Functional segregation of voltage-activated calcium channels in motoneurons of the dorsal motor nucleus of the vagus

Abbreviated title: Calcium dynamics in vagal motoneurons

Garry Cooper1*, Efrat Lasser-Katz2*, Alon Simchovitz3, Ronit Sharon4, Hermona Soreq3, D. James Surmeier1 and Joshua A. Goldberg2

1Department of Physiology, Feinberg School of Medicine, Northwestern University, Chicago, IL, 60611
2Department of Medical Neurobiology, Institute of Medical Research Israel–Canada, Faculty of Medicine, The Hebrew University of Jerusalem, Jerusalem, 9112102, Israel
3The Edmond and Lily Safra Center for Brain Sciences & The Life Science Institute, The Hebrew University of Jerusalem, Jerusalem, 91404, Israel
4Department of Biochemistry and Molecular Biology, Institute of Medical Research Israel–Canada, Faculty of Medicine, The Hebrew University of Jerusalem, Jerusalem, 9112102, Israel

* These authors contributed equally to this study.

Author contribution: G.C. and E. L.-K. conducted experiments; A.S. conducted the qPCR analysis; R.S. and H.S. oversaw parts of the study; D.J.S. designed experiments and oversaw parts of the study; J.A.G. designed and conducted experiments, analyzed the results, designed and conducted the numerical modeling, wrote the manuscript and prepared the figures. All authors read and approved the final version of the manuscript.

Correspondence to:
Joshua A. Goldberg, PhD
Department of Medical Neurobiology
Institute of Medical Research Israel-Canada
The Faculty of Medicine
The Hebrew University of Jerusalem
POB 12272
9112102 Jerusalem, Israel
Tel: +972-2-675-7385
fax: +972-2-643-9736
E-mail: joshg@ekmd.huji.ac.il
Abstract
Calcium influx elevates mitochondrial oxidant stress (mOS) in dorsal motor nucleus of the vagus (DMV) neurons that are prone to Lewy body pathologies in presymptomatic Parkinson’s disease (PD) patients. In experimental PD models, treatment with isradipine – the dihydropyridine with the highest affinity to Cav1.3 channels – prevents subthreshold calcium influx via Cav1.3 channels into midbrain dopamine neurons and protects them from mOS. In DMV neurons, isradipine is also effective at reducing mOS despite overwhelming evidence that subthreshold calcium influx is negligible in comparison to spike-triggered influx. To solve this conundrum we combined slice electrophysiology, two-photon laser scanning microscopy, mRNA profiling and computational modeling. We find that the unusually depolarized sub-threshold voltage trajectory of DMV neurons is positioned between the relatively hyperpolarized activation curve of Cav1.3 channels and that of other high-voltage activated (HVA) calcium channels, thus creating a functional segregation between Cav1.3 and HVA calcium channels. The HVA channels flux the bulk of calcium during spikes, but can only influence pacemaking through their coupling to calcium-activated potassium currents. In contrast, CaV1.3 currents, which we show to be more than an order-of-magnitude smaller than the HVA calcium currents, are able to introduce sufficient inward current to speed up firing. However, Kv4 channels that are constitutively open in the subthreshold range guarantee slow pacemaking, despite the depolarizing action of Cav1.3 and other pacemaking currents. We propose that the efficacy of isradipine in preventing mOS in DMV neurons arises from its mixed effect on Cav1.3 channels and on HVA Cav1.2 channels.

Keywords: calcium dynamics; vagal motoneurons; modeling; Traub model; Cav1.2; Cav1.3; Cav2; Kv4; NALCN; window current; Parkinson’s disease; Lewy body neurite pathology; two photon laser scanning microscopy; calcium imaging; isradipine; lingering; Hodgkin class I excitability; mouse; mitochondrial oxidative stress
Introduction

The cholinergic motoneurons of the dorsal motor nucleus of the vagus (DMV) are reported to be among the first neurons to exhibit Lewy body and Lewy neurite pathology in Parkinson’s disease (PD) (Lewy 1912), before its appearance in dopamine (DA) neurons of the substantia nigra pars compacta (SNc) (Braak et al. 2004). The functional decline of DMV neurons may underlie PD dysautonomia (Miller et al. 2009), and the loss of DMV neurons correlates with the severity of PD (Gai et al. 1992). We have shown that DMV neurons in brain slices taken from the DJ-1 mouse model of PD exhibit higher basal mitochondrial oxidant stress (mOS) than DMV neurons from littermate wild-type mice. Pre-incubation of these brain slices in isradipine, an antagonist of plasma membrane Cav1 (L-type) calcium (Ca\(^{2+}\)) channels, reduced the basal mOS in the DMV neurons of the DJ-1 mice. Isradipine also lowered the cytosolic Ca\(^{2+}\) concentration in DMV neurons (Goldberg et al. 2012). The drop in cytosolic Ca\(^{2+}\) levels, which presumably leads to a drop in mitochondrial Ca\(^{2+}\) levels and a slowing of respiration, reduces basal mOS (Goldberg et al. 2012).

Isradipine was chosen for that study because of its high affinity to Cav1.3 Ca\(^{2+}\) channels (Sinnegger-Brauns et al. 2009) that are activated in the subthreshold range (Lipscombe et al. 2004). This choice was motivated by previous studies in SNc DA neurons that showed that Ca\(^{2+}\) flux through Cav1.3 Ca\(^{2+}\) channels, during subthreshold dendritic Ca\(^{2+}\) oscillations (Guzman et al. 2009; Guzman et al. 2010; Puopolo et al. 2007), was a determinant of basal cytosolic Ca\(^{2+}\) levels and basal mOS, both of which were reduced by isradipine.

