stimuli. The H-reflex size progressively decreases with increasing stimulation
frequencies from 0.1 to 10 Hz (Calancie et al. 1993; Schindler-Ivens and Shields 2000) and its
variability at these depressed levels appears to be random (Ishikawa et al. 1966). The different
behavior of the repetitively elicited PRM reflexes may arise from the synchronous stimulation
of afferents of several spinal cord segments bilaterally, the resulting mixed homo- and
heteronymous inputs and the recruitment of a wider range of interneurons, as well as the
concomitant elicitation of reflex responses in several lower limb muscle groups.

Entrainment of spinal reflex circuits by repetitive inputs and interaction with peripheral
events

Synaptic summation of inhibitory and excitatory actions with different time courses may have
been underlying the periodic PRM-reflex modulation. Such summation processes would
require at least one component to not have fully decayed by the time the next activation
arrives at the motoneuron pool, or secondary actions that affect the PRM-reflex transmission
only after a certain delay. Candidate mechanisms include post-activation depression,
presynaptic inhibition, recurrent inhibition and facilitation as well as reafferent feedback
activation following the muscle twitches.

Recurrent inhibition is mediated by Renshaw cells that respond to an activation with a burst of
spikes, lasting for 40–100 ms (Eccles et al. 1954; Hultborn et al. 1971a; Katz and Pierrot-
Deseilligny 1999). Axon collaterals of alpha motoneurons synapse on the Renshaw cells, which in turn project back to populations of homonymous and heteronymous motoneurons, including those that excited them (Pierrot-Deseilligny and Burke 2012). Renshaw cells would discharge following the motoneuronal activity of a PRM reflex evoked by a given stimulus pulse. An input volley elicited by a succeeding stimulus pulse that arrives to the motoneuron pool before full decay of the ongoing Renshaw-cell activity, as could be the case at e.g. 16-Hz stimulation, would recruit a smaller number of motoneurons. In turn, the smaller reflex discharge would activate fewer Renshaw cells, and the degree of recurrent inhibition upon the motoneuron pool would be reduced when the next stimulus pulse is applied. This process may eventually lead to a temporarily stable state of alternating degrees of recurrent inhibition exerted on the motoneuron pools and hence to the observed PRM-reflex modulation.

Another mechanism contributing to the PRM-reflex modulation could be a periodically emerging excitation superimposed upon a longer-lasting reduced reflex transmission. Such excitation could be caused by reafferent feedback activation induced by the substantial stimulation-evoked muscle twitch contractions and the subsequent relaxation. Reafferent feedback activation has been suggested to contribute to the facilitation of H reflexes elicited by double stimuli at inter-stimulus intervals of 100–400 ms (Paillard 1959; Schiepatti 1987; Kagamihara et al. 1998). Here, post-activation depression and presynaptic inhibition presumably led to a longer-lasting reduction in reflex transmission when SCS was applied at 5–26 Hz, since even the larger responses within the periodic modulation patterns were smaller than the unconditioned initial response to the first stimulus of the respective train. Thus, for a train of epidural stimulation delivered at a frequency around 16 Hz, the first pulse would set inhibitory mechanisms into action, leading to a smaller response to the second stimulus pulse. The afferent discharge following the muscle twitch contractions induced by the first pulse
would cause rebound volleys to the spinal cord that would arrive in time to facilitate the
response to the third stimulus (after 2 x 62.5 ms inter-stimulus interval = 125 ms). The weaker
contractions evoked by the second stimulus pulse would lead to less facilitatory impact on the
PRM reflex elicited by the fourth stimulus pulse, and eventually, an alternation between
smaller and larger PRM reflexes may develop.

Periodic reflex modulation occurred in about 30% of the recording sections of the proximal
and 20% of the distal muscle groups. This uneven distribution may in part result from
differences in efficacy of recurrent inhibition to motoneurons innervating proximal and distal
muscles. Contraction and relaxation times of the large thigh muscles together with their
shorter reflex arcs may result in different timing and patterns of rebound facilitation as
compared to the leg muscles. Differences in the secondary activation of muscle-spindle
afferents may also arise from muscle-receptor sensitivity and the range of twitch-related
movements. Further, different interactions between proximal and distal stretch reflexes
elicted in succession could either facilitate or impede the generation of the periodic reflex
modulation. Delwaide and colleagues (1976) demonstrated that the reflex activation of the
soleus muscle resulted in two distinct phases of facilitation of the quadriceps stretch reflex,
whereas the stretch reflex of quadriceps led to a long-lasting inhibition of the soleus reflex.