However, three findings seem to marginalize the contribution of subthreshold Ca\(^{2+}\) influx to basal cytosolic Ca\(^{2+}\) levels in DMV neurons. First, imaging of dendritic Ca\(^{2+}\) concentrations in DMV neurons during autonomous pacemaking failed to reveal substantial Ca\(^{2+}\) influx between spikes. Second, once DMV neurons were silenced by hyperpolarization, further hyperpolarization did not further reduce their cytosolic Ca\(^{2+}\) concentration. Third, measurement of subthreshold Ca\(^{2+}\) currents in DMV neurons demonstrated that they were quite small, in comparison, for example, to leak and voltage-activated sodium (Na\(^{+}\))
currents. These findings suggest a more prominent role for the high-voltage
activated (HVA) Cav1.2 Ca\(^{2+}\) channels, to which isradipine has an equal affinity
(Sinnegger-Brauns et al. 2009), in determining Ca\(^{2+}\) levels and basal mOS in DMV
neurons.

In the present study we set out to clarify the specific physiological role of
Cav1.2 and Cav1.3 channels in Ca\(^{2+}\) dynamics – as well as the autonomous firing
patterns – observed in DMV neurons (Goldberg et al. 2012). Using a combination
of (a) slice electrophysiology; (b) two-photon laser scanning microscopy (2PLSM)
Ca\(^{2+}\) imaging; (c) mRNA profiling; and (d) numerical modeling, we found that the
depolarized autonomous voltage oscillations, due to NALCN, Nav1 Na\(^{+}\) and Kv4
potassium (K\(^{+}\)) channel currents create a functional segregation between Cav1.3
channels and Cav1.2 (as well as other HVA, such as Cav2) Ca\(^{2+}\) channels in DMV
neurons. A very small Cav1.3 current suffices to speed up the firing rate, without
significantly contributing to the basal Ca\(^{2+}\) concentration, which is attributable
almost entirely to spike triggered Ca\(^{2+}\) influx via HVA Ca\(^{2+}\) channels. This latter
influx activates Ca\(^{2+}\) activated K\(^{+}\) currents that underlie afterhyperpolarizations
(AHPs) (Sah and McLachlan 1992) that slow the pacemaking (Goldberg et al.
2012). We propose that equal affinity of isradipine to Cav1.2 and Cav1.3 channels
reduces spike-triggered Ca\(^{2+}\) influx, and prevents Cav1.3 channels from driving
pacemaking, thereby explaining the drug’s efficacy in protecting DMV neurons
from mOS.

Materials and Methods

2PLSM imaging: The 2PLSM system was described previously (Guzman et al.
2010). Briefly, the two-photon excitation source was a Chameleon Ultra 2 tunable
laser system (680–1,080 nm) (Coherent Laser Group, Santa Clara, Ca). Optical
signals were acquired using an 820-nm excitation beam to excite Alexa and Fluo-4
dyes simultaneously. Laser power attenuation was achieved with two Pockels
cells electro-optic modulators (model 350-80, Conoptics, Danbury, CT). The
fluorescence emission was collected with non-descanned photomultiplier tubes
(PMTs) (Prairie Technologies, Madison, WI). A Dodt contrast detector system was
used to provide a bright-field transmission image in registration with the fluorescent images.

**Slice Preparation:** Experimental procedures adhered to the Northwestern University and the Hebrew University Animal Care and Use Committees. Three to 8 week old C57BL/6 mice of both sexes were deeply anesthetized with ketamine–xylazine and perfused transcardially with ice-cold modified artificial cerebrospinal fluid (ACSF), bubbled with 95% O2-5% CO2, and containing (in mM): 2.5 KCl, 26 NaHCO3, 1.25 Na2HPO4, 0.5 CaCl2, 10 MgSO4, 0.4 ascorbic acid, 10 glucose, and 210 sucrose. The cerebellum, pons and medulla were rapidly removed, blocked in the coronal plane, and sectioned at a thickness of 240 µm in ice-cold modified ACSF. Slices were then submerged in ACSF, bubbled with 95% O2-5% CO2, and containing (in mM): 2.5 KCl, 126 NaCl, 26 NaHCO3, 1.25 Na2HPO4, 2 CaCl2, 2 MgSO4, and 10 glucose, and stored at room temperature for at least one hour prior to recording and/or imaging.

**Slice visualization and electrophysiology:** The slices were transferred to the recording chamber mounted on an Olympus BX51 upright, fixed-stage microscope and perfused with oxygenated ACSF at 32°C. The only exception was that voltage sensitive Na+ currents were recorded in a HEPES-based solution that contained (in mM): 2.5 KCl, 137 NaCl, 1.8 CoCl2, 1 MgCl2, 5.4 tetraethylammonium (TEA)-Cl, 10 4-aminopyridine, 10 glucose, 10 HEPES (pH=7.4 with HCl). A 60X, 0.9 NA water-immersion objective was used to examine the slice using standard infrared differential interference contrast video microscopy. Patch pipette resistance was typically 3-4.5 MΩ. For whole-cell current clamp recordings and for voltage-clamp measurements of K+ conductances the pipette contained (in mM): 135.5 K-MeSO4, 5 KCl, 2.5 NaCl, 5 Na-phosphocreatine, 10 HEPES, 0.2 EGTA, 0.21 Na2GTP, and 2 Mg1.5ATP (pH=7.3 with KOH, 280-290 mOsm/kg). For Ca2+ imaging experiments in conjunction with current clamp recordings the pipette contained (in mM): 135 K-MeSO4, 5 KCl, 5 Na-phosphocreatine, 5 Tris-phosphocreatine, 10 HEPES, 0.1 Fluo-4, 0.05 Alexa Fluor 568, 0.21 Na2GTP, and 2 Mg1.5ATP (pH=7.3 with KOH, 280-290 mOsm/Kg). For voltage clamp Ca2+ imaging the pipette contained (in mM): 120 Cs-
Cooper et al. 2015

Me-\(\text{SO}_3\), 15 CsCl, 8 NaCl, 10 TEA, 5 QX-314, 10 HEPES, 0.3 Na\(_2\)GTP, and 3 Mg\(_{1.5}\)ATP, 0.1 Fluo-4, 0.05 Alexa Fluor 568 (pH=7.3 with CsOH, 280-290 mOsm/kg).

Electrophysiological recordings were obtained with a Multiclamp 700B amplifier (Molecular Devices, Sunnyvale, CA). Junction potential, which was 7-8 mV, was not corrected. Signals were digitized at 20–100 kHz and logged onto a personal computer with the Clampex 9.2 software (Molecular Devices) or, in the imaging experiments, using the custom-written shareware package WinFluor (John Dempster, Strathclyde University, Glasgow, Scotland, UK), which automates and synchronizes the 2PLSM imaging and electrophysiological protocols. The voltage ramps had a slope of 25 mV/s.

**Drugs and reagents:** For recording of autonomous discharge the following cocktail of synaptic receptor blockers was used that included (in µM): 50 D-APV to block NMDA receptors, 5 NBQX to block AMPA receptors, 10 SR 95531 (gabazine) to block GABA\(_A\) receptors, 1 CGP 55845 to block GABA\(_B\) receptors, 10 mecamylamine to block nicotinic receptors, 10 atropine to block muscarinic receptors, and – in some of the experiments – 1 ketanserin to block 5-HT\(_2A\) receptors. Voltage sensitive Na\(^+\) channels were blocked with 1 µM tetrodotoxin (TTX). Cav1 channels were antagonized with 5 µM of either nifedipine or isradipine, or 200 nM calciseptine. The acute effects of solution exchanges or drug applications were measured at least 5 minutes after wash on, except for dihydropyridine where 10 minutes were given. The only exception was the application of phrixotoxin-2, a selective Kv4 (A-type) channel blocker. In these experiments we pipetted 20 µl of a 100 µM stock solution into the recording chamber whose volume was approximately 1-2 ml, thereby reaching a nominal concentration of 1-2 µM phrixotoxin-2 in the bath until the solution was replaced (Subramaniam et al. 2014) Fluo-4 pentapotassium and Alexafluor 568 hydrazide Na\(^+\) salts were obtained from Invitrogen (Carlsbad, CA). TTX, phrixotoxin-2 and calciseptine were obtained from Alomone Labs (Jerusalem, Israel). The rest of the drugs and reagents were obtained from Tocris (Ellisville, MO) or Sigma (St. Louis, MO).
RNA extraction from mouse brain slices: The DMV was dissected out from brain slices taken from 5 C57BL/6 mice. RNA was extracted from brain slices using the QIAGEN (Venlo, Netherlands) RNAeasy kit, which ensures full representation of all RNA length groups. Briefly, brain slices were homogenized with 500 µl QIAzol lysis buffer, lysed for 5 minutes, mixed with 100 µl chloroform to allow full neutralization, suspended for 3 minutes and centrifuged for 15 minutes (at 12,000x g and 4ºC). The aqueous phase was mixed with 1.5 volume of ethanol, loaded on RNA-binding spin column and centrifuged for 30 seconds in 8400xg. The column was washed once by centrifugation (30 seconds at 8400x g), and discarding the flow-through with 700 µl RWT buffer (85% ethanol); and twice with 500 µl RPE buffer (QIAGEN, 70%), for the removal of DNA and protein remnants, respectively. Finally, spin column was centrifuged for 1 minute in 21,067xg to further dry the column. 35 µl of nuclease-free water were used to elute the RNA, which was immediately put on ice to prevent degradation. RNA concentration was determined by Nanodrop-2000 (Thermo-Scientific, Waltham, MA), and its integrity was assessed via 1% agarose gel and identification of two distinct rRNA bands (28S and 18S).

cDNA synthesis from mouse brain slices: cDNA preparation was performed using the Quanta qScript cDNA Synthesis Kit (Quanta biosciences Inc., Gaithersburg, MD). 130 ng of RNA were diluted in 15 µl of nuclease-free water, and mixed with 4 µl 5x Reaction Buffer and 1µl of Reverse Transcriptase (Except for the no-RT controls, where reverse transcriptase was not added), for a final 20 µl reaction volume. Mixture was put in a 200 mL PCR tube, and placed in a MJ Research PTC 200 Thermal Cycler (GMI Inc., Ramsey, MN) programmed for 5 minutes in 22ºc, 30 minutes in 42ºc and 5 minutes in 85ºc. cDNA was then diluted 1:10, by adding 180 mL DDW.

qRT-PCR - Quantitative real-time PCR and primers: For each reaction, iTaq Universal SYBR green Supermix 2x (7.5 µL, Biorad Inc., Hercules, CA) was used for both the target and reference genes employed for normalization together with 0.75 µl of left and right primers (10 mM of each), and 6 µl of cDNA on a Biorad CFX96 Touch Real-Time PCR Detection System. Amplification was performed in
triplicates for each primer pair and tested samples, for 95°C (3 minutes), 95°C (15 seconds) and 40 repeats of 60°C for 30 seconds, followed by increasing the temperature from 67.0 to 94.6°C in 0.3°C increments every 5 seconds to create a melting curve. The data was obtained using Bio-Rad CFX Manager 3.0 software. Also, for each primer pair a dilution curve was created in order to calculate primer efficiency that was then used to re-calculate the expression results. Fixed values were normalized to either 18S as a housekeeping gene or the neuronal-specific TUBB3 as a neuronal marker. The primers (Sigma) used to identify all known transcriptional variants of the Cav channels and reference genes were as follows:

<table>
<thead>
<tr>
<th>Gene Name</th>
<th>Primer Direction</th>
<th>Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cav1.2</td>
<td>Forward</td>
<td>CGAAGGTACATCCCCAAGAA</td>
</tr>
<tr>
<td></td>
<td>Backward</td>
<td>CGATTITGAAGAGGCAGCTC</td>
</tr>
<tr>
<td>Cav1.3</td>
<td>Forward</td>
<td>ATCCCTACACCGAAGCTCCT</td>
</tr>
<tr>
<td></td>
<td>Backward</td>
<td>TCGGCAATCTCATGTTTTGT</td>
</tr>
<tr>
<td>Cav2.1</td>
<td>Forward</td>
<td>GACAGGGAGCGAGCACAC</td>
</tr>
<tr>
<td></td>
<td>Backward</td>
<td>GGTGGCCAGCTTGTATTTCT</td>
</tr>
<tr>
<td>Cav2.2</td>
<td>Forward</td>
<td>GGTGGAAGGGGATAAGGAAA</td>
</tr>
<tr>
<td></td>
<td>Backward</td>
<td>GGTGTCTGCATCCTCTGGT</td>
</tr>
<tr>
<td>Cav2.3</td>
<td>Forward</td>
<td>ACACCTTCCCTGCAGCTATC</td>
</tr>
<tr>
<td></td>
<td>Backward</td>
<td>GCGAATGCCATTGTACATCA</td>
</tr>
<tr>
<td>18S</td>
<td>Forward</td>
<td>ACAACAAGCTGCGTGAGGAC</td>
</tr>
<tr>
<td></td>
<td>Backward</td>
<td>CAAAGGCCAGAGACTCATT</td>
</tr>
<tr>
<td>TUBB3</td>
<td>Forward</td>
<td>CGCCTTTGGACACCATATTCA</td>
</tr>
<tr>
<td></td>
<td>Backward</td>
<td>TGCAGGCAGTCACAATTCT</td>
</tr>
</tbody>
</table>

Data and Statistical Analysis: Electrophysiological data were analyzed and curve fitting was done using custom-made code on custom-made (Winfluor, Dr. John Dempster, University of Strathclyde; NUPver, Nicholas Schwarz, Northwestern University; Oscilloscope, Dr. Charles Wilson, University of Texas at San Antonio) and commercial (Matlab, The Mathworks, Natick, MA) software. The two-tailed Wilcoxon signed-rank test (SRT) was used to test for changes of medians in matched-paired comparisons. The non-parametric Kruskal–Wallis one-way
analysis of variance (ANOVA) test was used to compare the relative expression of Cav channel mRNA. The null hypothesis of equal medians was rejected if the P value was below 0.05.

**Computational modeling:** To study the Ca\(^{2+}\) dynamics exhibited by the DMV cells we converted the Traub model – a single compartment model of a non-autonomously spiking neuron, exactly as it appears in Ermentrout (1998) – into an autonomous pacemaker by replacing the externally applied current term with two voltage activated Ca\(^{2+}\) currents and by shifting all voltages by +20 mV, to replicate the very depolarized potentials of the DMV pacemaker that result from the voltage-insensitive NALCN cation current that that they express (Goldberg et al. 2012). The two Ca\(^{2+}\) conductances differed only in their half activation voltages and in their maximal conductance: \(I_L\) with a half activation of \(V_{1/2} = -40\) mV, and maximal conductance of \(g_L\) representing the Cav1.3 current; and \(I_N\) with a half activation of \(V_{1/2} = -20\) mV and maximal conductance of \(g_N\) representing the HVA currents such as the Cav1.2 and Cav2 currents. The units of the maximal conductance are mS/cm\(^2\).

The Ca\(^{2+}\) currents were given by

\[
I_X = g_X(V - E_{Ca}) / \left[1 + \exp \left(-\frac{(V - V_{1/2}^X)}{V_S^X}\right)\right]^3 \quad \text{for } X = L, N
\]

where \(E_{Ca} = 120\) mV and \(V_{1/2}^X = 8\) mV for both \(X = L, N\) (Goldberg et al. 2009). The equation for the concentration of free Ca\(^{2+}\) in the cell is given by

\[
\frac{d}{dt}[Ca] = -\frac{Ca}{\tau_{Ca}} - r \left(\frac{I_L + I_N}{zF}\right)
\]

where \([Ca]\) are in units of nM, \(r=600\) cm\(^{-1}\) and \(\tau_{Ca} = 50\) ms. The units of the Faraday constant are kC/mole and \(z=2\) for Ca\(^{2+}\) ions.

**Results**

**Blockade of Cav1 channels reduces Ca\(^{2+}\) influx per spike and speeds up pacemaking**

Motoneurons of the DMV are autonomous pacemakers that discharge at a median rate of approximately 1 spike/s in the presence of synaptic blockers. The basal cytosolic Ca\(^{2+}\) concentration in DMV neurons is generated almost exclusively
by Ca\textsuperscript{2+} influx associated with this spiking (Goldberg et al. 2012). Because Cav1 antagonists reduce this basal concentration (Goldberg et al. 2012), these drugs should also reduce the amount of Ca\textsuperscript{2+} influx per individual spike. To test this we conducted 2PLSM imaging of somatic Ca\textsuperscript{2+} in DMV neurons while injecting various holding currents to either hyperpolarize or depolarize the cells (Fig. 1A). As described previously (Goldberg et al. 2012), this generated a linear dependence of ∆F/F\textsubscript{0} on firing rate. Application of calciseptine, a Cav1-selective neurotoxin, significantly reduced the slope indicating that Cav1 channels contribute to the amount of Ca\textsuperscript{2+} influx that occurs with each spike. Previous studies have shown that Ca\textsuperscript{2+} influx via Cav1.2 channels generates the slow AHP (sAHP) in DMV neurons (Goldberg et al. 2012; Sah and McLachlan 1992). We have previously shown that Ca\textsuperscript{2+} influx through Cav2.2 channels (another HVA Ca\textsuperscript{2+} channel) activates the small-conductance Ca\textsuperscript{2+}-activated K\textsuperscript{+} (SK) current that underlies medium AHP (mAHP) that follows each individual spike and slows DMV pacemaking significantly (Goldberg et al. 2012). In contrast, in several neuronal types the sAHP usually affects mostly evoked discharge rates (Gustafsson and Wigstrom 1983; Hotson and Prince 1980; Schwindt et al. 1988). Nevertheless, we tested whether antagonizing Cav1 channels affected peacemaking. Application of 5 µM nifedipine modestly raised the firing rate of all 7 DMV neurons tested (median firing rate increase from 1.05 spikes/s to 1.28 spikes/s, P < 0.05, SRT, Fig. 2A). Therefore, Cav1 Ca\textsuperscript{2+} channels contribute to spike-triggered Ca\textsuperscript{2+} influx, and may contribute to the slowing of pacemaking in DMV neurons by activating AHPs.