The occurrence of the periodic reflex modulation could be partially linked to the
reorganization of the spinal circuitry following SCI (Edgerton et al. 2001). These changes
together with an elevated central state of excitability alter the input processing by the neural
circuitry, which may increase the probability of the periodic reflex modulation to occur.

Further, some descending long-tract fibers or propriospinal connections may survive even
severe SCI clinically classified as motor complete (Sherwood et al. 1992; Kakulas 2004;
McKay et al. 2004) and may influence the modulation patterns produced across subjects.
The lack of attenuation of the PRM reflexes elicited by 2-Hz SCS may have resulted from the multi-root input provided by SCS together with alterations in the spinal circuits’ excitability after SCI (Ishikawa et al. 1966; Grey et al. 2008). At such stimulation frequency, the net result of inhibition and excitation may have evened each other out. Also, time-dependent mechanisms related to recurrent inhibition and rebound discharges would have ceased before the next response is elicited.

*Coupling of mechanisms underlying the reflex modulation in pairs of muscle groups*

Periodic PRM-reflex modulation generated in a single muscle group showed that the mechanisms modulating the excitability of the respective motoneuron pool did not require the occurrence of such pattern in other muscles (cf. Clair et al. 2011). Yet, prevalence, consistency and attenuation were significantly higher when the periodic patterns occurred in pairs of related muscle groups. This observation hints on interactions and mutual drive of oscillating or time-dependent mechanisms acting on separate motoneuron pools. Coupling effects are also put forward by the observation that not all possible phase-relations of reflex modulation between muscle groups occurred with same odds. Influences affecting different motoneuron pools may arise from inter-connections of myotatic reflex arcs (Delwaide et al. 1976; Meunier et al. 1990; Cheng et al. 1995) as well as crossed spinal connections (Koceja and Kamen 1992; Mezzarane and Kohn 2002). For instance, recurrent inhibition of Ia inhibitory interneurons would periodically release antagonistic motoneurons from their tonic inhibitory activity, a phenomenon known as recurrent facilitation (Hultborn et al. 1971b; Pierrot-Deseilligny and Burke 2012). Independently, the preferential generation of synchronous reflex modulation in pairs of muscle groups may be explained by the same initial conditions across muscles with the first stimulus pulse leading to large, unconditioned PRM reflexes.
Involvement of spinal oscillating networks

An alternative mechanism underlying the periodic reflex modulation could be the activation of a spinal network with inherent oscillating capability that cyclically modified the excitability of the central components of the PRM-reflex arcs. One network capable of producing rhythmic motor output in response to tonic SCS is the spinal locomotor rhythm and pattern generator (Danner et al. 2015). Yet, the higher stimulation frequencies of 22–60 Hz required for its activation as well as the lower cycle frequency of the produced rhythmic outputs of 0.27–1.84 Hz (Dimitrijevic et al. 1998; Minassian et al. 2004; Danner et al. 2015) render its involvement in the patterns described here highly unlikely.

Another spinal oscillating system that may underlie the periodic reflex modulation is indicated by the occurrence of the larger-amplitude PRM reflexes within these patterns at 5.5–8 Hz that are the usual rates of sustained clonus (Dimitrijevic et al. 1980). One hypothesis for the generation of clonus is that peripheral events such as stretch interact with a spinal generator that produces alternating excitatory-refractory cycles which sustain and regulate the repetitive clonic bursts (Dimitrijevic et al. 1980; Beres-Jones et al. 2003). Dimitrijevic and colleagues (1980) also found that Achilles tendon reflexes evoked in clonic muscles attained large EMG amplitudes when tapping at rates similar to that of clonus. When tapping rates were further increased, reflex attenuation occurred, affecting each response, or – with tapping rates close to twice the rate of clonus – a large response to alternate taps was evoked. The periodic reflex modulation described here is reminiscent of the latter finding. Hence, the SCS inputs may have initiated and entrained centrally organized cyclical changes of excitability similar to those underlying clonus, which in turn affected the excitability cycles of the PRM reflexes.