**Acute isradipine application slows pacemaking**

In contrast to the robust suprathreshold Ca\textsuperscript{2+} influx through HVA Ca\textsuperscript{2+} channels, the subthreshold Ca\textsuperscript{2+} influx in between spikes (that must flow through Cav1.3 channels because they are the only non-inactivating low-voltage activated Ca\textsuperscript{2+} channels) is small or nonexistent (Goldberg et al. 2012). If Cav1.3 channels do not flux much Ca\textsuperscript{2+} what is their physiological role? To answer this question we tested the effect of isradipine, which is the dihydropyridine with the highest affinity to Cav1.3 channels, on autonomous firing in DMV neurons recorded in the loose patch configuration. We have previously shown that preincubation of DMV
slices with 200 nM isradipine does no affect basal firing rates (Goldberg et al. 2012). However, it is possible that application of a higher concentration would lead to a different effect by antagonizing Cav1.3 channels acutely. Indeed, acute application of 5 µM isradipine reduced the median firing rate from 0.99 spikes/s to 0.23 spikes/s (P < 0.01, SRT, n=9). Furthermore, in all 6 cells in which we attempted to wash off the effect of isradipine the firing rate recovered from a median of 0.22 spikes/s to 1.08 spikes/s (P < 0.05, SRT) (Fig. 2B). The caveat in these experiments is that voltage dependent persistent sodium currents are necessary and sufficient to drive pacemaking of DMV motoneurons (Goldberg et al. 2012), and there is a possibility that 5 µM isradipine also antagonizes them. However, direct measurements of the persistent TTX-sensitive current (in the presence of an equimolar replacement of calcium chloride with cobalt chloride to block Cav currents) in DMV neurons demonstrated (Goldberg et al. 2012) that acute application of 5 µM isradipine did not antagonize the persistent sodium current in DMV neurons (data not shown).

A functional segregation between the Cav1.3-like and HVA-like Ca\(^{2+}\) currents in a model of a slow autonomous pacemaker

The fact that 5 µM isradipine slows pacemaking while 5 µM nifedpine speeds it up, is probably due to the facts that i) the isradipine is the dihydropyridine with the highest affinity to Cav1.3 channels; and that conversely ii) nifedpine more-or-less completely blocks Cav1.2 channels whereas isradipine does so only partially. Thus, Cav1.3 channels that contribute only a small inward current to DMV motoneurons (Goldberg et al. 2012), are capable of speeding up pacemaking while fluxing little Ca\(^{2+}\) into the cell (Goldberg et al. 2012). In contrast, Cav1.2 and Cav2 channels generate large Ca\(^{2+}\) currents and flux much more Ca\(^{2+}\) into the cell that slows pacemaking by activating AHPs (Goldberg et al. 2012). How can such a small Cav1.3 current robustly speed up pacemaking? We hypothesized that the answer to this question lies in the fact that the membrane voltage of DMV motoneurons during autonomous pacemaking is very depolarized, which we have previously shown to arise from the action of the “leak” NALCN channels (Goldberg et al. 2012). These channels should place the Cav1.3 current in a position
Cooper et al. 2015

to dictate how closely the cell’s steady-state I-V curve approaches its zero current line. The proximity to the zero current line presumably causes the membrane potential to linger for longer just beneath spike threshold and consequently lengthens the interspike intervals (ISIs). Thus, because the DMV motoneurons’ empirical negative conductance region is within a few picoamperes of the zero current point (Goldberg et al. 2012), even a very small Cav1.3 current – also on the order of a few picoamperes – should suffice to dramatically influence the lingering. In contrast, according to this hypothesis the HVA currents, as large as they are, are not activated in this voltage region and therefore cannot influence the pacemaking directly. Instead, they do so via their coupling to K⁺ channels that underlie AHPs, as discussed above.

To test this hypothesis, we constructed a model of a slow pacemaking neuron (Ermentrout 1998) that like the DMV neurons possesses an N-shaped steady-state I-V curve with a negative conductance region. We also shifted the voltage of the trajectories to correspond to the relatively depolarized voltage oscillations of DMV neurons (Goldberg et al. 2012). The autonomous pacemaking of the model was achieved by including in the model two voltage-activated Ca²⁺ currents that shared the same functional shape but differed in that one half-activated at -20 mV (representing the HVA currents, denoted $I_N$) and the other at -40 mV (representing the Cav1.3 current, denoted $I_L$). We also assumed that the maximal conductance of $I_N$ was 25 times larger than the maximal conductance of $I_L$ (Fig. 3). Under these conditions the model cell fired autonomously and displayed Ca²⁺ transients with amplitudes of approximately 75 nM. Abolishing $I_N$ reduced the amplitude to less than 20 nM but did not abolish pacemaking. In contrast, abolishing $I_L$ alone arrested pacemaking and reduced the Ca²⁺ concentration to less than 1 nM (Fig. 3A).

How does pacemaking depend on $I_L$ despite its maximal conductance being 25 times smaller than the maximal conductance of $I_N$? The answer to this question is given by considering the steady-state I-V curve of the model, and the value of the two Ca²⁺ conductances in the voltage region where the I-V curve approaches the zero current line. Despite being larger, the voltage-dependence of $I_N$ prevents it from making a significant contribution in this subthreshold range.
Therefore, abolishing $I_N$ hardly moves the I-V curve any closer to the zero current line (Fig. 3B). In contrast, $I_L$ is strongly activated at this region, and therefore despite being 25 times smaller than $I_N$, abolishing $I_L$ raises the I-V such that it crosses the zero current line. The zero crossing establishes a steady-state voltage, which arrests pacemaking. $I_L$ also controls the degree of lingering in the model. For example, doubling the amplitude of its conductance, pushes the I-V curve further from the zero current line, thereby increasing the inward current near threshold that speeds up the trajectory of the membrane voltage in that region (Fig. 3C).

Voltage dependency of somatic Ca$^{2+}$ currents and dendritic Ca$^{2+}$ concentrations in DMV neurons fit the functional segregation model

In the model, in order for the bulk of Ca$^{2+}$ influx to result from $I_N$, the maximal conductance of $I_N$ needed to be more than an order of magnitude larger than the maximal conductance of $I_L$ (as mentioned above, we used a factor of 25). To quantify Ca$^{2+}$ currents and the rises in Ca$^{2+}$ concentrations that they generated in the model, we measured them in response to a simulated voltage ramp. These simulations were conducted either for $I_L$ alone or for the total current ($I_L+I_N$). Indeed, the currents and associated concentrations were roughly 20 times larger when both currents were activated compared to when only $I_L$ was activated. If such a discrepancy in the sizes of the Cav1.3 and HVA currents was present in real DMV neurons, it should be detectable by conducting the same voltage ramp experiment on actual DMV neurons. We applied somatic voltage ramps to cesium (Cs$^+$)-loaded DMV neurons and measured the total current and the 5 µM isradipine-sensitive subthreshold current as well as the dendritic Ca$^{2+}$ signals associated with them. To accurately determine the isradipine-sensitive subthreshold response required averaging measurements from several cells. These measurements revealed that an isradipine-sensitive somatic Ca$^{2+}$ current is indeed activated in the subthreshold range, which explains why applications of 5 µM isradipine slowed pacemaking (Fig. 2B). Moreover this current and the associated dendritic Ca$^{2+}$ signals were indeed approximately 20 times smaller then the full-blown responses (Fig. 4A), in agreement with the model prediction.
Because the isradipine-sensitive subthreshold current is generated by Cav1.3 channels, these results suggest that DMV neurons express much less Cav1.3 channels in comparison to other HVA Cav channels. We tested this with qRT-PCR analysis that revealed a differential tissue-level expression in the DMV of the various channels, when normalized either to housekeeping genes or to the tubulin marker, TUBB3, that is specific to neurons (P < 0.001, Kruskal-Wallis one-way ANOVA). The transcript levels of Cav1.3 channel were approximately 20-fold less than the sum of the transcript levels of all the other HVA channels (Fig. 4B).

We have previously reported that the dendritic Ca\(^{2+}\) transient measured in current clamp in DMV neurons rose rapidly with each spike and then decayed exponentially (Goldberg et al. 2012). This shape contrasted with that observed in SNc DA neurons that exhibited substantial Ca\(^{2+}\) influx prior to the spike (Guzman et al. 2009). The current model, nevertheless, predicted a very small albeit visible increase in Ca\(^{2+}\) prior to the occurrence of a spike (Fig. 3A, green arrow), despite the size of \(I_L\) relative to \(I_N\). This discrepancy may have stemmed from the distal location of the dendritic spike-triggered changes in fluorescence, which were typically >50 µm from the soma (Fig. 4C), vs. the measurements of Ca\(^{2+}\) currents, which despite being done in Cs\(^-\)-loaded cells, probably represented current flowing through proximally located channels. Similarly the fluorescence measurements conducted during the voltage ramps (Fig. 4A) were taken from proximal dendrites (20–50 µm from the soma). Thus it is possible that relative complement of Cav1.3 is even smaller more distally, thereby reducing the putative subthreshold calcium entry via these channels. Nevertheless, upon closer scrutiny of the experimental data, we found some examples of DMV neurons in which a small subthreshold component was visible in the spike-triggered average of the dendritic Ca\(^{2+}\) oscillations (Fig. 4C).

**A window A-current slows pacemaking in DMV neurons**

The voltage-clamp recordings, Ca\(^{2+}\) imaging and mRNA profiling support our conclusion that Cav1.3 is an order-of-magnitude less abundant than other HVA Ca\(^{2+}\) currents in DMV motoneurons. Using the model we explained how a small Cav1.3-like current can nevertheless significantly modulate the slow pacemaking of the model neuron. However, this modulation relied on the ability of the model...
neuron to exhibit class I Hodgkin excitability (Hodgkin 1948) meaning that it can exhibit very long ISIs (or lingering) near the transition point (bifurcation) of its voltage to oscillations (Ermentrout 1996). We therefore need to identify the biophysical mechanism that confers such behavior in DMV motoneurons. Lingering behavior depends on two biophysical mechanisms. First, the cell must have a steady-state negative conductance region. Our previous work has shown that DMV cells exhibit a negative conductance region due to the action of the persistent Na⁺ current (Goldberg et al. 2012). However, while the negative conductance region is necessary, it may not be sufficient, because the oscillations might become very fast even with a small deviation from the bifurcation point. Thus, the second mechanism that serves to slow pacemaking is the outward fast inactivating (A-type) K⁺ current. This current is activated when the cell depolarizes towards spike threshold and resists the depolarization, thereby causing the cell to slow its ramp up to spike threshold (Connor and Stevens 1971; Ermentrout and Terman 2010).

The influence of the A-type current on the autonomous spike rate and membrane voltage trajectory near spike threshold is evident from biophysical measurements of its voltage activation and inactivation curves. These curves reveal a ‘window current’ in the subthreshold region (centered around -50 mV) just below spike threshold (Fig. 5A) – at a much more depolarized voltage range than was previously reported in the rat (Sah and McLachlan 1992). Whole cell current clamp measurements in DMV neurons strongly suggest that A-type currents indeed temper the autonomous firing rate of DMV motoneurons, because dialysis of the intracellular milieu after rupturing of the membrane always significantly reduces the A-current (P < 0.01, SRT) (Fig. 5B), which is accompanied by a speed-up of autonomous pacemaking (P < 0.01, SRT) (Fig. 5C). To verify that antagonizing A-currents affects the firing rate in an intact cell, we tested the effect of phrixotoxin-2 – a selective antagonist of Kv4 channels that give rise to A-currents – on the discharge rate of DMV neurons recorded in the loose patch mode. Application of nominally 1-2 µM phrixotoxin-2 to the bath (see Methods)(Subramaniam et al. 2014) transiently increased the firing rate of these neuron by 21% (P < 0.05, SRT) (Fig. 5D).
Discussion