Role of stimulation frequency in central and muscular effects
Clinical applications of epidural SCS in individuals with severe SCI are motivated by the finding that sustained stimulation at 5–15 Hz can generate bilateral extension (Jilge et al. 2004), sufficient to induce standing (Harkema et al. 2011), while 22–60 Hz can produce alternating flexion-extension movements (Dimitrijevic et al. 1998). It was suggested that the different frequencies recruited different elements of spinal pattern generating networks (Jilge et al. 2004; Minassian et al. 2007a; Danner et al. 2015). Since the SCS-induced motor outputs are composed of series of stimulus-triggered PRM reflexes (Jilge et al. 2004; Minassian et al. 2004), the stimulation frequency entrains the motoneuron-firing rate and thus critically influences the muscle force produced. This relationship between the stimulation frequency and muscle force was also used in a recent study to achieve control of hindlimb kinematics during SCS-induced locomotion in paralyzed rats (Wenger et al. 2014). The phenomenon described in the present manuscript shows that, under specific conditions, the effective output frequencies (given by the larger amplitude responses within the periodic modulation) may be lower than the stimulation frequency applied. For inducing standing, increasing the stimulation frequency slightly above the ones optimal to centrally organize an extension function may be required to avoid PRM-reflex alternation (cf. Fig. 2A) and to produce adequate levels of extensor-muscle force.

Conclusion

The repetitive elicitation of monosynaptic spinal reflexes at distinct rates led to a simple form of oscillating motor output with unanticipated prevalence in individuals after chronic loss of supraspinal input. Apparently, the bilateral, multi-segmental activation as provided by SCS was essential to reveal this type of input-processing and output-generation by the human lumbosacral spinal circuitry. The recent renaissance of epidural lumbar SCS in SCI people (Harkema et al. 2011; Angeli et al. 2014; Danner et al. 2015) as well as novel neurophysiological studies employing transcutaneous SCS (Dy et al. 2010; Hofstoetter et al. 2015)
2013, 2014; Knikou 2014; Roy et al. 2014) may provide means to further advance the current understanding of human spinal sensorimotor integration.

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Disclosures

No conflicts of interest, financial or otherwise, are declared by the authors.

References


**Figure 1.** Series of PRM reflexes without and with periodic amplitude modulation elicited by epidural stimulation of the lumbosacral spinal cord. A(i) Upper trace shows responses of hamstrings (Ham) to 2-Hz stimulation with unmodulated amplitudes, lower trace illustrates periodic reflex modulation elicited at 22 Hz in the same muscle group (note different time scaling). (ii) Same PRM reflexes as shown in (i) but presented stimulus-triggered in enlarged time scale. Inserted values for n are numbers of superimposed responses. B(i) Examples illustrating the characteristic short-build up phase of periodic modulation patterns, here elicited at 16 Hz, established either with the first pair of stimuli applied (upper trace) or after a few stimulus pulses (lower trace). (ii) Peak-to-peak amplitudes (black circles) of the consecutively elicited PRM reflexes shown in (i). Arrows mark times of stimulus application. Data derived from subject 2, epidural electrode combination 0–3+, 6 V in A(i), both traces, and B(i), lower trace; and 0+3–, 3V in B(i), upper trace.

**Figure 2.** Relationship between the prevalence of periodic reflex modulation and stimulation parameters. Number (#) of recording sections (each with constant stimulation parameters) with periodic reflex modulation of quadriceps (Q), hamstrings (Ham), tibialis anterior (TA), and triceps surae (TS) relative to the total number of available recording sections across subjects at a given stimulation A frequency, B intensity (in multiples of the respective reflex thresholds, thr.), and C site.

**Figure 3.** Prevalence and consistency of periodic reflex modulation per muscle group. A Number (#) of recording sections with segments featuring periodic reflex modulation (grey areas within bars) in relation to total number nt total of recording sections (each with a given set of stimulation parameters) from quadriceps (Q), hamstrings (Ham), tibialis anterior (TA), and triceps surae (TS) across subjects. B Percentage of recording sections with periodic reflex modulation per muscle with a consistency below 50% (grey, hatched area) or above 50%
(grey area). The percentage of modulation patterns occurring with a consistency of 100% (i.e., throughout a recording section) is marked by dashed rectangles within bars.

**Figure 4.** Stimulation-frequency dependence of periodic reflex modulation. A Average consistency of periodic reflex modulation (describing its prevalence per recording section with unchanged stimulation parameters) within each muscle and B attenuation of the smaller relative to the larger responses as a function of the stimulation frequency; error bars are standard errors.