Functional segregation between the Cav1.3 and the HVA Ca^{2+} currents

DMV neurons express two kinds of L-type Ca^{2+} channel: Cav1.2 and Cav1.3 (Goldberg et al. 2012), that differ in their voltage dependency with the half-activation voltage of Cav1.3 channels being approximately 20 mV more hyperpolarized (Lipscombe et al. 2004). In the present study we have shown that DMV neurons manipulate this voltage difference to bestow Cav1.3 channels with complementary (if not opposing) role to that of Cav1.2 channels and at least one other HVA channel, the Cav2.2 (N-type) channel. The cells do so by expressing a substantial “leak” NALCN channel that positions their subthreshold membrane trajectories in between the half-activation voltages of the Cav1.3 channel and the HVA Ca^{2+} channels. This segregation causes the majority of the Ca^{2+} influx into the cytosol to enter via the HVA channels, with a negligible contribution from subthreshold flux via Cav1.3 channels. Conversely, the HVA currents can only influence the cells’ autonomous firing rate via their coupling to the K^{+} currents that underlie the AHPs: Cav1.2 channels activate the sAHP and Cav2.2 channels activate SK channels that underlie the mAHP (Goldberg et al. 2012; Sah and McLachlan 1992). The end result is a slowing of the firing rate. In contrast, Cav1.3 channels are able to speed up the firing rate of the cells by controlling the proximity of the cells’ N-shaped steady-state I-V curve to their zero (ionic) current point. In doing so, Cav1.3 channels determine the degree of subthreshold voltage lingering and the duration of the ISI. In our analysis, we used a wide range of ages (3-8 weeks), spanning immature to young adults. This could give rise to a potential confound as we may have overlooked any age-dependence of Ca^{2+} regulation in these cells. However, the 3 biophysical properties that are essential for our argument, namely: i) an N-shaped steady state IV curve; ii) a depolarized subthreshold region due to NALCN channels; and iii) having the majority of Ca^{2+} ions entering during spiking – were similar across all these ages. Hence, although this is a legitimate concern, we do not see any evidence of age-dependent Ca^{2+} regulation.
Our physiological estimate of the relative sizes of the Cav1.3 current vs. the HVA currents shows that the former is over an order of magnitude smaller than the latter. We reached this conclusion by comparing the predictions concerning the Ca\textsuperscript{2+} currents and the Ca\textsuperscript{2+} cytosolic concentration that arose from our simplified one-compartment model to the empirically measured Ca\textsuperscript{2+} currents and Ca\textsuperscript{2+} concentration. Molecular profiling revealed that the transcript level of neuronal Cav1.3 mRNA was an order of magnitude less than the total transcript level of mRNA of the other HVA channels, in close agreement with the physiological and imaging estimates.

**Multiple conductances activated in the subthreshold range guarantee robust pacemaking in DMV neurons**

Cav1.3 channels are not alone in influencing the subthreshold trajectory of pacemaking. Persistent Na\textsuperscript{+} and HCN currents are other depolarizing currents that drive pacemaking in these neurons (Goldberg et al. 2012). Despite the driving to spike threshold by at least three pacemaking currents, DMV neurons fire at a relatively slow rate of approximately 1 spike/s. We have shown that this slowing of the discharge rate is attributable at least in part to the constitutive activity of a Kv4 channels (although we cannot rule out a contribution from Kv1 channels) that give rise to an A-type window current that is conveniently targeted precisely to the subthreshold voltage range. Previous measurements using sharp electrodes in the rat found the A-type window current to reside at a much more hyperpolarized voltage range: even after correcting for junction potential the difference is at least 20 mV (Sah and McLachlan 1992). The difference is attributable to our use of a different recording technique. Whole cell patch electrodes afford better voltage control than do high resistance sharp electrodes.

**Clinical implications**

We have shown the Cav1 channels contribute directly to the Ca\textsuperscript{2+} influx that occurs with each spike. Because Ca\textsuperscript{2+} influx into the weakly buffered DMV neurons generates basal mOS, Cav1 channels must contribute to it. Unlike the case of SNc DA neurons, Cav1.3 channels themselves do not flux much Ca\textsuperscript{2+} into DMV cells. However, by virtue of the ability of Cav1.3 channels to speed up spiking they too could contribute indirectly to Ca\textsuperscript{2+} influx during DMV neuron discharge,
and consequently increase mOs. It is also possible that Ca\(^{2+}\) influx via Cav1.3 channels has direct access to the endoplasmic reticulum and from there to the mitochondria. In such a scenario, it is possible that the Cav1.3 current is particularly efficacious in driving basal mOS despite it small amplitude. Because the clinically approved drug isradipine has equal affinity to both Cav1.2 and Cav1.3 Ca\(^{2+}\) channels (Sinnegger-Brauns et al. 2009), it should antagonize either of these putative mechanisms. As we have shown previously, isradipine is effective at lowering basal mOS in DMV neurons in wild-type as well as in the transgenic DJ-1 knockout mouse model of PD. Our present results suggest that the beneficial effect of isradipine in treating early-stage PD, and particularly the symptoms of dysautonomia (Miller et al. 2009), arises from its equal affinity to Cav1.2 and Cav1.3 Ca\(^{2+}\) channels. Antagonizing these channels reduces Ca\(^{2+}\) influx during spiking via both channels, and prevents Cav1.3 Ca\(^{2+}\) channels from driving pacemaking in DMV motoneurons. Because nifedipine failed to slow down pacemaking in DMV motoneurons, it would seem that dihydropyridines that have a lower affinity to Cav1.3 channels, while still able to reduce spike-related Ca\(^{2+}\) influx, will not have the added benefit of preventing subthreshold Ca\(^{2+}\) influx from exacerbating mOS in neurons known to be vulnerable in PD (Goldberg et al. 2012; Guzman et al. 2010; Sanchez-Padilla et al. 2014).

**Grants**

This work was supported by grants from the Hartman Foundation, the IDP Foundation, the Picower Foundation, the US National Institutes of Health (P50 NS047085 and T32 NS041234) and the Department of Defense (W81XWH-11-1-051) to D. J. Surmeier; by a grant from the Israel Science Foundation (ISF) to H. Soreq; and by a grants from the ISF (No. 154/14) and from the Michael J. Fox Foundation to J. A. Goldberg.