**Figure 5.** Coordination of periodic reflex modulation in pairs of related muscles. A Relationship of motor outputs in antagonistic muscle groups with (i) periodic modulation occurring in one of the antagonists only; (ii) reciprocal modulation; and (iii) synchronous modulation. B Periodic modulation patterns occurring in the same muscle group bilaterally with (i) reciprocal, and (ii) synchronous relationship. Recordings from unilateral quadriceps (Q) and hamstrings (Ham) as well as right (R) and left (L) tibialis anterior (TA) and triceps surae (TS), subject 2. Stimulation parameters: A, (i) 0+2–, 11 Hz, 5 V; (ii) 0–3+, 16 Hz, 5 V; and (iii) 0+2–, 16 Hz, 4 V; B, (i) 0–3+, 11 Hz, 7 V; and (ii) 0–3+, 16 Hz, 7 V.

**Figure 6.** Consistency and attenuation of periodic reflex-modulation patterns. Patterns found in one (white bars) or both (black bars) muscle groups of a pair, A, with antagonistic relation, and B, bilaterally. The consistency of a pattern, measured within each muscle, is significantly larger and the attenuation of the smaller responses relative to the larger ones significantly stronger when the periodic reflex modulation is concomitantly occurring in the related muscle (bars are mean values across all available data sets, error bars are standard errors; all P<.00001).

**Figure 7.** The two most frequently detected output relationships of periodic reflex modulation across muscles of the same side. Data derived from A, subject 2, 0–3+, 16 Hz, 6 V and B, subject 5, c+1–, 16 Hz, 6 V. Arrows mark times of stimulus application.
Figure 8. Characteristic EMG waveforms and onset latencies of PRM reflexes. A Responses to 2-Hz stimulation. B Responses from patterns of periodic amplitude modulation. Arrows indicate times of stimulus delivery, EMG waveforms are exemplary and were characteristic across subjects, triangles represent onset latencies averaged over all available data sets and subjects, horizontal lines show standard errors. Two characteristic waveforms were found in Ham and TA. Data derived from A, Q, subject 4, 0+3–, 8V; Ham, subject 8, 0+3–, 4V (solid line), and subject 2, 0+3–, 5V (dashed line); TA, subject 8, 2+3–, 5V (solid line), and subject 2, 0+2–, 6V (dashed line); and TS, subject 4, 1+3–, 9V; and B, Q, subject 4, 0+3–, 11Hz, 5V; Ham, subject 2, 0+3–, 16Hz, 4V (solid line), and subject 8, 1+3–, 16Hz, 4V (dashed line); TA, subject 2, 0+2–, 11Hz, 5V (solid line), and subject 8, c+3–, 11Hz, 4V (dashed line); and TS, subject 2, 0+2–, 11Hz, 5V. Waveforms are presented amplitude-matched.
Table 1. Subject demographic data.

<table>
<thead>
<tr>
<th>Subj.</th>
<th>Sex</th>
<th>Age (y)</th>
<th>AIS</th>
<th>Neurol. injury level</th>
<th>Years post-injury</th>
</tr>
</thead>
<tbody>
<tr>
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<td>M</td>
<td>29</td>
<td>B</td>
<td>C6</td>
<td>3</td>
</tr>
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<td>M</td>
<td>33</td>
<td>A</td>
<td>T5</td>
<td>13</td>
</tr>
</tbody>
</table>

AIS, American Spinal Injury Association Impairment Scale
Table 2. Relationship of periodic reflex modulation in pairs of antagonistic muscle groups (top) and in a given muscle group bilaterally (bottom).