**Disclosures**

No conflicts of interest, financial or otherwise, are declared by the authors.
Figure Captions

Figure 1. Antagonizing Cav1 Ca^{2+} channels reduces the amount of Ca^{2+} influx per spike. A. Bottom: a sequence of 10-s current injections from -60 pA to +60 pA either hyperpolarize and silence discharge or depolarize and increase discharge. 2PLSM fluo-4 imaging of the soma reveals that the level of fluorescence in the silenced cell is relatively insensitive to the degree of hyperpolarization. In contrast, the fluorescence increases linearly as the cell speeds up. Imaging was conducted in the final 5 seconds of each current pulse and black bar indicates the average level of fluorescence. B. Plotting fluorescence levels as a function of the firing rate also reveals a linear relationship. Application of 200 nM calciseptine – a Cav1 channel specific neurotoxin – reduces the slope of the dependence of fluorescence on firing rate indicating that the amount of Ca^{2+} flux per spike is reduced in the presence of this toxin. *p<0.05, SRT.

Figure 2. Acute application of various dihydropyridines affects the rate of pacemaking differentially. A. Application of 5 µM nifedipine, slightly but significantly speeds up pacemaking in DMV neurons. B. Acute application of 5 µM isradipine significantly and reversibly slows pacemaking in DMV neuron (shown on a logarithmic scale). *p<0.05, SRT. In order to better gauge the autonomous firing rate of the DMV neurons, the slices were bathed in ACSF (at an elevated temperature) that included a cocktail of glutamatergic, GABAergic, cholinergic and serotonergic receptor antagonists.

Figure 3. A two Ca^{2+} channel single-compartment model of pacemaking DMV neurons. A. The model neuron discharges rhythmically and each action potential is accompanied by a rapid influx of Ca^{2+} that decays with time (red). The green arrow marks a small yet visible rise in Ca^{2+} just prior to the spike. Elimination of the HVA-like Ca^{2+} current (I_{n}) eliminated the rapid influx and reveals much smaller subthreshold oscillations (blue, note the change in scale). Elimination of the Cav1.3-like Ca^{2+} current (I_{L}) arrests pacemaking and creates a stable resting potential. Ca^{2+} concentration all but vanishes (black). B. Left: Steady-state I-V
curves of the 3 various parameter regimes in panel A, with corresponding color-
coding. Right, top: Zoom-in on the region of the I-V curves nearest to the zero-
current line (dashed black line). The green arrow marks the value of the stable
resting potential. Right, bottom: the corresponding steady-state I-V curves for the
two calcium currents. C. Voltage trajectory for two values of \( I_L \). The larger current
passes further away from the zero-current line and does so faster (as indicated by
the density of the corresponding marks) resulting in a shorter ISI interval.

Figure 4. Ca\(^{2+}\) responses to voltage ramps and the spike-triggered waveform of
Ca\(^{2+}\) in DMV neurons are consistent with the two Ca\(^{2+}\) channel model. A. Left:
Voltage-dependence of \( I_L \) (black) and \( I_L + I_N \) (gray) and the corresponding Ca\(^{2+}\) rises
they induce in response to a slow voltage ramp in the model. Right: Measurement
of the total (gray) and isradipine-sensitive (black) somatic Ca\(^{2+}\) currents and the
corresponding raw dendritic fluorescence from DMV neurons in response to a
slow voltage ramp. Ramp speed was 100 mV/s for the full range (gray trace) and
50 mV/s for the subthreshold range (black one). Black trace is the average of 5
measurements from 3 cells. Note the difference in scale between black and gray
traces both in the model and in the experiment. The internal pipette solution
included 5 mM QX-314 to block Nav channels. B. Neuronal Cav channel mRNA
expression normalized to TUBB3 in the DMV. The expression level is expressed in
percentages of the median value of the expression of Cav1.2 mRNA. C. A spike-
triggered average of Ca\(^{2+}\) oscillations (as in Fig. 6C, different cell) reveals a slight
rise in dendritic Ca\(^{2+}\) concentration (70 µm from the soma) just prior to the spike
marked by black arrow. Compare to red trace in panel Fig. 4A.

Figure 5. A-type Kv4 \( K^+ \) currents slow pacemaking of DMV neurons. A. Left: An
example of measurements of the activation (black) and inactivation (gray) of the
transient \( K^+ \) current in the presence of TTX. Right: The normalized Boltzman
sigmoidal functions describing the voltage dependence of activation and
inactivation of the Kv4 current, and the distribution of the voltages of half
activation (\( n=13 \)) and inactivation (\( n=15 \)), reveal a window Kv4 current that spans
the typical subthreshold range of the voltage trajectory of DMV neurons during
pacemaking. B. Left: Time course of the reduction in the peak A-current as a function of time after membrane rupture in whole cell configuration. Right: Comparison of the peak A-current at membrane rupture vs. past 4 minutes from rupture. C. Left: Time course of the instantaneous firing rate of a DMV neuron in cell-attached mode prior to membrane rupture and after whole-cell configuration was achieved. Right: Comparison of autonomous discharge rate before and after rupturing the membrane. D. Brief application of nominally 1-2 µM phrixotoxin-2 (PaTX-2), a selective Kv4 antagonist, transiently increases the firing rate of DMV neurons recorded in the loose-patch configuration (shown on a logarithmic scale). *p<0.05, **p<0.01, SRT.

References


Cooper et al. Figure 1
Cooper et al. Figure 2

Blockers

+5 μM Isradipine

Wash (Blockers)

AB spikes/s

0.05 0.5 1 1.5

spikes/s

Nifedipine

Blockers

0 1 1.5

Cooper et al. Figure 2
Cooper et al. Figure 3
Cooper et al. Figure 4

A

Model

Experiment

Total

Isradipine sensitive

0 pA

B

Neuronal mRNA expression level

% (normalized to TUBB3)

C

Neuronal mRNA expression level

40 mV

0 mV

ΔF=5

ΔF=2

100 nM

20 μM
Cooper et al. Figure 5

A. 

B. 

C. 

D. 

Activation of A-current at rupture > 4 min.