<table>
<thead>
<tr>
<th>Muscles</th>
<th>( n_{\text{mod_total}} )</th>
<th>( n_{\text{single}} )</th>
<th>( n_{\text{reciprocal}} )</th>
<th>( n_{\text{synchron.}} )</th>
<th>( n_{\text{fluctuating}} )</th>
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</thead>
<tbody>
<tr>
<td>Q</td>
<td>136</td>
<td>36</td>
<td>52</td>
<td>45</td>
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</tr>
<tr>
<td>Ham</td>
<td>154</td>
<td>54</td>
<td>52</td>
<td>45</td>
<td>3</td>
</tr>
<tr>
<td>TA</td>
<td>61</td>
<td>17</td>
<td>2</td>
<td>41</td>
<td>1</td>
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<tr>
<td>TS</td>
<td>80</td>
<td>36</td>
<td>2</td>
<td>41</td>
<td>1</td>
</tr>
</tbody>
</table>

- \( n_{\text{mod\_total}} \), total number of recording sections containing segments with periodic reflex modulation per muscle group; 
- \( n_{\text{single}} \), periodic alternation in one of the antagonistic muscles only; 
- \( n_{\text{reciprocal}} \), reciprocal modulation, and 
- \( n_{\text{synchron.}} \), synchronous modulation in pairs of antagonistic muscle groups; 
- \( n_{\text{fluctuating}} \), fluctuation between reciprocal and synchronous modulation.

<table>
<thead>
<tr>
<th>Muscles</th>
<th>( n_{\text{bilateral}} )</th>
<th>( n_{\text{reciprocal}} )</th>
<th>( n_{\text{synchron.}} )</th>
<th>( n_{\text{fluctuating}} )</th>
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</thead>
<tbody>
<tr>
<td>Q bilateral</td>
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<td>36</td>
<td>3</td>
</tr>
<tr>
<td>Ham bilateral</td>
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<td>84</td>
<td>6</td>
</tr>
<tr>
<td>TA bilateral</td>
<td>36</td>
<td>8</td>
<td>24</td>
<td>4</td>
</tr>
<tr>
<td>TS bilateral</td>
<td>46</td>
<td>6</td>
<td>36</td>
<td>4</td>
</tr>
</tbody>
</table>

- \( n_{\text{bilateral}} \), number of recording sections with periodic modulation patterns occurring bilaterally in a given muscle group, of which \( n_{\text{reciprocal}} \) are modulating reciprocally and \( n_{\text{synchron.}} \) synchronously; 
- \( n_{\text{fluctuating}} \), fluctuation between reciprocal and synchronous modulation.

Quadriceps (Q), hamstrings (Ham), tibialis anterior (TA), and triceps surae (TS)
Table 3. Observed patterns of periodic reflex modulation across subjects.

<table>
<thead>
<tr>
<th>Subj.</th>
<th>Modulation in pairs of antagonists</th>
<th>Modulation occurring in a muscle bilaterally</th>
<th>Modulation in all 4 unilateral muscles</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>x</td>
<td>x</td>
<td>x</td>
</tr>
<tr>
<td>2</td>
<td>x</td>
<td>x</td>
<td>x</td>
</tr>
<tr>
<td>3</td>
<td>x</td>
<td></td>
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<tr>
<td>4</td>
<td>x</td>
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<td>5</td>
<td>x</td>
<td>x</td>
<td>x</td>
</tr>
<tr>
<td>6</td>
<td>x</td>
<td>x</td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>x</td>
<td>x</td>
<td>x</td>
</tr>
<tr>
<td>8</td>
<td>x</td>
<td>x</td>
<td>x</td>
</tr>
</tbody>
</table>

Periodic reflex modulation in one of the antagonistic muscle groups only (Single); periodic reflex modulation occurring concomitantly in pairs of related muscle groups with reciprocal (Recip.) or synchronous (Synch.) relationship; fluctuation (Fluct.) between reciprocal and synchronous modulation; x = pattern present.
A

(i) Ham, 2 Hz

(ii) Ham, 2 Hz

B

(i) Ham, 16 Hz - pattern established with first pair of stimuli

(ii) Ham, 16 Hz - pattern established after short build-up phase

Ham, 16 Hz - pattern established with first pair of stimuli

Ham, 16 Hz - pattern established after short build-up phase

1.5
1.0
0.5
0
peak-to-peak amplitude [mV]
response number

1 5 10 15 20

1 5 10 15 20

peak-to-peak amplitude [mV]
response number

1 5 10 15 20

1 5 10 15 20
A

B

n_{total} total # recording sections available

# rec. sections with patterned segments

Q Ham TA TS

n_{total} 530
n_{total} 491
n_{total} 381
n_{total} 335

# rec. sections available

# rec. sections with patterned segments

consistency ≥ 50%
consistency < 50%
consistency = 100%

Q Ham TA TS
Modulation in

- □ one muscle group only
- ▪ pairs of related muscle groups